

## ORIGINAL RESEARCH



# Differential impact of decreasing relative humidity on photosynthesis under fluctuating light between maize and tomato

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## Abstract

Photosynthetic CO<sub>2</sub> assimilation ( $A_N$ ) under fluctuating light (FL) plays a critical role in plant growth and crop yield. C<sub>4</sub> crops usually reach greater yields than C<sub>3</sub> crops when grown in regions with low relative humidity, which might be partly due to the CO<sub>2</sub> concentration mechanism and higher  $A_N$  under FL of C<sub>4</sub> crops. To test this hypothesis, we measured gas exchange and chlorophyll fluorescence under different changing frequencies of FL at contrasting relative air humidity (60% and 15%) for leaves of maize (*Zea mays*, C<sub>4</sub>) and tomato (*Solanum lycopersicum*, C<sub>3</sub>). In general, a more rapid changing frequency of FL would cause a larger loss in  $A_N$  during shade-sun transitions, with 20–38% in maize and 16–52% in tomato. Low relative humidity induced a larger decrease of stomatal conductance ( $g_s$ ) under FL in tomato than in maize, leading to a much stronger loss in  $A_N$  in tomato than in maize. After transition from low to high light,  $A_N$  of maize was mainly limited by  $g_s$  with negligible non-stomatal limitation, and the stomatal limitation was slightly affected by relative humidity. By comparison, low relative humidity significantly increased the stomatal limitation of  $A_N$  in tomato but hardly affected non-stomatal limitation. These results highlight the differential impact of decreasing relative humidity on dynamic photosynthesis and productivity between C<sub>3</sub> and C<sub>4</sub> crops.

## 1 | INTRODUCTION

Photosynthesis is an important remaining opportunity for further improving the genetic yield potential of our major crops. Measurement, analysis, and improvement of leaf CO<sub>2</sub> assimilation ( $A_N$ ) have focused largely on photosynthetic capacity under light-saturated steady-state conditions (Scafaro et al. 2011; Driever et al. 2014; Yamori et al. 2016). However, in modern crop canopies of several leaf layers, light is rarely constant, and most of the leaves experience marked light fluctuations throughout the day, owing to cloudy movement, wind-induced change in leaf angle and shading from upper leaves. As a result, most leaves of field crops are rarely under steady-state photosynthesis and dynamic photosynthesis contributes to a significant fraction of carbon gain (Kromdijk et al. 2016a; Slattery et al. 2018; De Souza et al. 2022). Upon transition from sun to shade

and back into sunlight, photosynthesis needs several minutes to return to steady-state efficiency, leading to approximately 10–40% loss in potential  $A_N$  (Long et al. 2022). Therefore, increasing photosynthetic efficiency under fluctuating light (FL) is a promising strategy for improving plant growth and crop yield (Zhu et al. 2004; Adachi et al. 2019; De Souza et al. 2020; Kimura et al. 2020; Yamori et al. 2020).

Low-light-adapted C<sub>3</sub> and C<sub>4</sub> species showed similar light utilization in FL (Percy and Calkin 1983; Percy et al. 1985; Krall and Percy 1993), but high-light-adapted C<sub>4</sub> plants utilize FL less efficiently than high-light-adapted C<sub>3</sub> plants under low VPD conditions (Kubásek et al. 2013; Li et al. 2021). Therefore, increasing sunfleck utilization efficiency has the potential to improve plant growth and yield of C<sub>4</sub> plants under natural field conditions. Many recent studies investigated the limiting factor of sunfleck utilization efficiency in C<sub>3</sub> crops

(Soleh et al. 2016; Salter et al. 2019; Acevedo-Siaca et al. 2020; De Souza et al. 2020; Li et al. 2020; Sakoda et al. 2022). Unlike immediate CO<sub>2</sub> fixation by Rubisco in the C<sub>3</sub> plant tomato, CO<sub>2</sub> in the C<sub>4</sub> plant maize is first hydrated to HCO<sub>3</sub><sup>−</sup> by carbonic anhydrase and assimilated to oxaloacetate by phosphoenolpyruvate carboxylase in mesophyll cells (Yamori et al. 2014). Oxaloacetate is subsequently reduced to malate in mesophyll cells and then transported to bundle sheath cells, in which CO<sub>2</sub> is regenerated from the decarboxylation of malate and fixed by Rubisco. The CO<sub>2</sub> concentration mechanism facilitates relatively high rate of photosynthesis under low g<sub>s</sub> conditions. However, little is known about the major limiting factor of sunfleck utilization efficiency in C<sub>4</sub> crops (Li et al. 2021; Wang et al. 2021; Lee et al. 2022). Under the background of global climatic change, uneven rainfall has become increasingly frequent, leading to drought stress or a decrease in relative air humidity. C<sub>4</sub> crops usually reach greater yields than C<sub>3</sub> crops when grown in regions with low precipitation. Many previous studies investigated the effects of drought and dry air on steady-state photosynthesis in C<sub>3</sub> and C<sub>4</sub> plants (Leakey et al. 2006; Ibrahim et al. 2008; Carmo-Silva et al. 2010; Vilagrosa et al. 2012; Zivcak et al. 2013; Sancho-Knapik et al. 2022), but it is unclear whether C<sub>4</sub> crops have higher sunfleck utilization efficiency in dry air than C<sub>3</sub> crops.

Fluctuating light can induce a gradual decrease of stomatal conductance (g<sub>s</sub>) owing to the low-light-induced stomatal closure (Kimura et al. 2020; Yamori et al. 2020; Eyland et al. 2021). Upon transition from low to high light, the induction rapidity of g<sub>s</sub> is much slower than the activation rate of Rubisco (Sakoda et al. 2021). The absolute g<sub>s</sub> at low light and the induction speed of g<sub>s</sub> are critical for the induction kinetics of A<sub>N</sub> (Kaiser et al. 2020; Kimura et al. 2020; Eyland et al. 2021). Therefore, g<sub>s</sub> imposes a significant limitation on A<sub>N</sub> under FL in some C<sub>3</sub> plants (Kaiser et al. 2020; Kimura et al. 2020; Eyland et al. 2021; Liu et al. 2022). Under high VPD conditions, g<sub>s</sub> usually declines to prevent hydraulic failure, leading to subsequent detrimental effects on steady-state A<sub>N</sub>, growth and productivity (Shirke and Pathre 2004; Zhang et al. 2005; Peguero-Pina et al. 2016; Qiu et al. 2017; Merilo et al. 2018; Grossiord et al. 2020; Inoue et al. 2021; Sancho-Knapik et al. 2022). Furthermore, a high VPD slows down the induction rate of g<sub>s</sub> in tomato (Kaiser et al. 2017, 2020). Owing to the similar effects of VPD and FL on g<sub>s</sub>, dry air might aggravate the decrease of g<sub>s</sub> under FL. However, little is known about the impact of high VPD on g<sub>s</sub> behavior under FL in C<sub>3</sub> and C<sub>4</sub> plants.

