

褐薄小齿菌的化学成分研究

杨小龙^{1,2}, 王飞^{1,2}, 邵红军^{1,2}, 董泽军¹, 丁智慧¹, 杨婉秋^{1,2}, 刘吉开^{1*}

¹中国科学院昆明植物研究所 植物化学与西部植物资源持续利用国家重点实验室, 昆明 650204;

²中国科学院研究生院, 北京 100049

摘要:从褐薄小齿菌(*Hydnellum concrescens*)子实体中分离得到9个已知化合物,借助光谱手段,它们的化学结构分别鉴定为:friedelin(1), (22E,24R)-麦角甾-5,7,22-三烯-3 β -醇(2), 5 α ,8 α -过氧-(22E,24R)-麦角甾-6,22-二烯-3 β -醇(3), (22E,24R)-麦角甾-5,22-二烯-3 β -羟基-7-酮(4), 6-甲氧基-cerevisterol(5), thelephantin K(6), thelephantin I(7), thelephantin J(8), thelephantin L(9)。

关键词:褐薄小齿菌;三萜;甾醇;对联三苯

中图分类号:R284.2;Q946

文献标识码:A

Study on the Chemical Constitutes of *Hydnellum concrescens*

YANG Xiao-long^{1,2}, WANG Fei^{1,2}, SHAO Hong-jun^{1,2}, DONG Ze-jun¹,

DING Zhi-hui¹, YANG Wan-qiu^{1,2}, LIU Ji-kai^{1*}

¹State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204, China;

²Graduate University of Chinese Academy of Sciences, Beijing 100049, China

Abstract: Nine known compounds have been isolated from the fruiting bodies of *Hydnellum concrescens*. Their structures were established as friedelin(1), (22E,24R)-ergosta-5,7,22-triene-3 β -ol(2), 5 α ,8 α -epidioxy-(22E,24R)-ergosta-6,22-dien-3 β -ol(3), (22E,24R)-ergosta-5,22-dien-3 β -hydroxyl-7-one(4), 6-methoxy-cerevisterol(5), thelephantin K(6), thelephantin I(7), thelephantin J(8), thelephantin L(9) by spectroscopic methods. All compounds are reported from this fungus for the first time.

Key words: *Hydnellum concrescens*; triterpenoid; sterols; p-terphenyls

Introduction

Hydnellum concrescens, a fungus belonging to the family Hydnaceae, is mainly distributed in the province of Zhejiang, Yunnan of China^[1]. As a part of our search for naturally occurring bioactive metabolites of the higher fungi in Yunnan Province, the chemical constituents of the fruiting bodies of *H. concrescens* were investigated and nine known compounds have been isolated, including friedelin(1), (22E,24R)-ergosta-5,7,22-triene-3 β -ol(2), 5 α ,8 α -epidioxy-(22E,24R)-ergosta-

6,22-dien-3 β -ol(3), (22E,24R)-ergosta-5,22-dien-3 β -hydroxyl-7-one(4), 6-methoxy-cerevisterol(5), thelephantin K(6), thelephantin I(7), thelephantin J(8), thelephantin L(9).

Experimental

General

NMR spectra were recorded on Bruker AV-400 and DRX-500 spectrometers, δ in ppm, J in Hz. IR spectra were obtained with a Bruker Tensor 27 FT-IR with KBr pellets. EI-MS spectra were recorded with a VG Auto-spec-3000 spectrometer, m/z (rel. int.). HR-ESI-MS was recorded with an API QSTAR Pulsar 1 spectrometer.

Material

Column chromatography was carried out on silica gel (200-300 mesh, Qingdao Marine Chemical Ltd., Qing-

Received August 31, 2006; Accepted November 15, 2006

Foundation Item: This work was supported by the National Natural Science Foundation of China (30470027 and 30225048) and Natural Science Foundation of Yunnan Province(2005C0052M and 2005C0011R).

* Corresponding author Tel: 86-871-5216327; E-mail address: jkliu@mail.kib.ac.cn

dao, China) and Sephadex LH-20 (Amersham Biosciences, Uppsala, Sweden).

