# CYCLODIPEPTIDES FROM THE ROOTS OF PANAX NOTOGINSENG

WANG Shuang-ming<sup>1,2</sup>, TAN Ning-hua<sup>1\*</sup>, YANG Ya-bin<sup>1</sup>, HE Min<sup>1</sup>

- (1. State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204, China;
  - 2. Tianiin Tianshili Pharmaceutical Group Co. Ltd, Tianjin 300142, China)

Abstract From the roots of Panax notoginseng fourteen cyclodipeptides 1 ~ 14 were isolated including one new compound(1), seven new natural compounds( $4 \sim 10$ ) and six known compounds( $2 \sim 3, 11 \sim 14$ ). Their structures were elucidated as following based on spectral methods: cyclo-(Leu-Thr)(1), cyclo-(Leu-Ile)(2), cyclo-(Leu-Val) (3), cyclo-(Ile-Val)(4), cyclo-(Leu-Ser)(5), cyclo-(Leu-Tyr)(6), cyclo-(Val-Pro)(7), cyclo-(Ala-Pro)(8), cyclo-(Phe-Tyr)(9), cyclo-(Phe-Ala)(10), cyclo-(Phe-Val)(11), cyclo-(Leu-Ala)(12), cyclo-(Ile-Ala)(13), cyclo-(Val-Ala) (14). Among them Compounds 2 and 11, 3 and 4, 12 and 13 are mixtures with 2:1, 1:1, 2:1 ratios, respectively.

Key words Panax notoginseng; Araliaceae; cyclodipeptides

### 三七环二肽成分

王双明1,2 谭宁华1\* 杨亚滨1 何 敏1

(1.中国科学院昆明植物研究所植物化学与西部植物资源持续利用国家重点实验室 昆明 650204; 2. 天津天士力制药集团有限公司药物研究所 天津 300142)

摘 要 从三七(Panax notoginseng)的根中分离得到 14 个环二肽成分,通过波谱解析其结构分别鉴定为环-(亮氨酸-苏氨酸)(1)、环-(亮氨酸-异亮氨酸)(2)、环-(亮氨酸-缬氨酸)(3)、环-(异亮氨酸-缬氨酸)(4)、环-(亮 氨酸-丝氨酸)(5)、环-(亮氨酸-酪氨酸)(6)、环-(缬氨酸-脯氨酸)(7)、环-(丙氨酸-脯氨酸)(8)、环-(苯丙氨酸-酪氨酸)(9)、环-(苯丙氨酸-丙氨酸)(10)、环-(苯丙氨酸-缬氨酸)(11)、环-(亮氨酸-丙氨酸)(12)、环-(异亮氨 酸-丙氨酸)(13)、环-(缬氨酸-丙氨酸)(14)。其中化合物 1 为新化合物,化合物 4 ~ 10为新天然化合物,化合 物 2~3、11~14为已知化合物;化合物 2和11、3和4、12和13分别为一对混合物,比例分别为2:1、1:1和2:

关键词 三七; 五加科; 环二肽

#### Introduction

Panax notoginseng (Bur.) F. H. Chen (Araliaceae) is one of famous Traditional Chinese Medicines. There are a lot of chemical studies on it, especially saponins. In the communication<sup>[1]</sup> we isolated cyclodipeptides  $1 \sim 14$  from the roots of P. notoginseng, in which compound 1 is one new compound, compounds 4

fractions of the  $H_2O$  extracts of the roots of P. notoginseng by column chromatography as described in the experimental.

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Colorless needles (CH3OH), gave a Compound 1 negative ninhydrin reaction but positive after

 $\sim 10$  are new natural compounds, compounds 2  $\sim 3$  and

 $11 \sim 14$  are known compounds, and compounds 2 and 11, 3 and 4, 12 and 13 are mixtures with 2:1,1:1,2:1

ratios, respectively. In this full paper we report the

Cyclodipeptides 1 ~ 14 were isolated from the EtOAc

structure elucidation of compounds  $1 \sim 14$ .