In nature, FL occurs with different frequencies (Valladares et al. 1997; Kimura et al. 2020). Based on the characteristics of g<sub>s</sub> under FL (i.e., decrease of g<sub>s</sub> at low light and increase of g<sub>s</sub> at high light), the duration of low light significantly affects the absolute g<sub>s</sub> before transitioning to high light (Yamori et al. 2020; Eyland et al. 2021). Therefore, the changing frequency of FL can significantly influence the photosynthetic efficiency during shade-sun transitions (Arce Cubas et al. 2023). The effects of changing the frequency of FL on g<sub>s</sub> and CO<sub>2</sub> assimilation rate have been well clarified in the C<sub>3</sub> model plant *Arabidopsis thaliana* (Kimura et al. 2020; Yamori et al. 2020). Furthermore, the changing

frequency of FL influences CO<sub>2</sub> assimilation rate during shade-sun transitions in C<sub>4</sub> species, and C<sub>4</sub> species had stronger and more sustained CO<sub>2</sub> assimilation rate during the low-light phase in FL compared with C<sub>3</sub> species (Arce Cubas et al. 2023). However, it is unclear whether relative humidity interacts with the changing frequency of FL to influence photosynthetic efficiency under FL.

Based on current knowledge about dynamic photosynthesis and its responses to environmental stresses, dry air theoretically aggravates the loss in photosynthetic carbon gain under FL and thus reduces crop production. Therefore, understanding the responses of dynamic photosynthesis to dry air in C<sub>3</sub> and C<sub>4</sub> plants can help us actively solve the global food crisis under future climatic change. In this study, we measured gas exchange and chlorophyll fluorescence under different frequencies of fluctuating light at contrasting relative air humidity for leaves of maize (*Zea mays*, C<sub>4</sub>) and tomato (*Solanum lycopersicum*, C<sub>3</sub>). The main aims are: 1) to assess whether maize has a higher sunfleck utilization efficiency than tomato in dry air; and 2) to evaluate whether dry air imposes different effects on maize and tomato in terms of g<sub>s</sub> and A<sub>N</sub> under FL.

## 2 | MATERIALS AND METHODS

### 2.1 | Plant materials and growth conditions

Maize (*Zea mays* L. cv. Zhengdan 958) and tomato (*Solanum lycopersicum* Miller cv. Hupishizi) plants were used in this study. Plants were grown in 13-litre plastic pots containing humus soil, with an initial soil N content of 2.1 mg g<sup>−1</sup>. After seed germination, pots were placed in an open field at the Kunming Institute of Botany, Chinese Academy of Sciences, China (102°44′31″E, 25°08′24″N, 1950 m of elevation). During the growth period (from July to August 2022), the day/night air temperatures were approximately 30°C/20°C and the maximum light intensity to which the leaves were exposed was approximately 2000 μmol photons m<sup>−2</sup> s<sup>−1</sup> (measured by LI-1400, Li-Cor Biosciences). These plants were fertilized with 0.15 g N plant<sup>−1</sup> every two days using Peters Professional's water solution (N:P:K = 15:4.8:24.1) to avoid any nutrient stress. To prevent any water stress, plants were watered every day with a gradually increasing amount of water to match the plant growth. After normal growth for a month, mature canopy leaves on the top of plants were used for photosynthetic measurements.

### 2.2 | Gas exchange and chlorophyll fluorescence measurements

The gas exchange and chlorophyll fluorescence were measured simultaneously using a portable photosynthesis system (LI-6400XT; Li-Cor Biosciences) equipped with a leaf chamber fluorometer (Li-Cor Part No. 6400-40, enclosed leaf area: 2 cm<sup>2</sup>). This leaf chamber can be full of all individual maize and tomato leaves. The atmospheric CO<sub>2</sub> concentration (400 μmol mol<sup>−1</sup>), the flow rate of air through the

measuring system ( $300 \text{ mmol min}^{-1}$ ), and leaf temperature ( $30^\circ\text{C}$ ) were controlled by an automatic control device in the LI-6400XT. Irradiance was provided by a mixture of red (90%) and blue (10%) LEDs in the fluorometer. Relative air humidity of 60% or 15% was controlled by using a buffer bottle (water was injected in it) and desiccant. Leaves were first illuminated at  $1500 \text{ } \mu\text{mol photons m}^{-2} \text{ s}^{-1}$  and  $400 \text{ } \mu\text{mol mol}^{-1} \text{ CO}_2$  concentration for at least 30 min to reach

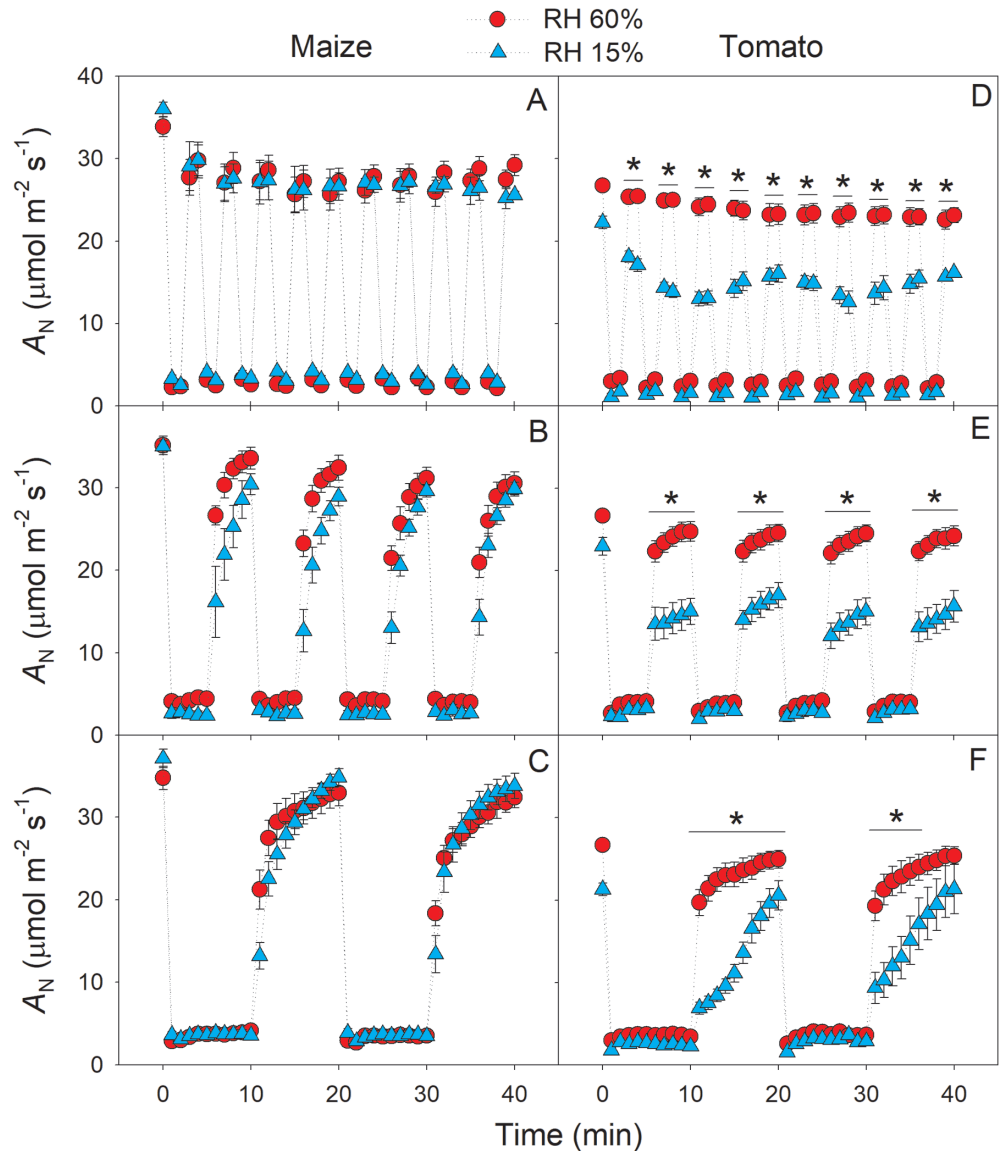
steady-state photosynthesis and stomatal conductance. Afterwards, light intensity was altered between low and high ( $100$  and  $1500 \text{ } \mu\text{mol photons m}^{-2} \text{ s}^{-1}$ ) every 2, 5, or 10 min for 40 min to simulate different frequencies of FL. Photosynthetic parameters were logged every 1 minute to calculate the kinetics of photosynthesis under FL. After transition from low to high light, the measured cumulative  $\text{CO}_2$  assimilation ( $\text{CCA}_{\text{measured}}$ ) in high-light phase was calculated by integrating

