

# New Cytotoxic Cycloartane Triterpenes from the Aerial Parts of *Actaea heracleifolia* (syn. *Cimicifuga heracleifolia*)

## Authors

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## Key words

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## Bibliography

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## ABSTRACT

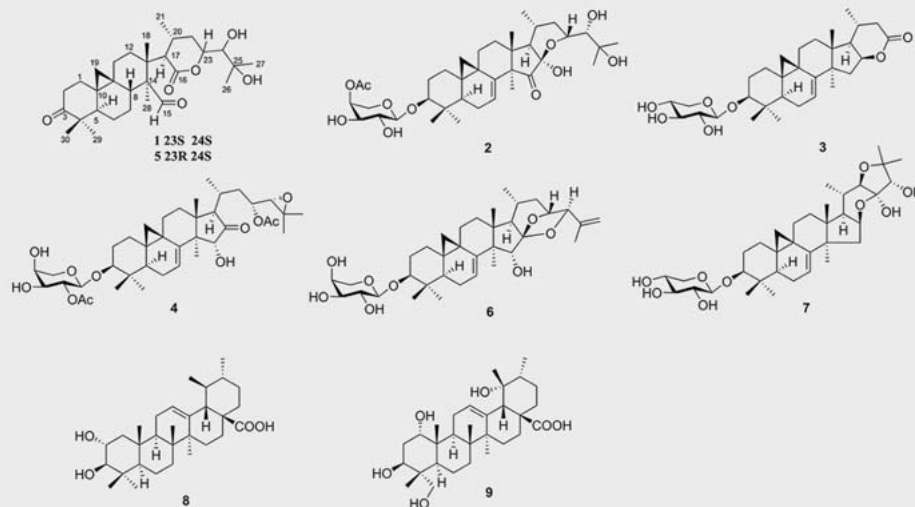
One new 15,16-seco-cycloartane triterpene (1), three new cycloartane triterpene glycosides (2–4), and five known compounds (5–9) were isolated from the aerial parts of *Actaea heracleifolia*. The chemical structures of these compounds were determined on the basis of NMR analysis, HRTOF-ESIMS data, and other spectroscopic methods. Selected compounds were evaluated for their cytotoxicity against human tumor cell lines (HL-60, SMMC-7721, A549, MCF-7, and SW480) *in vitro*. Compounds 3 and 4 showed weak activity against the HL-60, A-549, and MCF-7 cell lines with IC<sub>50</sub> values ranging from 21.34 to 36.98 μM.

## Introduction

The *Actaea* species, belonging to the family Ranunculaceae, has a long history of uses as a medicinal herb worldwide. In China, the rhizomes of *Actaea heracleifolia* Kom., *Actaea dahurica* (Turcz.) Maxim., and *Actaea foetida* L., officially listed in the Chinese Pharmacopoeia with the name “shengma”, are used as cooling and detoxifying agents for the treatment of headache, sore throat, and toothache [1]. In Europe and the United States, *Actaea racemosa* (L.) Nutt., commonly called black cohosh, is also used as a dietary supplement for women’s health during climacteric periods [2, 3]. Triterpene glycosides have been considered the main active components of *Actaea* species, and phytochemical investigations of plant species led to the isolation of a series of 9,19-cycloartane triterpenes characterized with unique structural features [4], including ring A opened cycloartane triterpenes, such as 3,4-seco-4-hydroxy-3-cimigenolate [5]; ring A expanded cimigenol-type triterpenes, such as cimihaclein A [6]; 15,16-seco-

