

# 化感植物向日葵叶化学成分的研究

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**摘要:** 从向日葵 (*Helianthus annuus* L.) 叶子的甲醇提取物中分离得到了 8 个已知化合物, 其结构经波谱解析分别确定为: (—)-kaur-16-en-19-oic acid (1)、(6*R*, 10*R*)-6, 10, 14-三甲基-十五烷-2-酮(2)、维生素 E(3)、dehydrocostus lactone(4)、(—)- $\alpha$ -tocospirone(5)、angeloygrandifloric acid(6)、*trans*-phytol(7) 及 3(20)-phytene-1, 2-diol(8)。其中化合物 2, 5 和 8 为首次从该植物中分离得到。

**关键词:** 向日葵; 叶; 化学成分

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## Chemical Component from Allelopathic Cultivar Sunflower Leaves

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**Abstract:** The methanol extracts of the leaves of cultivar sunflower (*Helianthus annuus* L.) afforded eight known compounds (1~8), by spectroscopic analysis, their structures were elucidated as (—)-kaur-16-en-19-oic acid(1), (6*R*, 10*R*)-6, 10, 14- trimethyl-2-pentadecanone(2),  $\alpha$ -tocopherol(3), dehydrocostus lactone(4), (—)- $\alpha$ -tocospirone(5), angeloygrandifloric acid(6), *trans*-phytol(7), and 3(20) phytene-1, 2-diol(8). Compound 2, 5 and 8 were isolated from this plant for the first time.

**Key words:** sunflower; leaves; constituents

Our indiscriminate use of synthetic chemicals for herbicides and pest control poses a serious threat to our health and environment<sup>[1]</sup>. One way to overcome such problem is to find natural phytotoxins which is degradable and safe for agricultural production and human beings *via* isolation, identification and synthesis of active compounds from phytotoxic plant species<sup>[2]</sup>. Sunflower (*Helianthus*

*annuus* L.), a well-known allelopathic plant, produced a variety of secondary metabolites, some of which possess significant allelopathic property<sup>[3]</sup>. To search for bioactive natural products from medicinal plants, the chemical component of cultivar sunflower were carried out. In this paper, we herein described the isolation and characterization of compounds 1~8 (Fig. 1) from sunflower leaves.

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## 1 Experimental

### 1.1 General

Melting points were obtained on a XRC-1 apparatus and uncorrected. Optical rotations were measured on a Horiba SEPA-300 polarimeter. NMR spectra were recorded on Bruker AV-400 and DRX-500 spectrometers with TMS as an internal standard.  $\delta$  in  $10^{-6}$ ,  $J$  in Hz. IR spectra were obtained with a Bruker Tensor 27 FT-IR with KBr pellets. UV spectrum was measured on a Shimadzu double-beam 210A spectrometer. MS (EI, FAB) were recorded with a VG Autospec-3000 spectrometer,  $m/z$  (rel. int.). ESI and HR-ESI-MS was recorded with an API QSTAR Pulsar 1 spectrometer. Column chromatography (CC) was carried out on silica gel (200~300 mesh, Qingdao Marine Chemical Ltd., Qingdao, P. R. China) and Sephadex LH-20 (Amersham Biosciences, Uppsala, Sweden).

### 1.2 Plant material

Leaves of *Helianthus annuus* L. were collected in September 2003 in Wuzhong City, Ningxia, China and were identified by Mr. Wu Z H, at Northwest Institute of Botany, Yangling, Shaanxi, and were deposited at Natural Medicine Chemistry Research Centre, College of Sciences, Northwest A & F University.

### 1.3 Extraction and isolation

The dried and powdered leaves (20 kg) were extracted three times with MeOH at room temperature. The combined extracts were concentrated *in vacuo* to give a deep green gum (2 230 g), which was partitioned between  $\text{CHCl}_3$  and  $\text{H}_2\text{O}$ . After removal of the organic solvent under reduced pressure, the resulting residue (498 g) was subjected to CC on silica gel eluting with  $\text{CHCl}_3/\text{MeOH}$  mixtures of increasing polarity to yield 8 fractions (frs 1~8) according to TLC analysis.