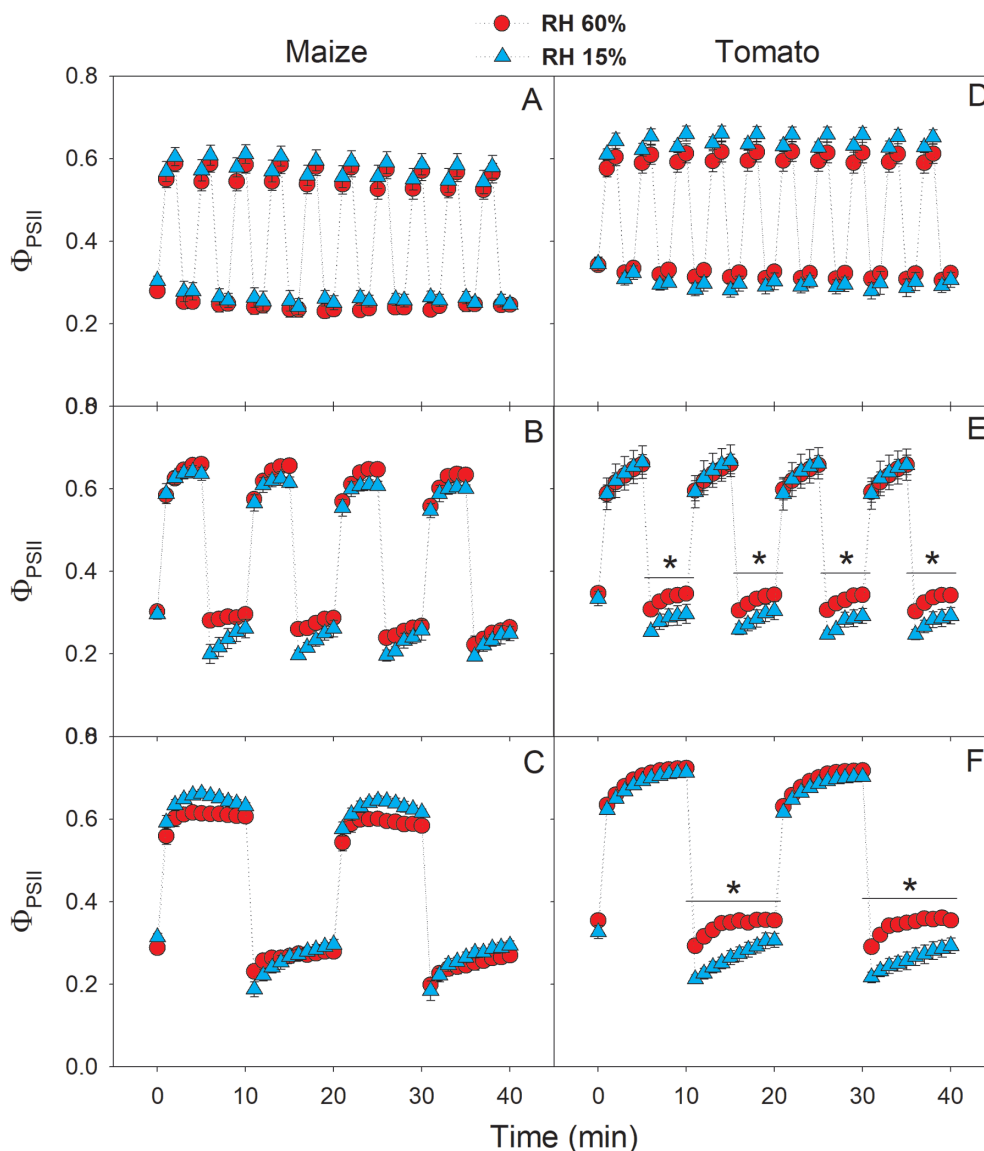
**TABLE 1** Steady state photosynthetic parameters of maize and tomato measured at different relative humidity (60% and 15%) conditions. Data are shown as mean  $\pm$  SE ( $n = 5-6$ ). Different letters indicate significant differences among these treatments (Tukey's test,  $P < 0.05$ )

| Species and conditions | $A_N$ ( $\mu\text{mol m}^{-2} \text{ s}^{-1}$ ) | $\Phi_{\text{PSII}}$ | $g_s$ ( $\text{mol m}^{-2} \text{ s}^{-1}$ ) | $T_r$ ( $\text{mmol m}^{-2} \text{ s}^{-1}$ ) |
|------------------------|-------------------------------------------------|----------------------|----------------------------------------------|-----------------------------------------------|
| Maize RH 60%           | $35.4 \pm 1.3^a$                                | $0.30 \pm 0.01^b$    | $0.22 \pm 0.01^c$                            | $4.4 \pm 0.3^c$                               |
| Maize RH 15%           | $34.9 \pm 0.7^a$                                | $0.30 \pm 0.01^b$    | $0.23 \pm 0.01^c$                            | $9.7 \pm 0.5^b$                               |
| Tomato RH 60%          | $26.8 \pm 0.5^b$                                | $0.35 \pm 0.01^a$    | $0.51 \pm 0.02^a$                            | $8.5 \pm 0.5^b$                               |
| Tomato RH 15%          | $22.9 \pm 1.1^c$                                | $0.33 \pm 0.02^b$    | $0.34 \pm 0.04^b$                            | $12.6 \pm 1.2^a$                              |

$A_N$ , net  $\text{CO}_2$  assimilation rate;  $g_s$ , stomatal conductance;  $\Phi_{\text{PSII}}$ , effective quantum yield of photosystem II photochemistry;  $T_r$ , transpiration rate.