cycloartane triterpenes [5–10]; ring B opened triterpenes [6, 11, 12]; and cycloartane triterpenes with new skeletons, such as cimicifugadine [13], cimicifine B [14], and cimiyunnins A–D [15]. In a further investigation of the aerial parts of *A. heracleifolia*, we found four new cycloartane triterpenes (1–4) and five known ones: cimihaclein D (5) [6], 25-anhydrocimigenol-3-O-α-L-arabinopyranoside (6) [16], cimiaceroside A (7) [17], 2α-hydroxyursolic acid (8) [18], and 1α,3β,19α,23-tetrahydroxyurs-12-en-28-oic acid-28-O-β-D-xylopyranoside (9) [19] (► **Fig. 1**). Compound 1 was a D ring-cleaved 15,16-seco-cycloartane triterpene. Compounds 2–4 were 9,19-cycloartane triterpenoid saponins with a fused monosaccharide moiety. Herein, we reported the structure determination by 1D/2D NMR analysis of the new compounds and the evaluation of the cytotoxic activity of selected compounds against the HL-60, SMMC-7721, A549, MCF-7, and SW480 cell lines.

\* These authors contributed equally to the work reported in this article.



► **Fig. 1** Chemical structures of compounds 1–9.

## Results and Discussion

Compound **1** was obtained as white powder. The HRTOF-ESIMS ion signal at  $m/z$  525.3180  $[M + Na]^+$  (calcd for 525.3192) indicated the molecular formula  $C_{30}H_{46}O_6$  with eight degrees of unsaturation. Its IR spectra showed the presence of hydroxy ( $3438\text{ cm}^{-1}$ ) and carbonyl functional groups ( $1707\text{ cm}^{-1}$ ). The  $^1\text{H}$  and  $^{13}\text{C}$  NMR data for **1** (► **Table 1**) were similar to those of cimiheraclen D (**5**) [6], and the slight difference indicated that compound **1** was a configurational isomer of **5**, which was confirmed by its 2D NMR spectra (HSQC,  $^1\text{H}$ - $^1\text{H}$  COSY, and HMBC) (► **Fig. 2**). The ROESY correlations of H-23 to H-20 and of H-20 to Me-18 suggested a 23R configuration in compound **5**, while the correlations of H-23 to H-17 and of H-17 to Me-28 suggested a 23S configuration in compound **1** (► **Fig. 3**). In addition, the configuration of C-24 in compound **5** was proposed to be *S* by comparison of the chemical shifts and coupling constants of H-23 (5.07, d,  $J = 11.2$ ) and H-24 (3.76, s) of **5** with 23R, 24S configuration analogue [H-23 (5.01, d,  $J = 11.1$  Hz), H-24 (3.70, s)] [8]. The configuration at C-24 in **1** was finally confirmed by molecular modeling, in which  $\theta = 73.8^\circ$  in 23R 24S configuration,  $\theta = 177.7^\circ$  in 23R 24R configuration,  $\theta = 168.9^\circ$  in 23S 24R configuration, and  $\theta = 61.1^\circ$  in 23S 24S configuration (**Fig. 10S**, Supporting Information), the coupling constants of H-23 and H-24 were 10.9 Hz and 0 Hz, so the configuration of C-24 in **1** was proposed as *S* based on the function of dihedral angle and  $^3J_{\text{H-C-C-H}}$ . Also, this was consistent with other 15,16-seco-cycloartane derivatives reported previously [5–10]. Therefore, the structure of **1** was established as 15,16-seco-14-formyl-(23S, 24S)-16-oxohydroshengmanol-3-one (► **Fig. 1**) and given the name cimiheraclen E.

Compound **2** possessed a molecular formula of  $C_{37}H_{56}O_{11}$  based on its HRTOF-ESIMS ion signal at  $m/z$  699.3715  $[M + Na]^+$  (calcd for 699.3720). Its  $^1\text{H}$  NMR spectrum displayed resonances for cyclopropane methylene protons at  $\delta_{\text{H}}$  0.50 (1H, d,  $J = 4.0$  Hz) and 1.03 (1H, d,  $J = 4.0$  Hz) for  $\text{CH}_2$ -19, six tertiary methyl singlets