Fr. 2 was subjected to CC on silica gel eluting with petroleum ether/EtOAc (50 : 1 to 10 : 1) to provide four subfractions (A~D). Subfraction A was separated by silica gel CC using petroleum ether/EtOAc (40 : 1) to afford compound **1** (3.15 g). Subfraction B was rechromatographed over silica gel CC with petroleum ether/ $\text{CHCl}_3$  (6 : 1), and

Sephadex LH-20 column ( $\text{CHCl}_3/\text{MeOH}$  1 : 1), followed by Prep-TLC (petroleum ether/ether, 7 : 1) to give compound **2** (13 mg). Subfraction C was rechromatographed over silica gel column using petroleum ether/EtOAc (50 : 1), then Sephadex LH-20 column ( $\text{CHCl}_3/\text{MeOH}$ , 1 : 1) to furnish compound **3** (14 mg). Subfraction D was rechromatographed over silica gel column eluting with petroleum ether/ $\text{CHCl}_3$  (1 : 2.5), followed by Sephadex LH-20 column ( $\text{CHCl}_3/\text{MeOH}$ , 1 : 1), and Prep-TLC (petroleum ether/ether, 2.5 : 1), yielding compound **4** (12 mg). Four fractions (E~G) were obtained from fr. 3 by CC on silica gel eluted with petroleum ether/EtOAc mixtures of increasing polarity (20 : 1, 10 : 1, 3 : 1). Fraction E was rechromatographed over silica gel column using petroleum ether/acetone (60 : 1), and on Sephadex LH-20 column ( $\text{CHCl}_3/\text{MeOH}$ , 1 : 1), and further purified by Prep-TLC (petroleum ether/EtOAc, 5 : 1), yielding **5** (17 mg). Fraction F was chromatographed over silica gel eluting with petroleum ether/EtOAc (15 : 1) to afford a mixture containing compound **6** (258 mg), which was further purified by recrystallization in petroleum ether/ $\text{CHCl}_3$ . Fraction G was chromatographed over silica gel column using petroleum ether/ $\text{CHCl}_3$  (3 : 2), followed by Sephadex LH-20 column ( $\text{CHCl}_3/\text{MeOH}$ , 1 : 1) to give compounds **7** (91 mg) and **8** (18 mg).

## 2 Results and discussion

A chloroform-soluble fraction of the methanolic extract of the leaves of sunflower was subjected to repeated silica gel column chromatography, followed by Sephadex LH-20 to give compounds **1**~**8**.

(-)-**Kaur-16-en-19-oic acid (1)**  $\text{C}_{20}\text{H}_{30}\text{O}_2$ , colorless crystals, mp. 179~180°C;  $[\alpha]_{\text{D}}^{24.5} -111^\circ$  (c 1.6,  $\text{CHCl}_3$ ); + TOF-MS  $m/z$ : 303  $[\text{M}+1]^+$ . The  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR data were identical to those recorded for an authentic specimen of (-)-kaur-16-en-19-oic acid<sup>[4,5]</sup>.

(**6R, 10R**)-**6, 10, 14-Trimethyl-2-pentadecanone (2)**  $\text{C}_{18}\text{H}_{36}\text{O}$ , FAB+MS  $m/z$ : 269  $[\text{M}]^+$  (100);

$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  2.41 (2H, m), 2.13 (3H, s, H-1), 0.86 (6H, d,  $J=6.6$  Hz, H-15 and H-18), 0.85 (3H, d,  $J=6.8$  Hz), 0.83 (3H, d,  $J=6.8$  Hz), 1.60~1.00 (19H, m);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  29.9 (q, C-1), 209.5 (s, C-2), 44.2 (t, C-3), 21.4 (t, C-4), 36.5 (t, C-5), 32.6 (d, C-6), 37.2 (t, C-7), 24.4 (t, C-8), 37.2 (t, C-9), 32.8 (d, C-10), 37.4 (t, C-11), 24.8 (t, C-12), 39.4 (t, C-13), 28.0 (d, C-14), 22.6 (q, C-15), 19.6 (q, C-16), 19.7 (q, C-17), 22.7 (q, C-18). These data were identical to those recorded for an authentic specimen of (6*R*, 10*R*)-6, 10, 14-Trimethyl-2-pentadecanone<sup>[6]</sup>. The data of  $^1\text{H}$  NMR were assigned after comparison by those data of (–)- $\alpha$ -tocospirone<sup>[10]</sup> and 3(20)phytene-1, 2-diol<sup>[12]</sup>.

**$\alpha$ -Tocopherol (3)**  $\text{C}_{20}\text{H}_{50}\text{O}_2$ , colorless oil, UV  $\lambda_{\text{max}}^{\text{MeOH}}$ : 291 and 298 nm; EI-MS  $^1\text{H}$  NMR, and  $^{13}\text{C}$  NMR data were identical to those recorded for an authentic specimen of  $\alpha$ -tocopherol<sup>[7,8]</sup>, and the data of H-12'a, H-13', H-4'a and H-8'a were assigned after comparison by those data of (–)- $\alpha$ -tocospirone<sup>[10]</sup> and 3(20)phytene-1, 2-diol<sup>[12]</sup>.

**Dehydrocostus lactone (4)**  $\text{C}_{15}\text{H}_{18}\text{O}_2$ , colorless needles (EtOAc), mp. 49°C; IR  $V_{\text{max}}$   $\text{cm}^{-1}$ : 1762, 1644; + TOF-MS  $m/z$ : 231 [ $\text{M}+1$ ]<sup>+</sup>. The  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR data were identical to those recorded for an authentic specimen of dehydrocostus lactone<sup>[9]</sup>.