**FIGURE 1** Dynamic change of net  $\text{CO}_2$  assimilation rate ( $A_N$ ) under fluctuating light for leaves of maize (A, B, C) and tomato (D, E, F). Fluctuating light alternates between  $1500$  and  $100 \text{ } \mu\text{mol m}^{-2} \text{ s}^{-1}$  every 2 (A, D), 5 (B, E), or 10 min (C, F) at  $30^\circ\text{C}$  at different relative humidity (RH 60% and 15%). Asterisk indicates a significant difference between 60% and 15% relative humidity (T-test,  $P < 0.05$ ). Data are means  $\pm$  SE ( $n = 5-6$  replicates, each being an individual plant).





**FIGURE 2** Dynamic change of effective quantum yield of photosystem II photochemistry ( $\Phi_{PSII}$ ) under fluctuating light for leaves of maize (A, B, C) and tomato (D, E, F). Fluctuating light alternates between 1500 and 100  $\mu\text{mol m}^{-2} \text{s}^{-1}$  every 2 (A, D), 5 (B, E), or 10 min (C, F) at 30°C at different relative humidity (RH 60% and 15%). Asterisk indicates a significant difference between 60% and 15% relative humidity (T-test,  $P < 0.05$ ). Data are means  $\pm$  SE ( $n = 5-6$ ).

measured  $A_N$ . The maximum potential cumulative  $\text{CO}_2$  assimilation ( $\text{CCA}_{\text{maximum}}$ ) in high-light phase was calculated by integrating the maximum  $A_N$ . The maximum  $A_N$  indicates the steady-state  $A_N$  at 1500  $\mu\text{mol photons m}^{-2} \text{s}^{-1}$  and 400  $\mu\text{mol mol}^{-1} \text{CO}_2$  concentration. The percentage of loss in  $\text{CO}_2$  fixation after transition from low to high light was calculated by  $1 - \text{CCA}_{\text{measured}}/\text{CCA}_{\text{maximum}}$  (Long et al. 2022).

We determined chlorophyll fluorescence parameters using the multi-phase flash (MPF) protocol following recommended procedures (Loriaux et al. 2013). The measuring light intensity and the maximum flash intensity were 1 and 8000  $\mu\text{mol m}^{-2} \text{s}^{-1}$ , respectively. The flash intensity decreased by 60% during the second phase of the MPF and the durations of the three flash phases were 0.3 s, 0.7 s, and 0.4 s, respectively. The effective photochemistry quantum yield of photosystem II ( $\Phi_{PSII}$ ) and total electron transport rate through PSII (ETR) were calculated using the following equation (Genty et al. 1989; Krall and Edwards 1992):

$$\Phi_{PSII} = \frac{(F_m' - F_s)}{F_m'}$$

### 2.3 | Stomatal and nonstomatal limitations

Stomatal limitation ( $L_S$ ) and non-stomatal limitation ( $L_{NS}$ ) imposed on  $\text{CO}_2$  assimilation were calculated using the following equations (Farquhar and Sharkey 1982; Bellasio et al. 2018, 2023):

$$L_S = \frac{A_{\text{potCa}} - A_{\text{potCiop}}}{A_{\text{potCa}}}$$

$$L_{NS} = \frac{A_{\text{potCiop}} - A_{\text{op}}}{A_{\text{potCa}}}$$

where  $A_{\text{potCa}}$  is the potential A that would occur when  $C_i$  was equal to ambient  $[\text{CO}_2]$  at the leaf surface ( $C_a$ ) if there was no epidermal impediment to  $\text{CO}_2$  diffusion into the leaf, as predicted by the  $A/C_i$  curve;

$A_{\text{potC}_{\text{iop}}}$  is the potential  $A$  that would occur at the operational  $C_i$ , as predicted by the  $A/C_i$  curve;  $A_{\text{op}}$  is the operational  $A$  that is measured at the operational  $C_i$ . The response of  $A$  to  $C_i$  was measured at  $1500 \mu\text{mol photons m}^{-2} \text{s}^{-1}$  and different  $C_i$  conditions, and the  $A/C_i$  curve was modelled empirically as a nonrectangular hyperbola (Lee et al. 2022).

## 2.4 | Statistical analysis

Experimental design was a randomized complete block design with at least five replications from different individual plants. Tukey's test was used to determine whether significant differences ( $P < 0.05$ ) existed among all treatments in maize and tomato. A t-test was used to determine whether significant differences ( $P < 0.05$ ) existed between high and low relative humidity in maize or tomato. The software SigmaPlot 10.0 was used for graphing and fitting.