Results and Discussion

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\* Corresponding author Tel: 86-871-5223800; Fax: 86-871-5223800; E-mail: nhtan@mail. kib. ac. cn

hydrolysis with 6 mol/L HCl<sup>[2]</sup>. Its molecular formular was determined as C10H18O3N2 by means of DEPT spectrum and FAB+-MS in which its quasimolecular ion peak at m/z 215 [(M+1)<sup>+</sup>]. The 100 MHz <sup>13</sup>C NMR spectrum clearly showed the presence of two amides CO at 166. 8, 168. 6, four mehtines CH at  $\delta 66.9$ , 60.5, 52.8, 23.3, one methene CH<sub>2</sub> at  $\delta 45.0$ , three methyls CH<sub>3</sub> at  $\delta 23.1$ , 21. 5, 20. 1, respectively. The 400 MHz <sup>1</sup>H NMR spectrum clearly showed the presence of two amides NH at  $\delta 8.02, 8.21$ , three mehtines CH at  $\delta 4.00 \sim$ 5.02, one methine CH and one methene CH<sub>2</sub> at  $\delta$ 1.55  $\sim$  1.80, three methyls CH<sub>3</sub> at  $\delta$ 0.82  $\sim$  1.06, respectively. These facts indicated that 1 is a cyclodipeptide and composed of Leu (1eq) and Thr (1eq). Therefore, the structure of 1, a new cyclodipeptide, was elucidated as cyclo-(Leu-Thr).

Compound 2 and compound 11 A mixture with a 2: 1 ratio, gave a negative ninhydrin reaction but positive after hydrolysis with 6 mol/L  $HCl^{[2]}$ . The positive FAB-MS showed two different quasimolecular ion peaks at m/z 453[ $(2M+1)^+$ ], 227[ $(M+1)^+$ ] for 2 and 493[ $(2M+1)^+$ ], 247[ $(M+1)^+$ ] for 11.

Their moleclular formulars were determined as  $C_{12}H_{22}$   $O_2N_2$  for 2 and  $C_{14}H_{18}O_2N_2$  for 11 by means of DEPT spectrum and FAB<sup>+</sup>-MS. The 400 MHz <sup>1</sup>H NMR and 100 MHz <sup>13</sup>C NMR spectra clearly showed the presence of four amides NH at  $\delta 8.17, 8.12, 8.04, 7.92$  and two groups of amides CO at  $\delta 166.8, 168.3$ , respectively. Using 2D NMR including COSY, HMQC and HMBC we found that 2 was composed of Leu (leq) and Ile (leq), and 11 was composed of Phe (leq) and Val (leq). Thus, the structures of 2,a new cyclodipeptide, and 11,a known cyclodipeptide, were determined as cyclo-(Leu-Ile) and cyclo-(Phe-Val), respectively.

With the same method as compound 1, compound 7, compound 8, compound 9, compound 12, compound 13 and compound 14 were determined as cyclo-(Val-Pro), cyclo-(Ala-Pro), cyclo-(Phe-Tyr), cyclo-(Leu-Ala), cyclo-(Ile-Ala), cyclo-(Val-Ala), respectively. With the same method as compound 2 and compound 11, compound 3, compound 4, compound 5, compound 6 and compound 10 were elucidated as cyclo-(Leu-Val), cyclo-(Ile-Val), cyclo-(Leu-Ser), cyclo-(Leu-Tyr), cyclo-(Phe-Ala), respectively.

Fig. 1 Structures of cyclodipeptides  $1 \sim 14$  from Panax notoginseng

## Experimental

General Mps were recorded on XRC-1(uncorr.). IR spectra were taken on Bio-Rad FTS-135 infrared spectrometer with KBr disc. UV spectra were obtained on UV 210A spectrophotometer. NMR spectra were measured on Bruker AM-400 NMR spectrometer using TMS as the internal standard. MS

spectra were carried out on VG Autospec-3000 mass spectrometer.

Extraction and isolation The  $H_2O$  syrup (160 kg) was dissolved in  $H_2O$  and extracted with petroleum ether, EtOAc and n-BuOH, respectively. Then the EtOAc solution was evaporated and the residues (6 kg) was defatted with petroleum ether and then subjected to D-101 resin column eluting with  $H_2O$  and  $CH_3OH$  for removing a

lot of water-soluble principles. The CH<sub>3</sub>OH fraction (800 g) was chromatoghrophied on Al<sub>2</sub>O<sub>3</sub> column eluting with CH<sub>3</sub>OH, detected with the TLC chemical method for cyclopeptides<sup>[2]</sup>, and collected the cyclopeptide fraction (130 g). Then by the combination of silica gel and Rp-18 column, from the cyclopeptide fraction we obtained 1 (3 mg), 2 and 11(25 mg), 3 and 4(24 mg), 5(3 mg), 6(25 mg), 7(2 mg), 8(7 mg), 9(20 mg), 10, 12(7 mg), 13 (50 mg, including 12) and 14(15 mg), respectively.