Fungal material

The fresh fruiting bodies of *H. conrescens* were collect-

ed from Wuding of Yunnan Province in China, in August 2005. The voucher specimen was deposited at the herbarium of Kunming Institute of Botany, Chinese Academy of Sciences.

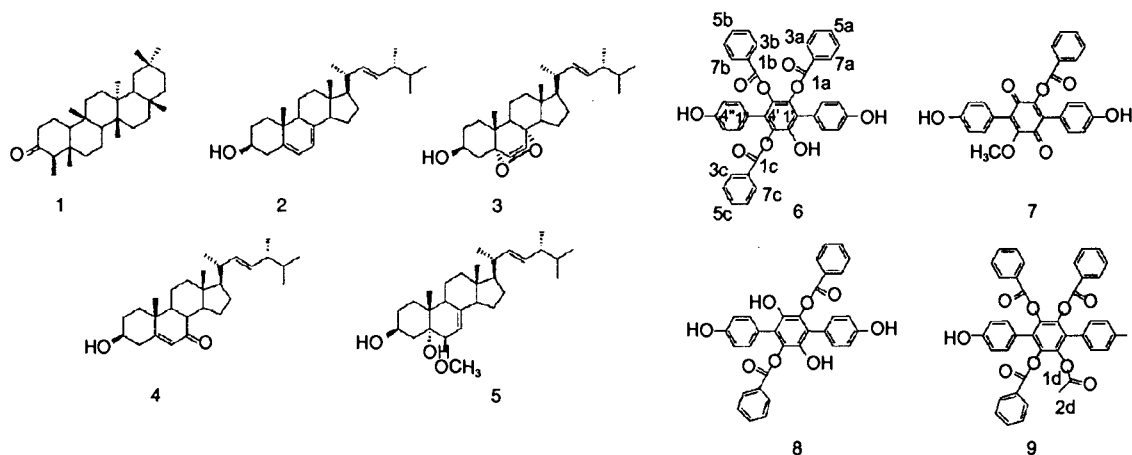


Fig. 1 Structures of compounds 1-9

Extraction and isolation

The fresh fruiting bodies of *H. conrescens* (dry wt. 570 g) were extracted three times with acetone and then three times with $\text{CHCl}_3/\text{MeOH}$ (1:1, v/v) at r. t. The combined extracts was concentrated *in vacuo* to give a deep brown gum (40 g), which was subjected to column chromatography (silica gel) with a gradient elution of $\text{CHCl}_3/\text{MeOH}$ (100:0, 98:2, 95:5, 90:10, 80:20, 50:50, 0:100 (v/v)), subsequently seven fractions (A-G) was obtained. Compounds **1** and **2** were obtained from fraction A as needles. Fraction B was rechromatographed over silica gel using petroleum ether/acetone (95:5) to afford compounds **3-5**. Fraction C was rechromatographed over silica gel using petroleum ether/acetone (5:1), Sephadex LH-20 ($\text{CHCl}_3/\text{MeOH}$, 1:1), Pre-TLC (petroleum ether/acetone, 3:2) and RP-8 (80% $\text{MeOH}-\text{H}_2\text{O}$) yielded compound **7**, further purified by repeated CC (silica gel; petroleum ether/acetone, 2:1) to afford compound **9**. Fraction D was further purified by repeated CC (silica gel; petroleum ether/acetone, 2:1) to afford compound **6**, then subjected to Sephadex LH-20 (Acetone) yielded compound **8**.