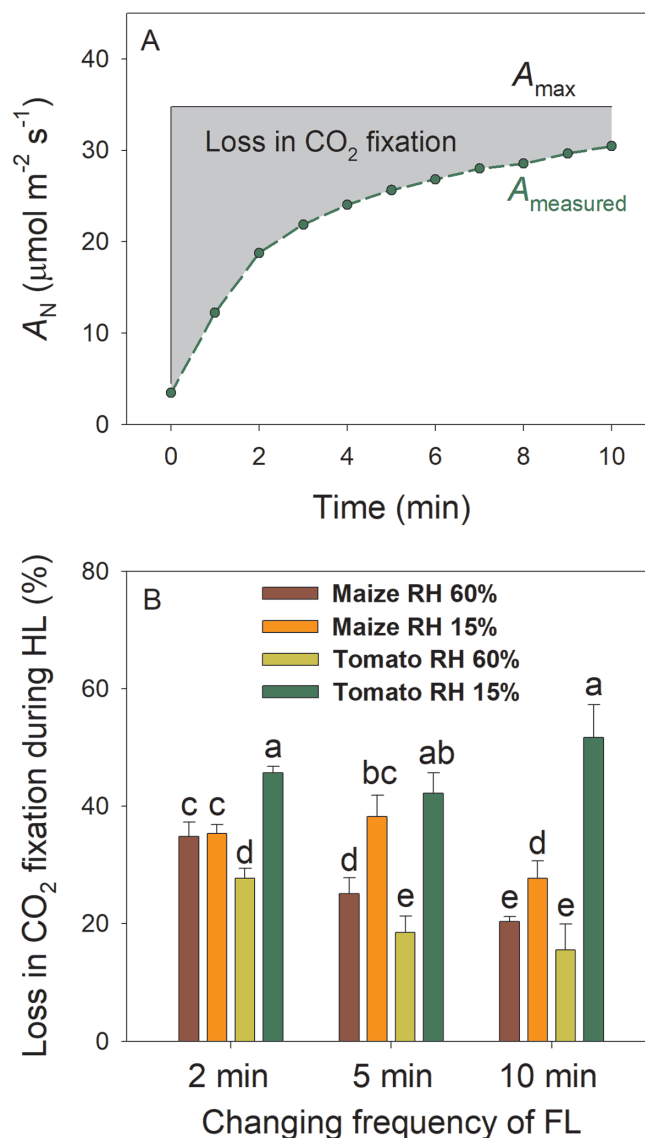
## 3 | RESULTS

### 3.1 | Effects of decreasing relative humidity on steady-state photosynthetic parameters

We first measured the steady-state photosynthetic performance at  $1500 \mu\text{mol photons m}^{-2} \text{s}^{-1}$  and contrasting relative humidity (Table 1). At 60% relative humidity, net  $\text{CO}_2$  assimilation rate ( $A_N$ ) in maize was  $35.4 \pm 1.3 \mu\text{mol CO}_2 \text{ m}^{-2} \text{s}^{-1}$ , which was significantly higher than that in tomato ( $26.8 \pm 0.5 \mu\text{mol CO}_2 \text{ m}^{-2} \text{s}^{-1}$ ) ( $P < 0.0001$ ). At 15% relative humidity,  $A_N$  in maize was  $34.9 \pm 0.7 \mu\text{mol CO}_2 \text{ m}^{-2} \text{s}^{-1}$  but significantly decreased to  $22.9 \pm 1.1 \mu\text{mol CO}_2 \text{ m}^{-2} \text{s}^{-1}$  in tomato ( $P < 0.0001$ ). The effective quantum yield of photosystem II photochemistry ( $\Phi_{\text{PSII}}$ ) at 60% relative humidity was  $0.30 \pm 0.01$  in maize and  $0.35 \pm 0.01$  in tomato. When relative humidity decreased to 15%, values for  $\Phi_{\text{PSII}}$  were  $0.30 \pm 0.01$  and  $0.33 \pm 0.03$  in maize and tomato, respectively. When relative humidity decreased from 60% to 15%, the steady-state  $g_s$  was maintained stable at  $0.22$ – $0.23 \text{ mol m}^{-2} \text{s}^{-1}$  in maize, but decreased significantly from  $0.51 \pm 0.02$  to  $0.34 \pm 0.04 \text{ mol m}^{-2} \text{s}^{-1}$  in tomato ( $P < 0.0001$ ). Concomitantly, transpiration rate ( $T_r$ ) significantly increased from  $4.4 \pm 0.3$  to  $9.7 \pm 0.5 \text{ mmol m}^{-2} \text{s}^{-1}$  in maize and from  $8.5 \pm 0.5$  to  $12.6 \pm 1.2 \text{ mmol m}^{-2} \text{s}^{-1}$  in tomato ( $P < 0.0001$ ).

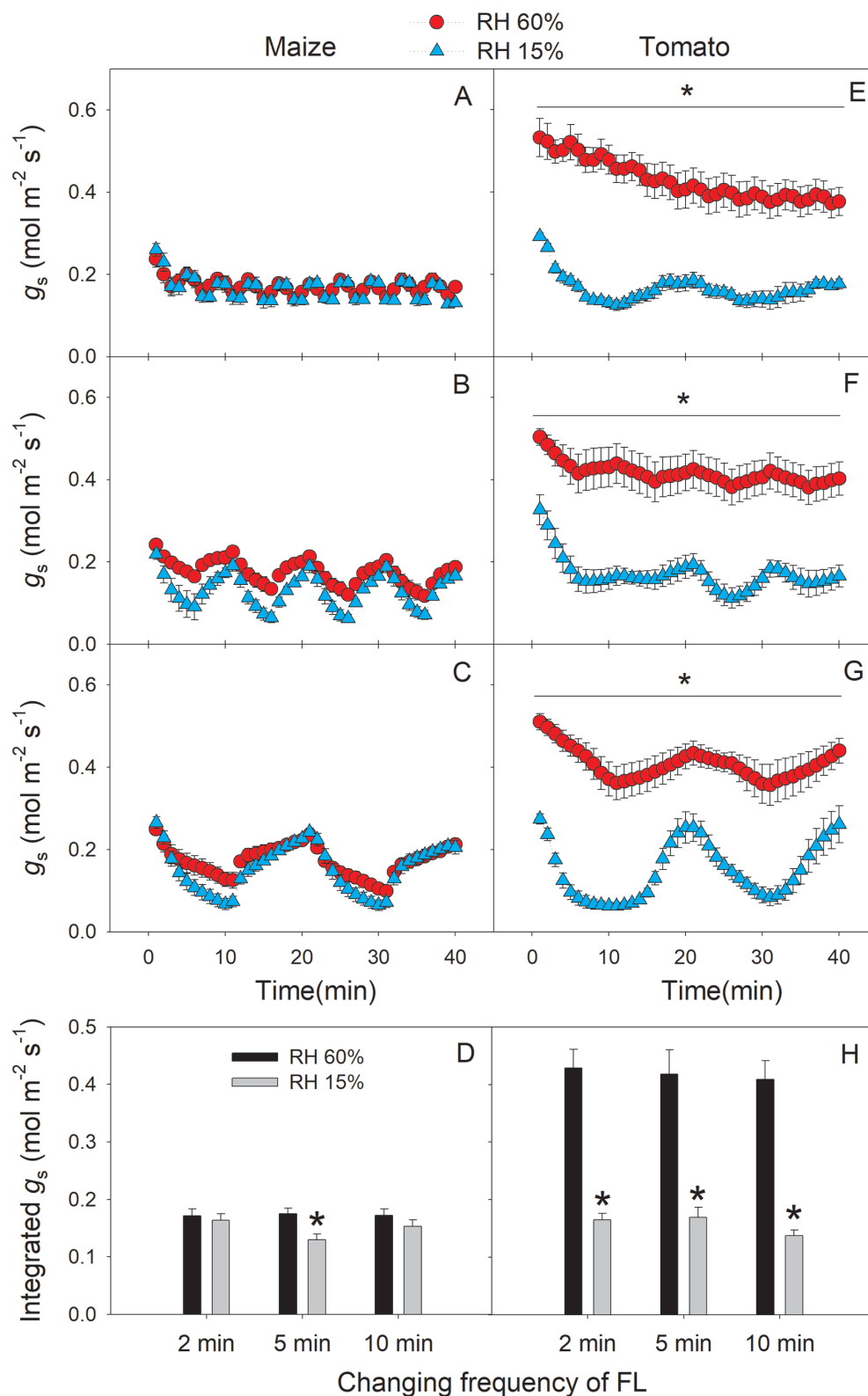
### 3.2 | Effects of dry air on temporal kinetics of photosynthetic parameters under FL