Cyclo-(Leu-Thr) (1)  $C_{10}H_{18}O_3N_2$ , needles (CH<sub>3</sub>OH), mp. 280 ~ 282 °C. ¹H NMR (DMSO): δ 8. 21 (1H, s, Leu<sub>NH</sub>)<sup>a</sup>, 8. 02 (1H, s, Thr<sub>NH</sub>)<sup>a</sup>, 5. 02 (1H, d, J = 5. 4Hz, Thr<sub>a</sub>)<sup>b</sup>, 4. 00 (2H, m, Thr<sub>β</sub> and Leu<sub>α</sub><sup>b</sup>), 1. 80 (1H, m, Leu<sub>γ</sub>), 1. 68 (1H, m, Leu<sub>γ</sub>), 1. 55 (1H, m, Leu<sub>γ</sub>), 1. 06 (3H, d, J = 6. 5 Hz, Thr<sub>γ</sub>), 0. 84 (3H, d, J = 6. 6 Hz, Leu<sub>δ1</sub>), 0. 82 (3H, d, J = 6. 5 Hz, Leu<sub>δ2</sub>). <sup>13</sup> C NMR (DMSO): δ 168. 6 (Leu<sub>CO</sub>)<sup>c</sup>, 166. 8 (Thr<sub>CO</sub>)<sup>c</sup>, 66. 9 (Thr<sub>β</sub>), 60. 5 (Thr<sub>α</sub>)<sup>d</sup>, 52. 8 (Leu<sub>α</sub>)<sup>d</sup>, 45. 0 (Leu<sub>β</sub>), 23. 3 (Leu<sub>γ</sub>), 23. 1 (Leu<sub>δ1</sub>), 21. 5 (Leu<sub>δ2</sub>), 20. 1 (Thr<sub>γ</sub>). Assignments<sup>a-d</sup> may be reversed. FAB<sup>+</sup>-MS m/z: 215 (M + 1)<sup>+</sup>, 115.

Cyclo-(Leu-Ile)(2) and Cyclo-(Phe-Val)(11)  $2:C_{12}H_{22}$  $O_2N_2$ ; 11:  $C_{14}H_{18}O_2N_2$ . <sup>1</sup>H NMR (DMSO):  $\delta$  2: 8. 17 (1H, s, Leu<sub>NH</sub>), 8. 04 (1H, s, Ile<sub>NH</sub>), 3. 77 (1H, br.,  $Leu_{n}$ ), 3.70(1H, br.,  $Ile_{n}$ ), 1.83(2H, m,  $Leu_{r}$  and  $Ile_{\theta}$ ),  $1.63(1H, m, Leu_{el}), 1.44(2H, m, Leu_{el}), 1.19$ (1H, m, Ile<sub>12</sub>), 0. 89 (12H, m, Leu<sub>31</sub>, Leu<sub>32</sub>, Ile<sub>13</sub> and  $Ile_{\delta}$ ); 11: 8. 12 (1H, s,  $Val_{NH}$ )<sup>a</sup>, 7. 92 (1H, s,  $Phe_{NH}$ )<sup>a</sup>, 7.  $19(5H, m, Phe_b)$ , 4.  $21(1H, br., Phe_a)^b$ , 3.  $53(1H, phe_a)^b$ br.,  $Val_{\alpha}$ )<sup>b</sup>, 3. 14(1H, dd, Phe<sub>81</sub>), 2. 90(1H, dd, Phe<sub>82</sub>), 1.63 (1H, m,  $Val_{\theta}$ ), 0.65 (3H, d, J = 6.9 Hz,  $Val_{\gamma 1}$ ),  $0.26(3H, d, J = 6.9 \text{ Hz}, \text{Val}_{12}).$  <sup>13</sup>C NMR(DMSO):  $\delta$ 2:168. 3 (Leu<sub>m</sub>)<sup>c</sup>, 166. 8 (Ile<sub>m</sub>)<sup>c</sup>, 58. 8 (Ile<sub>a</sub>), 52. 3  $(Leu_g)$ , 43. 6  $(Leu_g)$ , 38. 3  $(Ile_g)$ , 24. 3  $(Ile_{\gamma 1})$ , 23. 5  $(Leu_y)$ , 23.  $0(Leu_{\delta 1})$ , 21.  $7(Leu_{\delta 2})$ , 15.  $1(Ile_{\gamma 2})$ , 11. 6  $(Ile_{\delta})$ ; 11: 168.3  $(Val_{CO})^d$ , 166. 8  $(Phe_{CO})^d$ , 136. 0  $(Phe_{41'})$ , 130. 2  $(Phe_{43',5'})$ , 127. 9  $(Phe_{42',6'})$ , 126. 4  $(Phe_{44'})$ , 59. 2  $(Val_{y1} Val_{\alpha})^e$ , 55. 0  $(Phe_{\alpha})^e$ , 37. 9  $(Phe_{\beta})$ ,  $31.0(Val_9)$ ,  $18.2(Val_{71})$ ,  $16.3(Val_{72})$ . Assignments \*\*e may be reversed. FAB<sup>+</sup>-MS  $m/z:2:453(2M+1)^+,227$  $(M+1)^+$ ; 11:493(2M+1)+,247(M+1)+. The ratio of 2 and 11 is 2:1.