**(–)- $\alpha$ -Tocospirone (5)**  $\text{C}_{29}\text{H}_{50}\text{O}_4$ , colorless oil, UV  $\lambda_{\text{max}}^{\text{MeOH}}$ : 251.4 nm; IR  $V_{\text{max}}$   $\text{cm}^{-1}$ : 3487 (OH), 1697, 1679 (C=O), 1622; FAB+MS  $m/z$ : 463 [ $\text{M}$ ]<sup>+</sup> (100), 445 [ $\text{M}+\text{H}-\text{H}_2\text{O}$ ]<sup>+</sup> (35), 419 (85), 402 (15), 352 (8), 237 [ $\text{M}$ -side chain]<sup>+</sup> (8), 167 (23);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  0.83 (3H, d,  $J=6.8$  Hz, H-17a), 0.84 (3H, d,  $J=6.8$  Hz, H-13a), 0.85 (3H, d,  $J=6.8$  Hz, H-21a), 0.86 (3H, d,  $J=6.8$  Hz, H-22), 1.33 (3H, s, H-9a), 1.36 (3H, s, H-3a), 1.59 (1H, m, H-10a), 1.60 (1H, m, H-8a), 1.66 (1H, m, H-10b), 1.69 (1H, m, H-7a), 1.93 (1H, m, H-8b), 2.03 (1H, m, H-7b), 2.05 (3H, s, H-5a), 2.06 (3H, s, H-6a), 3.82 (1H, s, OH);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  198.8 (s, C-1), 93.3 (s, C-2), 81.2 (s, C-3), 24.2 (q, C-3a), 201.7 (s, C-4), 142.0 (s, C-5), 13.0 (q, C-5a), 146.9 (s, C-6),

13.4 (q, C-6a), 32.0 (t, C-7), 36.4 (t, C-8), 87.0 (s, C-9), 25.7 (q, C-9a), 41.3 (t, C-10), 22.3 (t, C-11), 37.5 (t, C-12), 32.8 (d, C-13), 19.7 (q, C-13a), 37.5 (t, C-14), 24.8 (t, C-15), 37.4 (t, C-16), 32.7 (d, C-17), 19.7 (q, C-17a), 37.3 (t, C-18), 24.4 (t, C-19), 39.3 (t, C-20), 28.0 (d, C-21), 22.6 (q, C-21a), 22.7 (q, C-22). These data were identical to those recorded for an authentic specimen of (–)- $\alpha$ -tocospirone<sup>[10]</sup>.

**Angeloygrandifloric acid (6)**  $\text{C}_{25}\text{H}_{36}\text{O}_4$ , colorless crystals, mp. 196~198°C; IR  $V_{\text{max}}$   $\text{cm}^{-1}$ : 3200~2500, 1702, 1250, 1040, 1005 and 896; EI-MS  $m/z$  (%): 400 [ $\text{M}$ ]<sup>+</sup> (8), 300 (65), 285 (70), 272 (34), 255 (22), 83 (100);  $^1\text{H}$  NMR (500 MHz,  $\text{CD}_3\text{COCD}_3$ ):  $\delta$  0.96 (3H, s, H-18), 1.23 (3H, s, H-20), 1.88 (3H, brs), 1.97 (1H, m), 2.80 (1H, m), 5.40 (1H, m), 5.08 (1H, brs), 5.13 (1H, m) and 6.07 (1H, m);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CD}_3\text{COCD}_3$ ):  $\delta$  40.6 (t, C-1), 19.0 (t, C-2), 35.1 (t, C-3), 43.8 (s, C-4), 56.6 (d, C-5), 20.9 (t, C-6), 37.4 (t, C-7), 47.6 (s, C-8), 53.0 (d, C-9), 39.9 (s, C-10), 20.7 (t, C-11), 32.7 (t, C-12), 42.6 (d, C-13), 37.7 (t, C-14), 82.6 (d, C-15), 155.6 (s, C-16), 109.9 (t, C-17), 28.9 (q, C-18), 184.0 (s, C-19), 15.8 (q, C-20), 168.1 (s, C-21), 128.3 (s, C-22), 137.3 (d, C-23), 15.8 (q, C-24), 18.5 (q, C-25). These data were identical to those recorded for an authentic specimen of angeloygrandifloric acid<sup>[4]</sup>.

**trans-Phytol (7)**  $\text{C}_{20}\text{H}_{40}\text{O}$ , EI-MS,  $^1\text{H}$  NMR, and  $^{13}\text{C}$  NMR data were identical to those reported for an authentic specimen of trans-phytol<sup>[11]</sup>.