Next, we examined photosynthetic performance under FL conditions alternating between  $100$  and  $1500 \mu\text{mol photons m}^{-2} \text{s}^{-1}$  every 2, 5, or 10 min (FL-2 min, FL-5 min, or FL-10 min). When relative humidity decreased from 60% to 15%,  $A_N$  in low light phases was almost maintained stable under all FL conditions in maize and tomato (Figure 1A–C). Under FL-2 min condition, sunfleck utilization efficiency of maize was hardly affected by the decrease in relative



**FIGURE 3** The loss in  $\text{CO}_2$  fixation after transition from low to high light. (A) A typical model for assessing the loss in  $\text{CO}_2$  fixation under fluctuating light. Solid and dashed lines represent the maximum  $A_N$  and measured  $A_N$ , respectively. (B) The percentage of loss in  $\text{CO}_2$  fixation during high-light phases of fluctuating light for maize and tomato measured at 60% and 15% relative humidity. Different letters indicate significant differences among these treatments (Tukey's test,  $P < 0.05$ ). Data are means  $\pm$  SE ( $n = 5$ – $6$ ).

humidity (Figure 1A). Under FL-5 min and FL-10 min conditions, sunfleck utilization efficiency of maize was slightly suppressed within the first minutes by the low relative humidity (Figure 1B,C). After transfer from low to high light for ten minutes,  $A_N$  reached at least 90% of the maximum value in maize at high and low relative humidity. By comparison, low relative humidity largely significantly suppressed sunfleck utilization efficiency in tomato under all FL conditions, especially within the first minutes after transition from low to high light (Figure 1D–F). After transitioning to high light for one minute under FL-5 min and FL-10 min conditions, values for  $A_N$  were higher than  $24 \mu\text{mol CO}_2 \text{ m}^{-2} \text{s}^{-1}$  at 60% relative humidity but were lower



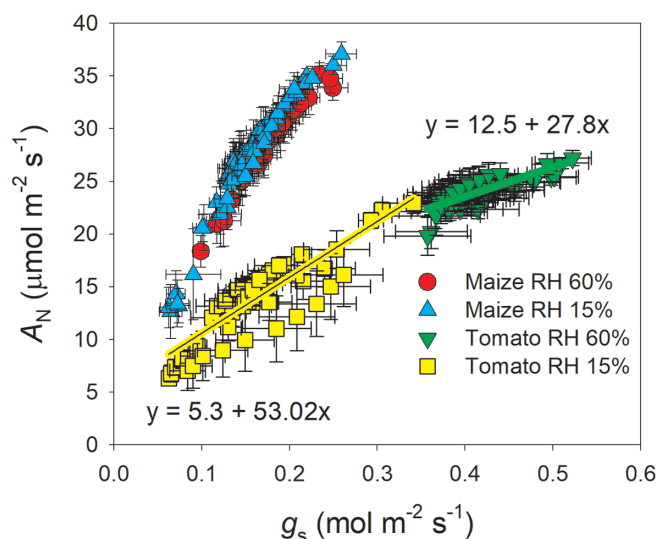
**FIGURE 4** Dynamic change of stomatal conductance ( $g_s$ ) under fluctuating light for leaves of maize (A, B, C) and tomato (E, F, G). Fluctuating light alternates between 1500 and 100  $\mu\text{mol m}^{-2} \text{s}^{-1}$  every 2 (A, E), 5 (B, F), or 10 min (C, G) at 30°C and different relative humidity (RH 60% and 15%). Integrated  $g_s$  values in maize (D) and tomato (H) were calculated. Asterisk indicates a significant difference between 60% and 15% relative humidity (t-test,  $P < 0.05$ ). Data are means  $\pm$  SE ( $n = 5-6$ ).

than 14  $\mu\text{mol CO}_2 \text{m}^{-2} \text{s}^{-1}$  at 15% relative humidity (Figure 1E,F). Consistent with the performance of  $A_N$ , dry air hardly affected  $\Phi_{\text{PSII}}$  in maize leaves under all FL conditions (Figure 2A-C). By comparison,  $\Phi_{\text{PSII}}$  during high-light phases slightly decreased in tomato leaves when exposed to FL-5 min and FL-10 min conditions (Figure 2E-F).

After transitioning to high light for five minutes under FL-10 min condition, values for  $\Phi_{\text{PSII}}$  were approximately 0.35 and 0.26 at 60% and 15% relative humidity, respectively.

We further estimated the loss in  $\text{CO}_2$  fixation by calculating the difference between potential  $\text{CO}_2$  fixation and measured  $\text{CO}_2$  fixation

(Figure 3A). At 60% and 15% relative humidity, cumulative CO<sub>2</sub> fixation of maize lost  $34.9\% \pm 0.025$  and  $35.4\% \pm 0.015$  in FL-2 min,  $25.1\% \pm 0.027$  and  $38.3\% \pm 0.036$  in FL-5 min,  $20.4\% \pm 0.008$  and