Cyclo-(Leu-Val)(3) and Cyclo-(Ile-Val)(4) 3 and 4:  $C_{11}H_{20} O_2N_2$ . <sup>1</sup>H NMR (DMSO): δ 3: 8. 20 (1H, s, Leu<sub>NH</sub>), 8.06(1H, s, Val<sub>NH</sub>), 3.74(1H, br., Leu<sub>α</sub>), 3.60 (1H, m, Val<sub>α</sub>), 2.08(1H, m, Val<sub>β</sub>), 1.83(1H, m, Leu<sub>γ</sub>), 1.58(1H, m, Leu<sub>γ1</sub>), 1.42(1H, m, Leu<sub>γ2</sub>), 0.80 ~ 0.94 (12H, m, Leu<sub>δ1</sub>, Leu<sub>δ2</sub>, Val<sub>γ1</sub> and Val<sub>γ2</sub>); 4:7.95(1H, s, Ile<sub>NH</sub>), 7.93(1H, s, Val<sub>NH</sub>), 3.74(1H, br., Ileα), 3.67

 $\begin{array}{l} (1H, br.\ , Val\alpha)\ , 2.\ 17(1H, m,\ Val_\beta)\ , 1.\ 83(1H, m,\ Ile_\beta)\ , \\ 1.\ 42(1H,\ m,\ Ile_{\gamma 1})\ , 1.\ 18(1H,\ m,\ Ile_{\gamma 2})\ , 0.\ 80 \sim 0.\ 94 \\ (12H,\ m,\ Leu_{\delta 1},\ Leu_{\delta 2},\ Val_{\gamma 1}\ \ and\ \ Val_{\gamma 2})\ .^{13}\ C\ \ NMR \\ (DMSO):\ \delta\ 3:\ 168.\ 6\ (Leu_{CO})\ , 167.\ 0\ (Val_{CO})\ , 59.\ 6 \\ (Val_{\alpha})\ , 52.\ 5\ (Leu_{\alpha})\ , 44.\ 0\ (Leu_{\beta})\ , 31.\ 6\ (Val_{\beta})\ , 23.\ 6 \\ (Leu_{\gamma})\ , 23.\ 1\ (Leu_{\delta 1})\ , 21.\ 6\ (Leu_{\delta 2})\ , 18.\ 8\ (Val_{\gamma 1})^a\ , 17.\ 4 \\ (Val_{\gamma 2})\ ;\ 4:\ 167.\ 6\ (Ile_{CO})\ , 167.\ 5\ (Val_{CO})\ , 59.\ 2\ (Val_{\alpha})\ , \\ 58.6\ (Ile_{\alpha})\ , 38.\ 0\ (Ile_{\beta})\ , 31.\ 0\ (Val_{\beta})\ , 24.\ 4\ (Ile_{\gamma 1})\ , 18.\ 7 \\ (Val_{\gamma 1})^a\ , 17.\ 4\ (Val_{\gamma 2})\ , 15.\ 1\ (Ile_{\gamma 2})\ , 11.\ 9\ (Ile_{\delta})\ . \\ Assignments\ ^a\ may\ be\ reversed.\ FAB^+\ -MS\ m/z\ ; 425(2M+1)^+\ , 213(M+1)^+\ , 113\ , 86\ , 72\ , 59.\ The\ ratio\ of\ 3\ and\ 4\ is\ 1:1. \end{array}$ 