Results and Discussion

Friedelin (1) White needles (acetone), mp. 260-263

$^{\circ}\text{C}$, $[\alpha]_{\text{D}}^{20} -22.5^{\circ}\text{C}$ (c 1.00, CHCl_3); EI-MS m/z (%): 426 $[\text{M}]^+$ (15), 411 (8), 341 (6), 302 (16), 273 (38), 273 (38), 218 (31), 205 (42); $\text{IR}_{\text{vmax}} \text{cm}^{-1}$: 1702 (CO); $^1\text{H NMR}$ (500 MHz, CD_3COCD_3) δ : 2.27 (1H, m), 1.76-1.31 (22H, m), 1.27 (3H, s), 1.07 (3H, s), 1.03 (3H, s), 1.02 (3H, s), 0.97 (3H, s), 0.90 (3H, s), 0.89 (3H, s), 0.74 (3H, s); $^{13}\text{C NMR}$ (125 MHz, CD_3COCD_3) δ : 22.3 (t, C-1), 41.5 (t, C-2), 213.5 (s, C-3), 58.2 (d, C-4), 42.1 (s, C-5), 41.3 (t, C-6), 18.2 (t, C-7), 53.1 (d, C-8), 37.5 (s, C-9), 59.5 (d, C-10), 35.6 (t, C-11), 30.5 (t, C-12), 39.7 (t, C-13), 38.3 (s, C-14), 32.4 (t, C-15), 36.0 (t, C-16), 30.0 (s, C-17), 42.8 (d, C-18), 35.4 (t, C-19), 28.2 (s, C-20), 32.8 (t, C-21), 39.2 (t, C-22), 6.8 (q, C-23), 14.6 (q, C-24), 17.9 (q, C-25), 20.2 (q, C-26), 18.6 (q, C-27), 32.1 (q, C-28), 35.0 (q, C-29), 31.8 (q, C-30) [2].

(22E,24R)-Ergosta-5,7,22-trien-3 β -ol (2) Colorless needles (acetone), mp. 152-154 $^{\circ}\text{C}$, $[\alpha]_{\text{D}}^{20} -129^{\circ}\text{C}$ (c 0.22, CHCl_3); EI-MS m/z (%): 396 $[\text{M}]^+$ (10), 363 (12), 337 (6), 253 (5), 137 (100), 69 (42); $\text{IR}_{\text{vmax}} \text{cm}^{-1}$: 3340, 1610; $^1\text{H NMR}$ (400 MHz, CDCl_3) δ : 5.56 (1H, m), 5.18 (1H, m), 5.18 (2H, m, H-22, 23), 3.62 (1H, m, H-3), 1.02 (3H, d, $J = 6.8$ Hz, H-4), 0.91 (3H, s, H-19), 0.90 (3H, d, $J = 6.4$ Hz, H-

21), 0.78(3H, d, $J = 6.4$ Hz, H-26, 27), 0.67(3H, s, H-18); ^{13}C NMR (100 MHz, CDCl_3) δ : 38.4 (t, C-1), 32.0 (t, C-2), 70.5 (d, C-3), 40.8 (t, C-4), 139.8 (s, C-5), 119.6 (d, C-6), 116.3 (d, C-7), 141.3 (s, C-8), 46.3 (d, C-9), 37.1 (s, C-10), 21.1 (t, C-11), 39.1 (t, C-12), 42.8 (s, C-13), 54.6 (d, C-14), 23.0 (t, C-15), 28.3 (t, C-16), 55.8 (d, C-17), 12.1 (q, C-18), 16.3 (q, C-19), 40.4 (d, C-20), 21.1 (q, C-21), 135.6 (d, C-22), 132.0 (d, C-23), 40.8 (d, C-24), 33.1 (q, C-25), 20.0 (d, C-26), 19.6 (q, C-27), 17.6 (q, C-28)^[3].