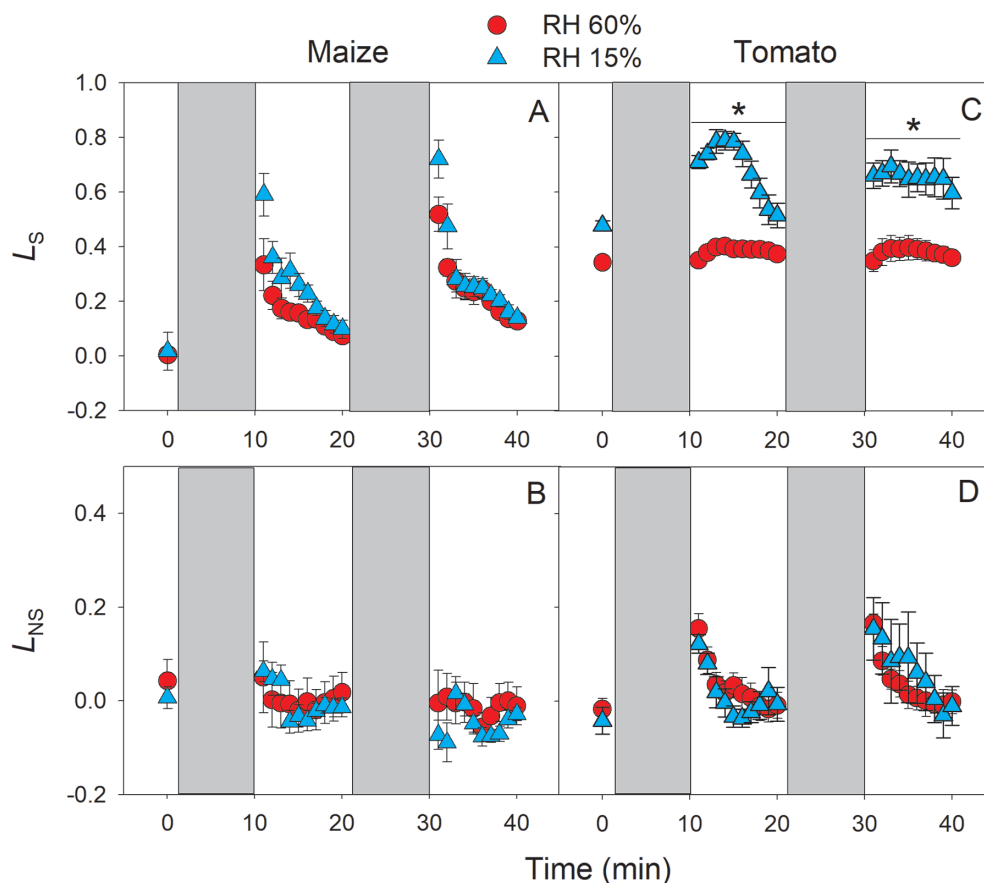


**FIGURE 5** Relationship between  $g_s$  and  $A_N$  during high-light phases of all fluctuating light conditions for maize and tomato measured at 60% and 15% relative humidity. Data are means  $\pm$  SE ( $n = 5-6$ ).

$27.7\% \pm 0.03$  in FL-10 min, respectively (Figure 3B). In tomato leaves, cumulative CO<sub>2</sub> fixation at 60% and 15% relative humidity lost  $27.8\% \pm 0.017$  and  $45.7 \pm 0.011$  in FL-2 min,  $18.5\% \pm 0.028$  and  $42.2\% \pm 0.035$  in FL-5 min,  $15.6\% \pm 0.044$  and  $51.7\% \pm 0.056$  in FL-10 min, respectively (Figure 3B). These results indicated that sunfleck utilization efficiency at 60% relative humidity was significantly lower in FL-2 min condition than in FL-5 min and FL-10 min ( $P < 0.05$ ). Furthermore, dry air induced a larger loss in cumulative CO<sub>2</sub> fixation under FL conditions in tomato than in maize.

Upon transition from high to low light, the decreased  $g_s$  rate in maize was aggravated by dry air, leading to a lower  $g_s$  before transition to high light in FL-5 min and FL-10 min (Figure 4B,C). When back into high light in FL-5 min and FL-10 min,  $g_s$  rapidly increased and the peak of  $g_s$  was hardly affected by dry air (Figure 4B,C). Compared with  $g_s$  at 60% relative humidity, the integrated  $g_s$  of maize leaves at 15% relative humidity decreased by 4%, 26%, and 11% in FL-2 min, FL-5 min and FL-10 min, respectively (Figure 3D). In tomato leaves,  $g_s$  under all FL conditions largely decreased at 15% relative humidity when compared with 60% relative humidity (Figure 4E-G). After transition from high to low light,  $g_s$  rapidly decreased at 15% relative humidity. Furthermore, within the first five minutes after transitioning to high light,  $g_s$  increased very slightly at 15% relative humidity. As a result, when relative humidity decreased from 60% to 15%, the integrated  $g_s$  of tomato leaves decreased by 62%, 60%, and 66% in FL-2 min,

**FIGURE 6** Dynamic changes of stomatal and non-stomatal limitations ( $L_s$  and  $L_{NS}$ ) for leaves of maize (A, B) and tomato (C, D) in high-light phases under fluctuating light alternating between 1500 and  $100 \mu\text{mol m}^{-2} \text{s}^{-1}$  every 10 min at 30°C and different relative humidity (RH 60% and 15%). Grey bars indicate low-light phases and open bars indicate high-light phases. Asterisk indicates a significant difference between 60% and 15% relative humidity (t-test,  $P < 0.05$ ). Data are means  $\pm$  SE ( $n = 5-6$ ).



FL–5 min and FL–10 min, respectively (Figure 4H). Therefore, dry air induced a larger decline of  $g_s$  under all FL conditions in tomato than in maize.

### 3.3 | Photosynthetic limitations under FL

Based on the concurrent changes of  $g_s$  and  $A_N$  under FL, the relationship between  $g_s$  and  $A_N$  was analyzed (Figure 5). During high-light phases under FL,  $g_s$  was positively correlated to  $A_N$  in both species, with a steeper slope in maize. In maize leaves, the slope of relationship between  $g_s$  and  $A_N$  was not altered by relative humidity. By comparison, in tomato, a steeper slope of relationship at 15% relative humidity than that at 60% relative humidity was found. To further examine the impact of  $g_s$  kinetics on  $A_N$ , we analyzed stomatal and non-stomatal limitations ( $L_s$  and  $L_{NS}$ , respectively) during high-light phases. In maize leaves,  $A_N$  during shade-sun transitions was significantly limited by  $g_s$  with little non-stomatal limitation (Figure 6A,B). Within the first two minutes after transition from low to high light, maize leaves showed high levels of  $L_s$  and such temporary  $L_s$  was aggravated by dry air. In tomato leaves,  $L_s$  and  $L_{NS}$  significantly occurred during shade-sun transitions (Figure 6C,D). Furthermore, dry air significantly increased  $L_s$  but hardly affected  $L_{NS}$  during the whole high-light phases in tomato leaves (Figure 6C,D). Therefore,  $L_s$  is more sensitive to dry air in  $C_3$  tomato leaves than in  $C_4$  maize leaves.

## 4 | DISCUSSION

### 4.1 | Sunfleck utilization efficiency is affected by the changing frequency of FL

Increasing the ability of plants to adjust their photosynthetic efficiency under FL is an attractive strategy for improving photosynthesis and crop yield (Kromdijk et al. 2016b; Tanaka et al. 2019; Acevedo-Siaca et al. 2020). Here, we determined the photosynthetic performance under FL with different changing frequencies in maize and tomato (Figure 1). The results indicated that the loss in potential photosynthetic carbon gain during sunfleck varied from 20–38% in maize and 16–52% in tomato (Figure 3B). The highest loss in potential photosynthetic carbon gain at 15% relative humidity in tomato (52%) is stronger than the loss in  $CO_2$  fixation under FL as calculated in previous studies (10–40%) (Lee et al. 2022; Long et al. 2022). Furthermore, we examined the loss in photosynthetic carbon gain under FL conditions with different changing frequencies. The results indicated that FL alternating between low and high light every 2 min (FL–2 min) induced a stronger loss in photosynthetic carbon gain than FL alternating between low and high light every 10 min (FL–10 min) (Figure 3B). These results indicated that a quicker changing frequency of FL would cause a lower sunfleck utilization efficiency. Therefore, the changing frequency of FL is an important factor affecting sunfleck utilization efficiency in both maize and tomato. Consistently, a recent

study found that photosynthetic efficiency under fluctuating light could be influenced by the fluctuation regime (Arce Cubas et al. 2023).