Cyclo-(Leu-Ser) (5)  $C_9H_{16}O_3N_2$ , needles (CH<sub>3</sub>OH), mp. 240 ~ 242 °C . IR  $v_{max}$  cm<sup>-1</sup>:3443,3197,3057,2959, 2927, 2856, 1669, 1468, 1333. <sup>1</sup>H NMR (DMSO): δ 8.23(1H, s, Leu<sub>NH</sub>),7.93(1H, s, Ser<sub>NH</sub>),5.14(1H, m, Ser<sub>β1</sub>),3.79(2H, m, Leu<sub>α</sub> and Ser<sub>α</sub>),3.42(1H, m, Ser<sub>β2</sub>), 1.79(1H, m, Leu<sub>γ</sub>),1.59(2H, m, Leu<sub>β</sub>),0.84(6H, m, Leu<sub>δ1</sub> and Leu<sub>δ2</sub>). <sup>13</sup>C NMR(DMSO): δ 168.2(Leu<sub>CO</sub>), 166.3(Ser<sub>CO</sub>),62.3(Ser<sub>β</sub>),57.2(Ser<sub>α</sub>),52.7(Leu<sub>α</sub>), 44.6(Leu<sub>β</sub>),23.3(Leu<sub>γ</sub>),23.1(Leu<sub>δ1</sub>),21.5(Leu<sub>δ2</sub>). FAB<sup>+</sup>-MS m/z:401(2M+1)<sup>+</sup>,201(M+1)<sup>+</sup>,115,86.

Cyclo-(Leu-Tyr)(6)  $C_{15}H_{20}O_3N_2$ , needles (CH<sub>3</sub>OH), mp. 260  $\sim\!262$  °C . UV  $\lambda_{max}^{CH,OH}$  nm; 202. 5, 225. 5, 277. 5. IR  $v_{\text{max}}$  cm<sup>-1</sup>:3439,3321,3215,2959,2932,1673,1616, 1598, 1515, 1469, 1384, 1335, 1244, 842, 818. <sup>1</sup>H NMR (DMSO):  $\delta$  9.27(1H, s, Tyr<sub>b4'-OH</sub>), 8.07(2H, m, Leu<sub>NH</sub> and  $Tyr_{NH}$ ), 6. 88 (2H, d, J = 8.2 Hz,  $Tyr_{42',6'}$ ), 6. 63  $(2H, d, J = 8.1 \text{ Hz}, Tyr_{43',5'}), 4.05(1H, br., Tyr_a),$  $3.44(1H, br., Leu_{q}), 3.00(1H, dd, J = 13.5, 3.3 Hz,$  $Tyr_{g1}$ ), 2. 67 (1H, dd, J = 13.6, 4. 6 Hz,  $Tyr_{g2}$ ), 1. 40  $(1H, m, Leu_{\gamma}), 0.72(1H, m, Leu_{\beta 1}), 0.62(6H, m, Leu_{\delta 1})$ and Leu<sub>22</sub>),  $0.08(1H, m, Leu_{22})$ . <sup>13</sup>C NMR(DMSO):  $\delta$  $167.5 (Leu_{CO}), 166.3 (Tyr_{CO}), 156.4 (Tyr_{44'}), 131.2$  $(Tyr_{42',6'})$ , 125. 9  $(Tyr_{41'})$ , 114. 9  $(Tyr_{43',5'})$ , 55. 7  $(Tyr_{\alpha})$ , 52. 3 (Leu<sub> $\alpha$ </sub>), 43. 7 (Leu<sub> $\beta$ </sub>), 37. 7 (Tyr<sub> $\beta$ </sub>), 23. 0  $(\text{Leu}_{\gamma})$ , 22. 7  $(\text{Leu}_{\delta 1})$ , 21. 4  $(\text{Leu}_{\delta 2})$ . FAB<sup>+</sup>-MS m/z:  $279(M+3)^{+},113.$ 