5 α , 8 α -Epidioxy-(22E, 24R)-ergosta-6, 22-dien-3 β -ol (3) White needles (acetone), mp. 177-178 °C, $[\alpha]_{\text{D}}^{20} +20$ °C (c 0.10, CHCl_3); EI-MS m/z (%): 428 $[\text{M}]^+$ (13), 410 $[\text{M}-\text{H}_2\text{O}]^+$ (5), 396 $[\text{M}-\text{O}_2]^+$ (100), 363 (35), 303 (8), 251 (20), 152 (30), 107 (24), 95 (35), 81 (43), 69 (65); $\text{IR}_{\text{v}_{\text{max}}}$ cm^{-1} : 3525, 3309, 2955, 1650, 1380, 1074; ^1H NMR (400 MHz, CDCl_3) δ : 6.49(1H, d, $J = 8.5$ Hz), 6.23(1H, d, $J = 8.5$ Hz), 5.20(1H, dd, $J = 7.6, 7.5$ Hz), 5.12(1H, dd, $J = 7.6, 7.5$ Hz), 3.94(1H, m), 2.08-1.49(20H, m), 1.20(3H, s), 0.98(3H, d, $J = 6.6$ Hz), 0.88(3H, d, $J = 6.8$ Hz), 0.86(3H, s), 0.82(3H, d, $J = 3.6$ Hz), 0.79(3H, d, $J = 3.4$ Hz); ^{13}C NMR (100 MHz, CDCl_3) δ : 34.1 (t, C-1), 30.2 (t, C-2), 66.5 (d, C-3), 37.0 (t, C-4), 82.1 (s, C-5), 135.5 (d, C-6), 130.8 (d, C-7), 79.4 (s, C-8), 51.2 (d, C-9), 37.0 (s, C-10), 23.4 (t, C-11), 39.4 (t, C-12), 44.6 (s, C-13), 51.7 (d, C-14), 20.6 (t, C-15), 28.6 (t, C-16), 56.3 (d, C-17), 12.9 (q, C-18), 18.2 (q, C-19), 39.7 (d, C-20), 20.9 (q, C-21), 135.2 (d, C-22), 132.4 (d, C-23), 42.8 (d, C-24), 33.1 (q, C-25), 19.1 (d, C-26), 19.6 (q, C-27), 17.6 (q, C-28)^[4].

(22E, 24R)-Ergosta-5, 22-dien-3 β -hydroxy-7-one (4) White needles (acetone), mp. 170-171 °C, $[\alpha]_{\text{D}}^{20} -93$ °C (c 0.19, CHCl_3); EI-MS m/z (%): 412 $[\text{M}]^+$ (33), 397 (5), 379 (4), 369 (9), 351 (5), 314 (62), 287 (79), 285 (52), 269 (24), 245 (18), 205 (20), 191 (30), 161 (44), 107 (69), 69 (78), 55 (100); $\text{IR}_{\text{v}_{\text{max}}}$ cm^{-1} : 3392, 1672; ^1H NMR (400 MHz, CDCl_3) δ : 5.67(1H, d, H-6), 5.17(2H, m, H-22 and H-23), 3.65(1H, bm, Hax-3), 2.48(1H, m, Heq-4),

2.35(1H, m, Hax-4), 2.23(1H, dd, H-8), 1.94(1H, m, H-20), 1.82(1H, m, H-24), 1.44(1H, m, H-25), 1.17(3H, s, H-19), 0.99(3H, d, $J = 6.6$ Hz, H-21), 0.89(3H, d, $J = 6.8$ Hz, H-28), 0.81(3H, d, $J = 6.7$ Hz, H-26 or H-27), 0.79(3H, d, $J = 6.8$ Hz, H-27 or H-26), 0.67(3H, s, H-18). ^{13}C NMR (100 MHz, CDCl_3) δ : 36.3 (t, C-1), 31.0 (t, C-2), 70.4 (d, C-3), 41.8 (t, C-4), 165.6 (s, C-5), 125.9 (d, C-6), 202.6 (s, C-7), 45.3 (d, C-8), 50.0 (d, C-9), 38.5 (s, C-10), 21.1 (t, C-11), 38.5 (t, C-12), 43.0 (s, C-13), 49.9 (d, C-14), 26.3 (t, C-15), 28.6 (t, C-16), 54.7 (d, C-17), 12.2 (q, C-18), 17.3 (q, C-19), 39.9 (d, C-20), 21.1 (q, C-21), 135.6 (d, C-22), 131.9 (d, C-23), 42.8 (d, C-24), 33.1 (d, C-25), 17.6 (q, C-26), 19.6 (q, C-27), 20.0 (q, C-28)^[5].