By transferring plants from low to high light under favorable conditions, photosynthetic induction was reported to be delayed in  $C_4$  plants when compared with  $C_3$  plants (Li et al. 2021; Lee et al. 2022). Consistently, the loss in photosynthetic carbon gain was significantly higher in maize than in tomato when exposed to FL–2 min and FL–5 min at 60% relative humidity (Figure 3B). These results strongly supported that the loss in photosynthetic carbon gain in maize mainly occurred in the first 2 min after transition from low to high light (Lee et al. 2022). However, under condition of FL–10 min at 60% relative humidity, no significant difference in the loss in photosynthetic carbon gain was observed between maize and tomato (Figure 3B), which was consistent with the scheme that photosynthetic efficiency during high-light phase is attributable to species rather than photosynthetic pathway (Arce Cubas et al. 2023). Therefore, the previous conclusion that  $C_4$  plants utilize FL less efficiently than  $C_3$  plants should be revised according to the changing frequency of FL.

### 4.2 | Photosynthesis under FL in maize is mainly limited by $g_s$

In previous studies, the difference in sunfleck utilization efficiency between  $C_4$  and  $C_3$  plants was hypothesized to be caused by the slower diffusion of metabolites from mesophyll cells to bundle sheath cells (Kubásek et al. 2013; Lee et al. 2022). However, our present data indicated that such delayed photosynthetic induction in maize was tightly correlated to the response of  $g_s$  to light conditions (Figure 5). After transition from high to low light at 60% relative humidity,  $g_s$  decreased much faster in maize than in tomato, leading to a much lower initial  $g_s$  under low light in maize (Figure 4), which strongly restricted  $A_N$  within the first minutes after transition to high light (Figure 6). Therefore, increasing the  $g_s$  under low light conditions or accelerating the rate of stomatal opening might provide a new opportunity to improve sunfleck utilization efficiency in maize grown under favorite conditions. Increased  $g_s$  or improved stomatal opening has been documented to be efficient to enhance photosynthetic rate and biomass under FL for  $C_3$  plants such as *Arabidopsis thaliana*, rice and tomato (Kaiser et al. 2020; Kimura et al. 2020; Sakoda et al. 2020; Yamori et al. 2020). Because of the stronger relationship between  $g_s$  and  $A_N$  during shade-sun transitions in maize than in tomato (Figure 5), manipulating  $g_s$  would improve dynamic photosynthesis to a greater extent in  $C_4$  crops than in  $C_3$  crops when cultivated in regions with high precipitation.

### 4.3 | Maize uses sunfleck more efficiently than tomato in dry air

In nature, FL often occurs with low relative humidity under the background of global climatic change, especially in semi-arid areas and in

dry season. However, little is known about how dry air affects photosynthetic efficiency under FL (Kaiser et al. 2015). We found that the absolute cumulative CO<sub>2</sub> fixation was much higher in maize than tomato (Figure 1). Although dry air generally accelerated the loss in photosynthetic carbon gain under FL in both maize and tomato, the loss extent was significantly lower in maize (Figure 3B). These results strongly indicate that maize utilizes sunfleck more efficiently than tomato in dry air, providing insight into why C<sub>4</sub> crops have higher yield potential than C<sub>3</sub> crops when cultivated in regions with dry air. In the context of adaptive evolution, these important photosynthetic characteristics of C<sub>4</sub> species result in them being more competitive against C<sub>3</sub> species in regions with dry air.

When relative humidity decreased from 60% to 15%, the closing rate of stomata under low light was accelerated in maize (Figure 4), leading to a lower initial  $g_s$  and thus restricting sunfleck utilization efficiency within the first minutes (Figure 6). Compared with maize, dry air not only induced a more rapid stomatal closure in low light phases but also inhibited stomatal opening during light induction in tomato (Figure 4), consistent with the impact of VPD on  $g_s$  in other C<sub>3</sub> plants (Grossiord et al. 2020). This large decrease of  $g_s$  in dry air made  $g_s$  the major limitation of photosynthesis during sunfleck in tomato (Figure 6), which was consistent with the significant limitation of  $g_s$  during light induction in C<sub>3</sub> plants (De Souza et al. 2020; Liu et al. 2022). Owing to the CO<sub>2</sub> concentration mechanism in maize, a relatively low  $g_s$  could be accompanied by a relatively high A<sub>N</sub> (Wang et al. 2021; Lee et al. 2022). Therefore, although maize and tomato leaves showed similar  $g_s$  under FL conditions in dry air (Figure 4), the absolute A<sub>N</sub> during high-light phases was much higher in maize than in tomato (Figures 1 and 5). Therefore, the maintenance of  $g_s$  and CO<sub>2</sub> concentration mechanism jointly facilitates the higher sunfleck utilization efficiency in maize than in tomato when exposed to FL conditions in dry air.

## 5 | CONCLUSIONS

Many previous studies investigated the photosynthetic limitation under FL in C<sub>3</sub> plants, but little is known about the photosynthetic limitation under FL in C<sub>4</sub> plants. We here found that sunfleck utilization efficiency of maize is mainly limited by  $g_s$ . Therefore, increasing  $g_s$  has the potential to improve dynamic photosynthetic efficiency and biomass for maize grown in high-precipitation regions. When exposed to FL in dry air,  $g_s$  decreased only slightly in C<sub>4</sub> maize but largely decreased in C<sub>3</sub> tomato. As a result, dry air induced a smaller loss in photosynthetic carbon gain under FL in maize than in tomato. This result partially explains why C<sub>4</sub> crops usually reach greater yields than C<sub>3</sub> crops when grown in regions with low relative humidity. At high relative humidity, the photosynthetic advantage of C<sub>4</sub> crops compared with C<sub>3</sub> crops is diminished by the relatively high  $g_s$  in FL. Taking together, the effect of relative humidity on dynamic photosynthesis should be taken into consideration when modelling crop yield under future climatic change.

## AUTHOR CONTRIBUTIONS

WH and YJY designed experiments; QS, XQW and BH performed experiments; QS, XQW and WH analyzed data; and WH wrote the paper, which was edited by other authors.

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## DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

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