Cyclo·(Val-Pro) (7)  $C_{10} H_{16} O_2 N_2$ , needles (CH<sub>3</sub>OH), mp, 145 ~ 147 °C. IR  $v_{max}$  cm<sup>-1</sup>: 3247,2959,2927,2856, 1650,1454,1289,1275. <sup>1</sup>H NMR(DMSO): δ 8. 38(1H, d, J = 3.8 Hz, Val<sub>NH</sub>), 4. 13(1H, m, Val<sub>α</sub>), 3. 34(3H, m, Pro<sub>α</sub> and Pro<sub>δ</sub>), 2. 14(1H, m, Val<sub>β</sub>), 1. 70 ~ 2. 02(4H, m, Pro<sub>β</sub> and Pro<sub>γ</sub>), 0. 90(3H, d, J = 11.8 Hz, Val<sub>γ1</sub>), 0. 86 (3H, d, J = 10.2 Hz, Val<sub>γ2</sub>). <sup>13</sup> C NMR (DMSO): δ 168.9 (Pro<sub>ΩO</sub>)<sup>a</sup>, 165.2 (Val<sub>QO</sub>)<sup>a</sup>, 62.7 (Pro<sub>α</sub>)<sup>b</sup>, 57.8 (Val<sub>α</sub>)<sup>b</sup>, 45.2 (Pro(<sub>δ</sub>), 32.5 (Val<sub>β</sub>), 29.0 (Pro<sub>β</sub>), 21.6 (Pro<sub>γ</sub>), 19.0 (Val<sub>γ1</sub>), 18.3 (Val<sub>γ2</sub>). Assignments <sup>a-b</sup> may

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be reversed. FAB<sup>+</sup>-MS m/z: 197(M+1)<sup>+</sup>, 115, 98, 70.

Cyclo-(Ala-Pro) (8)  $C_8H_{12}O_2N_2$ , needles (CH<sub>3</sub>OH), mp. 170 ~ 172 °C . IR  $v_{max}$  cm  $^{-1}$ : 3290, 2925, 2854, 1658, 1420. H NMR (DMSO):  $\delta$  8. 17 (1H, s, Ala<sub>NH</sub>), 4. 15 (1H, t, J = 7. 6 Hz,  $Pro_a$ ), 4. 07 (1H, dd, J = 13. 6, 6. 8 Hz, Ala<sub>a</sub>), 3. 34 (2H, m,  $Pro_b$ ), 1. 74 ~ 2. 12 (4H, m,  $Pro_b$ ) and  $Pro_y$ ), 1. 19 (3H, d, J = 6. 8 Hz, Ala<sub>b</sub>). C NMR (DMSO):  $\delta$  170. 0 ( $Pro_{\infty}$ ) a, 166. 6 (Ala<sub>\infty</sub>) a, 58. 7 ( $Pro_a$ ) b, 50. 2 (Ala<sub>a</sub>) b, 44. 9 ( $Pro_b$ ), 27. 7 ( $Pro_b$ ), 22. 4 ( $Pro_y$ ), 15. 3 (Ala<sub>b</sub>). Assignments ab may be reversed. FAB - MS m/z: 337 (2M + 1) + , 169 (M + 1) + , 115, 98,86,70.