6-Methoxy-cerevisterol (5) White needles (acetone), mp. 237-238 °C, $[\alpha]_{\text{D}}^{20} -104$ °C (c 0.15, CHCl_3); EI-MS m/z (%): 444 $[\text{M}]^+$ (2), 426 $[\text{M}-\text{H}_2\text{O}]^+$ (100), 411 (40), 393 (47), 377 (32), 355 (20), 337 (30), 283 (24), 269 (12), 251 (20); $\text{IR}_{\text{v}_{\text{max}}}$ cm^{-1} : 3540, 2955, 2812, 1465; ^1H NMR (400 MHz, CDCl_3) δ : 5.37(1H, dd, $J = 5.1, 2.6$ Hz, H-7), 5.14-5.23(2H, m, H-22, 23), 4.03(1H, m, H-3), 3.37(3H, s, CH_3), 3.14(1H, bd, $J = 5.1$ Hz, H-6), 1.20-2.10, 1.06(3H, s, H-19), 1.00(3H, d, $J = 6.6$ Hz, H-21), 0.89(3H, d, $J = 6.9$ Hz, H-28), 0.81(3H, d, $J = 6.6$ Hz, H-26), 0.80(3H, d, $J = 6.5$ Hz, H-27), 0.57(3H, s, H-18); ^{13}C NMR (100 MHz, CDCl_3) δ : 32.7 (C-1), 30.8 (C-2), 67.8 (C-3), 39.6 (C-4), 76.3 (C-5), 82.4 (C-6), 115.0 (C-7), 143.6 (C-8), 42.8 (C-9), 37.2 (C-10), 22.9 (C-11), 39.3 (C-12), 43.8 (C-13), 54.9 (C-14), 22.1 (C-15), 27.9 (C-16), 55.9 (C-17), 12.3 (C-18), 18.3 (C-19), 40.5 (C-20), 21.1 (C-21), 135.4 (C-22), 132.0 (C-23), 43.8 (C-24), 33.1 (C-25), 19.9 (C-26), 19.6 (C-27), 17.6 (C-28)^[6].

Thelephantin K (6) Grayish solid; Negative FAB-MS: 637 $[\text{M}-\text{H}]^-$; $\text{IR}_{\text{v}_{\text{max}}}$ cm^{-1} : 3447, 1745, 1611, 1526; $\text{UV}_{\lambda_{\text{max}}}^{\text{MeOH}}$ nm: 231, 260; ^1H NMR (500 MHz, $\text{DM}-\text{SO}-d_6$) δ : 9.50, 9.48, 9.42(3H, s, 3OH), 7.22(2H, d, $J = 8.4$ Hz, H-2, 6), 6.74(2H, d, $J = 8.4$ Hz, H-3, 5), 7.38(1H, d, $J = 8.4$ Hz, H-2''), 6.77(1H, d, $J = 8.4$ Hz, H-3''), 6.62(1H, d, $J = 8.4$ Hz, H-5''),

7.21(1H,d, $J = 8.4$ Hz,H-6''),7.76(2H,d, $J = 8.5$ Hz,H-3a,7a),7.28(overlap,H-4a,6a),7.44(1H,t, $J = 7.7$ Hz,H-5a),8.01(1H,d, $J = 8.5$ Hz,H-3b),7.47(2H,t, $J = 8.2$ Hz,H-4b,6b),7.59(1H,m,H-5b),7.97(1H,d, $J = 8.5$ Hz,H-7b),7.76(1H,d, $J = 8.5$ Hz,H-3c),7.28(overlap,H-4c),7.44(1H,t, $J = 7.7$ Hz,H-5c),7.28(overlap,H-6c),7.76(1H,d, $J = 8.5$ Hz,H-7c); ^{13}C NMR(125 MHz,DMSO- d_6) δ :124.9(s,C-1),132.6(d,C-2,6),116.0(d,C-3,5),158.0(s,C-4),124.1(s,C-1'),135.0(C-2''),134.8(s,C-3'),124.1(s,C-4'),135.0(s,C-5'),142.7(s,C-6'),124.9(s,C-1''),132.7(d,C-2''),116.1(d,C-3''),158.3(s,C-4''),115.9(d,C-5''),132.0(d,C-6''),166.4(s,C-1a),130.8(s,C-2a),130.7(d,C-3a,7a),129.5(d,C-4a,6a),134.6(d,C-5a),166.3(s,C-1b),129.8(s,C-2b),130.5(d,C-3b),129.5(d,C-4b,6b),134.2(d,C-5b),131.1(d,C-7b),166.4(s,C-1c),130.8(s,C-2c),130.7(d,C-3c),129.5(d,C-4c),134.6(d,C-5c),129.5(C-6c),130.7(d,C-7c)^[7,8].