**3(20)Phytene-1, 2-diol (8)**  $\text{C}_{20}\text{H}_{40}\text{O}_2$ , colorless oil, IR  $V_{\text{max}}$   $\text{cm}^{-1}$ : 3400, 1640 and 890; FAB+MS  $m/z$ : 313 [ $\text{M}+1$ ]<sup>+</sup> (18), 295 [ $\text{M}+1-\text{H}_2\text{O}$ ]<sup>+</sup> (18), 277 (6), 83 (32);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  4.98 (1H, s, H-20a), 5.13 (1H, s, H-20b), 4.21 (1H, dd,  $J=7.2, 3.2$  Hz, H-2), 3.71 (1H, dd,  $J=11.2, 3.2$  Hz, H-1a), 3.54 (1H, dd,  $J=11.2, 7.2$  Hz, H-1b), 0.87 (6H, d,  $J=6.8$  Hz, H-16 and H-17), 0.86 (3H, d,  $J=6.4$  Hz, H-19), 0.84 (3H, d,  $J=6.4$  Hz, H-18), 2.00 (2H, m, H-4), 1.60~1.00 (19H, m);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  65.6 (t, C-1), 75.0 (d, C-2), 148.6 (s, C-3), 33.0 (t, C-4), 25.4 (t, C-5), 37.2 (t, C-6), 32.7 (d, C-7), 37.4 (t, C-8), 24.4 (t, C-9), 37.4 (t, C-

10), 32.8 (d, C-11), 37.3 (t, C-12), 24.8 (t, C-13), 39.3 (t, C-14), 27.9 (d, C-15), 22.6 (q, C-16), 22.7 (q, C-17), 19.7 (q, C-18), 19.7 (q, C-19),

110.5 (t, C-20). These data were identical to those recorded for an authentic specimen of 3 (20) phytene-1,2-diol<sup>[11,12]</sup>.

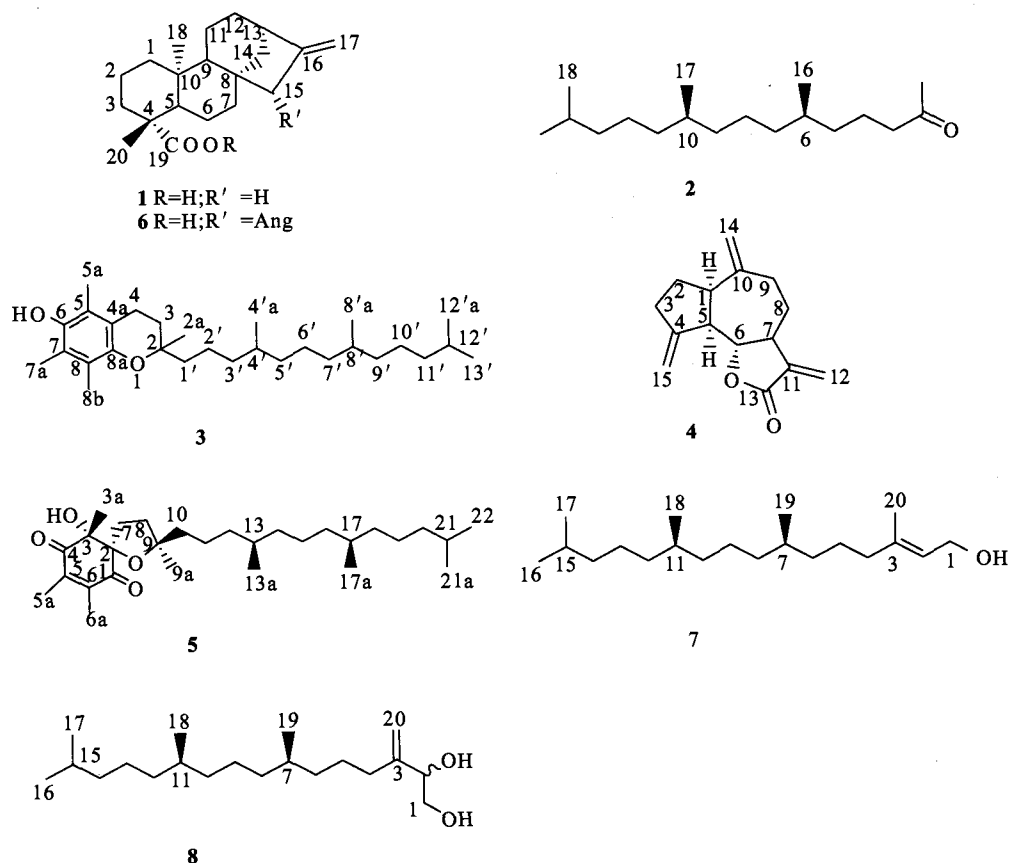


Fig. 1 Structures of compounds 1~8

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