Cyclo-(Phe-Tyr)(9)  $C_{18}H_{18}O_3N_2$ , needles (CH<sub>3</sub>OH), mp. 291 ~ 293 °C. UV  $\lambda_{max}^{CH,OH}$  nm: 202. 5, 223, 278. IR  $v_{\text{max}} \text{ cm}^{-1}$ : 3541, 3223, 3032, 2924, 2855, 1678, 1657, 1616,1518,1496,1466. <sup>1</sup>H NMR(DMSO):  $\delta$  9.27(1H,  $s, Tyr_{44'-OH}), 7.89(2H, m, Phe_{NH}), 7.26(2H, m)$  $t, J = 7.4 \text{ Hz}, Phe_{43',5'}), 7.20 (1H, d, J = 7.3 \text{ Hz},$  $Phe_{44'}$ ), 7.01(2H, d, J = 7.3 Hz,  $Phe_{42',6'}$ ), 6.81(2H, d,  $J = 8.3 \text{ Hz}, \text{Tyr}_{42'.6'}), 6.66 (2H, d, J = 8.2 \text{ Hz},$  $Tyr_{\phi 3',5'}$ ), 3.92(1H, br.,  $Tyr_{\sigma}$ )<sup>a</sup>, 3.87(1H, br.,  $Phe_{\sigma}$ )<sup>a</sup>, 2. 50 (2H, m, Phe<sub>B</sub>)<sup>b</sup>, 2. 14 (2H, m, Tyr<sub>B</sub>)<sup>b</sup>. <sup>13</sup>C NMR (DMSO):  $\delta$  166. 3 (Tyr<sub>CO</sub>)°, 166. 2 (Phe<sub>CO</sub>)°, 156. 1  $(Tyr_{44'})$ , 136. 6 (Phe<sub>41'</sub>), 130. 8 ( $Tyr_{42',6'}$ ), 129. 7  $(Phe_{43',5'})$ , 128. 1  $(Phe_{42',6'})$ , 126. 4  $(Phe_{44'}$  and  $Tyr_{41'})$ , 115. 0 ( $Tyr_{\phi 3',5'}$ ), 55. 7 ( $Tyr_{\alpha}$ )<sup>d</sup>, 55. 4 ( $Phe_{\alpha}$ )<sup>d</sup>, 39. 1 (Tyr<sub>8</sub>)<sup>e</sup>, 38.9 (Phe<sub>8</sub>)<sup>e</sup>. Assignments <sup>a-e</sup> may be reversed.  $FAB^+-MS m/z:310(M^+),282,85.$ 

Cyclo-(Phe-Ala) (10)  $C_{12}H_{14}O_2N_2$ . <sup>1</sup>H NMR(DMSO); δ 8.16(1H, s, Phe<sub>NH</sub>), 8.06(1H, s, Ala<sub>NH</sub>), 7.29(2H, t, J = 7.2 Hz, Phe<sub>43',5'</sub>), 7.23(1H, d, J = 6.8 Hz, Phe<sub>44'</sub>), 7.15 (2H, d, J = 7.1 Hz, Phe<sub>42',6'</sub>), 4.18 (1H, br., Phe<sub> $\alpha$ </sub>), 3.61(1H, m, Ala<sub> $\alpha$ </sub>), 3.13(1H, dd, J = 3.4, 13.4 Hz, Phe<sub> $\beta$ 1</sub>), 2.85(1H, dd, J = 4.9, 13.4 Hz, Phe<sub> $\beta$ 2</sub>), 0.44 (3H, d, J = 6.9 Hz, Ala<sub> $\beta$ 1</sub>). <sup>13</sup>C NMR(DMSO): δ 167.6 (Ala<sub>CO</sub>), 165.8 (Phe<sub>CO</sub>), 136.0 (Phe<sub> $\beta$ 1'</sub>), 130.3 (Phe<sub> $\beta$ 3',5'</sub>), 127.9 (Phe<sub> $\beta$ 2',6'</sub>), 126.5 (Phe<sub> $\beta$ 4'</sub>), 55.3 (Phe<sub> $\alpha$ 0</sub>), 49.7(Ala<sub> $\alpha$ 0</sub>), 38.3(Phe<sub> $\beta$ 0</sub>), 19.6(Ala<sub> $\beta$ 0</sub>). FAB<sup>+</sup>-MS m/z: 218 M<sup>+</sup>.

Cyclo-(Leu-Ala) (12)  $C_9H_{16}O_2N_2$ , needles (CH<sub>3</sub>OH), mp. 145~147 °C . IR  $v_{max}$  cm<sup>-1</sup>: 3424,3192,3051,2959,