Thelephantin I (7) Red powder; Negative FAB-MS: 441 [M-H]⁻; IR ν_{max} cm⁻¹: 3376, 1740, 1661, 1606, 1513; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 242, 260, 248; ^1H NMR(500 MHz, CD₃COCD₃) δ : 8.81, 8.68(2H,s,2OH), 7.36(2H,d, $J = 8.6$ Hz,H-2,6), 6.89(2H,d, $J = 8.6$ Hz H-3,5), 7.25(1H,d, $J = 8.6$ Hz,H-2'',6''), 6.86(1H,d, $J = 8.4$ Hz,H-3'',5''), 8.04(2H,d, $J = 7.3$ Hz,H-3a,7a), 7.55(2H,H-4a,6a), 7.71(1H,t, $J = 7.3$ Hz,H-5a), 3.93(3H,s,CH₃O); ^{13}C NMR(125 MHz, CD₃COCD₃) δ : 120.8(s,C-1), 132.8(d,C-2,6), 115.9(d,C-3,5), 159.5(s,C-4), 129.3(s,C-1'), 148.3(C-2'), 184.3(s,C-3'), 129.2(s,C-4'), 156.3(s,C-5'), 182.2(s,C-6'), 122.0(s,C-1''), 133.0(d,C-2''), 115.7(d,C-3''), 158.7(s,C-4''), 115.7(d,C-5''), 133.0(d,C-6''), 165.7(s,C-1a), 134.8(s,C-2a), 130.9(d,C-3a,7a), 129.7(d,C-4a,6a), 135.1(d,C-5a), 61.6(s,CH₃O)^[7,8].

Thelephantin J (8) Grayish solid; Negative FAB-MS: 533 [M-H]⁻; IR ν_{max} cm⁻¹: 3258, 1720, 1610, 1526; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 231, 262; ^1H NMR(400 MHz; CD₃OD) δ : 7.27(2H,d, $J = 8.8$ Hz,H-2,6), 6.75(2H,d, $J = 8.8$ Hz,H-3,5), 7.27(2H,d, $J = 8.8$ Hz,H-2'',6''), 6.75(1H,d, $J = 8.8$ Hz,H-3'',5''),

7.74(2H,dd, $J = 1.4,8.5$ Hz,H-3a,7a), 7.26(2H,t, $J = 8.5$ Hz,H-4a,6a), 7.43(1H,t, $J = 8.5$ Hz,H-5a), 7.74(2H,d, $J = 8.5$ Hz,H-3c,7c), 7.26(2H,t, $J = 8.5$ Hz,H-4c,6c), 7.43(1H,t, $J = 8.5$ Hz,H-5c); ^{13}C NMR(100 MHz, CD₃OD) δ : 124.5(s,C-1), 132.3(d,C-2,6), 115.8(d,C-3,5), 157.7(s,C-4), 124.3(s,C-1'), 132.3(C-2''), 141.9(s,C-3'), 124.3(s,C-4'), 132.3(s,C-5'), 141.9(s,C-6'), 124.5(s,C-1''), 132.3(d,C-2''), 115.8(d,C-3''), 157.7(s,C-4''), 115.9(d,C-5''), 132.3(d,C-6''), 166.9(s,C-1a), 130.3(s,C-2a), 130.4(d,C-3a,7a), 129.5(d,C-4a,6a), 134.7(d,C-5a), 165.0(s,C-1c), 130.2(s,C-2c), 130.7(d,C-3c), 129.5(d,C-4c), 134.7(d,C-5c), 129.5(d,C-6c), 130.4(d,C-7c)^[7,8].