2888, 1678, 1469, 1327. <sup>1</sup>H NMR(DMSO):  $\delta$  8. 22(1H, s, Leu<sub>NH</sub>)<sup>a</sup>, 8. 12(1H, s, Ala<sub>NH</sub>)<sup>a</sup>, 3. 94(1H, dd, J = 6. 9, 13. 7 Hz, Leu<sub>α</sub>)<sup>b</sup>, 3. 64(1H, m, Ala<sub>α</sub>)<sup>b</sup>, 1. 73(1H, m, Leu<sub>γ</sub>), 1. 49(2H, m, Leu<sub>β</sub>), 1. 21(3H, d, J = 6. 8 Hz, Ala<sub>β</sub>), 0. 87(3H, d, J = 6. 6 Hz, Leu<sub>δ1</sub>), 0. 85(3H, d, J = 6. 6 Hz, Leu<sub>δ2</sub>). <sup>13</sup>C NMR(DMSO):  $\delta$  168. 8(Leu<sub>α</sub>)<sup>c</sup>, 168. 4(Ala<sub>α</sub>)<sup>c</sup>, 53. 4(Leu<sub>α</sub>)<sup>d</sup>, 49. 0(Ala<sub>α</sub>)<sup>d</sup>, 42. 1(Leu<sub>β</sub>), 23. 6(Leu<sub>γ</sub>), 22. 8(Leu<sub>δ1</sub>), 21. 8(Leu<sub>δ2</sub>), 17. 8(Ala<sub>β</sub>). Assignments <sup>a-d</sup> may be reversed. FAB<sup>+</sup>-MS m/z: 184 (M<sup>+</sup>), 145, 113, 97, 87, 69.

Cyclo-(Ile-Ala) (13)  $C_9H_{16}O_2N_2$ . <sup>13</sup>C NMR(DMSO); δ 168. 4 (Ala<sub>CO</sub>)<sup>a</sup>, 166. 6 (Ile<sub>CO</sub>)<sup>a</sup>, 58. 8 (Ile<sub>α</sub>)<sup>b</sup>, 49. 6 (Ala<sub>α</sub>)<sup>b</sup>, 38. 0 (Ile<sub>β</sub>), 24. 2 (Ile<sub>γ1</sub>), 19. 9 (Ala<sub>β</sub>), 15. 0 (Ile<sub>γ2</sub>), 11. 8 (Ile<sub>δ</sub>). Assignments <sup>a-b</sup> may be reversed. FAB<sup>+</sup>-MS m/z: 370 (2M + 2)<sup>+</sup>, 184 M<sup>+</sup>, 169, 157, 141,128,115,98,86,70. The ratio of 12 and 13 is 2:1.

Cyclo-(Val-Ala) (14)  $C_8H_{14}O_2N_2$ , needles (CH<sub>3</sub>OH), mp. 265 ~ 267 °C . IR  $v_{max}$  cm<sup>-1</sup>: 3326, 3193, 3052, 2968, 2934, 2896, 1681, 1669, 1457, 1343, 1332, 1140, 861, 832. ¹H NMR (DMSO):  $\delta$  8. 15 (1H, s, Ala<sub>NH</sub>)<sup>a</sup>, 8. 01 (1H, s, Val<sub>NH</sub>)<sup>a</sup>, 3. 87 (1H, dd, J = 6.7, 13.5 Hz, Ala<sub>a</sub>), 3. 67 (1H, s, Val<sub>a</sub>), 2. 13 (1H, m, Val<sub>β</sub>), 1. 25 (3H, d, J = 7.0 Hz, Ala<sub>β</sub>), 0. 93 (3H, d, J = 7.0 Hz, Val<sub>γ1</sub>), 0. 81 (3H, d, J = 6.8 Hz, Val<sub>γ2</sub>). ¹³C NMR (DMSO):  $\delta$  168. 6 (Ala<sub>CO</sub>)<sup>c</sup>, 166. 5 (Val<sub>CO</sub>)<sup>c</sup>, 59. 4 (Val<sub>α</sub>)<sup>d</sup>, 49. 6 (Ala<sub>α</sub>)<sup>d</sup>, 31.0 (Val<sub>β</sub>), 20.0 (Ala<sub>β</sub>)<sup>e</sup>, 18.4 (Val<sub>γ1</sub>1)<sup>e</sup>, 16.6 (Val<sub>γ2</sub>)<sup>e</sup>. Assignments <sup>a-e</sup> may be reversed. FAB<sup>+</sup>-MS m/z: 171 (M+1)<sup>+</sup>, 98, 72.

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#### References

- Tan NH, Wang SM, Yang YB, et al. Cyclodipeptides of Panax notoginseng and lactams of Panax ginseng. Acta Botanica Yunnanica, 2003, 25(3):366-368
- Zhou J, Tan NH. Application of a new TLC chemical method for detection of cyclopeptides in plants. Chinese Science Bulletin, 2000, 45(20):1825-1831