Thelephantin L (9) Grayish solid; Positive FAB-MS: 703 [M + Na]⁺; IR ν_{max} cm⁻¹: 3452, 1748, 1611, 1525; UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm: 232, 267; ^1H NMR(500 MHz, CD₃OD) δ : 8.48, 8.38(2H,s,2OH), 7.22(2H,d, $J = 8.8$ Hz,H-2,6), 6.75(2H,d, $J = 8.8$ Hz H-3,5), 7.24(1H,d, $J = 8.8$ Hz,H-2'',6''), 6.65(1H,d, $J = 8.8$ Hz,H-3'',5''), 7.79(2H,d, $J = 8.2$ Hz,H-3a,7a), 7.28(t, $J = 8.2$ Hz,H-4a,6a), 7.47(1H,t, $J = 8.2$ Hz,H-5a), 7.79(1H,d, $J = 8.2$ Hz,H-3b), 7.28(2H,t, $J = 8.2$ Hz,H-4b,6b), 7.47(1H,t, $J = 8.2$ Hz,H-5b), 7.79(1H,d, $J = 8.2$ Hz,H-7b), 7.97(1H,d, $J = 8.5$ Hz,H-3c), 7.49(1H,t, $J = 8.5$ Hz,H-4c), 7.64(1H,t, $J = 8.5$ Hz,H-5c), 7.49(1H,t, $J = 8.5$ Hz,H-6c), 7.97(1H,d, $J = 8.5$ Hz,H-7c), 2.00(3H,s,CH₃CO); ^{13}C NMR(125 MHz, CD₃OD) δ : 123.7(s,C-1), 131.7(d,C-2,6), 116.0(d,C-3,5), 158.0(s,C-4), 131.8(s,C-1'), 140.9(C-2'), 140.9(s,C-3'), 131.8(s,C-4'), 140.9(s,C-5'), 140.9(s,C-6'), 123.7(s,C-1''), 131.7(d,C-2''), 116.0(d,C-3''), 158.0(s,C-4''), 116.0(d,C-5''), 131.7(d,C-6''), 164.9(s,C-1a), 129.3(s,C-2a), 130.8(d,C-3a,7a), 129.6(d,C-4a,6a), 134.4(d,C-5a), 164.9(s,C-1b), 129.3(s,C-2b), 130.8(d,C-3b), 129.6(d,C-4b,6b), 134.4(d,C-5b), 130.8(d,C-7b), 164.9(s,C-1c), 129.3(s,C-2c), 131.4(d,C-3c), 130.3(d,C-4c), 135.3(d,C-5c), 129.6(C-6c), 130.8(d,C-7c), 169.6(s,C-1d), 20.0(q,C-2d)^[7,8].

Acknowledgements This project was supported by

the National Natural Science Foundation of China (30470027 and 30225048) and the Natural Science Foundation of Yunnan Province (2005C0052M) and (2005C0011R).

References

- 1 Hashimoto T, Quang DN, Kuratsune M, *et al.* Hydnellins A and B, nitrogen- containing terphenyls from the mushrooms *Hydnellum suaveolens* and *Hydnellum geogerium*. *Chem Pharm Bull*, 2006, **54**:912-914.
- 2 Akihisa T, Yamamoto K, Tamura T, *et al.* Triterpenoid ketone from *Lingnania chungii* McClure: arborinone, friedelin and glutinone. *Chem Pharm Bull*, 1992, **40**:789-791.
- 3 Mishra PD, Wahidulla S, Souza LD, *et al.* Lipid constituents of marine sponge *Suberites carnosus*. *Ind J Chem*, 1996, **35B**: 806-809.
- 4 Ma WG, Li CX, Wang DZ, *et al.* Ergosterol peroxides from *cryptoporus volvatus*. *Acta Bot Yunnan*, 1994, **16**:196-200.
- 5 Notaro G, Piccialli V, Sica D. New steroidal hydroxyketones and closely related diols from the marine sponge *Cliona copiosa*. *J Nat Prod*, 1992, **55**:1588-1594.
- 6 Hu L. Study on chemical constitutes from twelve species of higher fungi in Yunnan. Kunming Institute of Botany, Chinese Academy of Sciences, Ph. D. thesis, 2002. 63-64.
- 7 Quang DN, Hashimoto T, Hitaka Y, *et al.* Thelephantins I-N: p-terphenyl derivatives from the inedible mushroom *Hydnellum caeruleum*. *Phytochemistry*, 2004, **65**:1179-1184.
- 8 Liu JK. Natural terphenyls developments since 1877. *Chem Rev*, 2006, **106**:2209-2223.