at  $\delta_{\text{H}}$  1.06, 1.27, 1.35, 1.61, 1.62, and 1.64 (each 3H, s), and one secondary methyl at  $\delta_{\text{H}}$  1.14 (3H, d,  $J = 6.3$  Hz) as well as an anomeric proton signal at  $\delta_{\text{H}}$  4.86 (1H, d,  $J = 7.6$  Hz), which is characteristic of a 9,19-cyclolanostane-type triterpene glycoside. The sugar obtained after acid hydrolysis was identified as L-arabinose by comparing its TLC mobility and specific rotation with those of a standard. Detailed analysis of its NMR data (► **Table 2**) indicated that **2** is a 16 $\alpha$ -hydroxyl dahurinol-type triterpene glycoside and is similar to cimidahuside C [20]. The major difference was the acetyl substituent. The observed  $^1\text{H}$ - $^1\text{H}$  COSY correlation of  $\delta_{\text{H}}$  4.28 (1H, m, H-5') to  $\delta_{\text{H}}$  5.26 (1H, brs, H-4') and the HMBC correlation between  $\delta_{\text{H}}$  5.26 (1H, brs, H-4') and  $\delta_{\text{H}}$  171.2 (-OAc) indicated the acetyl group was located at C-4' of the sugar moiety (► **Fig. 2**). The significant ROESY associations (► **Fig. 3**) of H-3/H-5 and H-20/H-17 suggested a 3S, 23R configuration. The hydroxyl group at C-24 was confirmed as *S* by comparison of the chemical shifts and coupling constant of **2** with those of cimidahuside C [20]. Accordingly, the structure of **2** was determined as 16 $\alpha$ -hydroxyl-7(8)-en-dahurinol-3-O-[4'-O-acetyl]- $\alpha$ -L-arabinopyranoside (► **Fig. 1**) and named as cimiheraclen F.

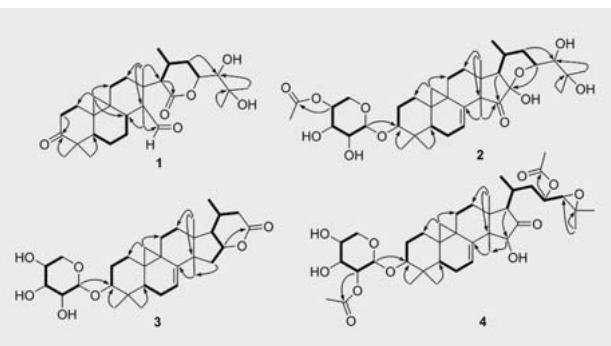
Compound **3** was isolated as a white powder and found to have the molecular formula  $C_{31}H_{46}O_7$  on the basis of the HRTOF-ESIMS ion peak at  $m/z$  553.3138  $[M + Na]^+$  (calcd for 553.3141). The NMR data (► **Table 2**) of **3** were very similar to those of 3 $\beta$ ,11 $\beta$ -dihydroxy-24,25,26,27-tetranor-cycloart-7-en-23,16 $\beta$ -olide 3-O- $\beta$ -D-xylopyranoside [21] except for the absence of the hydroxy group at C-11. The stereochemistry of **3** was determined from its ROESY spectrum (► **Fig. 3**). The crosspeaks of H-3 ( $\delta_{\text{H}}$  3.50, 1H, dd,  $J = 11.7, 4.1$  Hz) with H-5 ( $\delta_{\text{H}}$  1.26, 1H, dd,  $J = 12.5, 5.1$  Hz) and H-16 ( $\delta_{\text{H}}$  4.88, 1H, m) with H-17 ( $\delta_{\text{H}}$  1.92, 1H) and  $\text{CH}_3$ -28 ( $\delta_{\text{H}}$  1.06, 3H, s) indicated the  $\beta$ -orientation of the substituents at C-3 and C-16. Therefore, **3** was elucidated as 3 $\beta$ -hydroxy-24, 25, 26, 27-tetranor-cycloart-7(8)-en-23,16 $\beta$ -olide 3-O- $\beta$ -D-xylopyranoside (► **Fig. 1**) and given the name cimiheraclen G.

► **Table 1** The NMR data of compounds 1 and 5 ( $\delta$  in ppm).

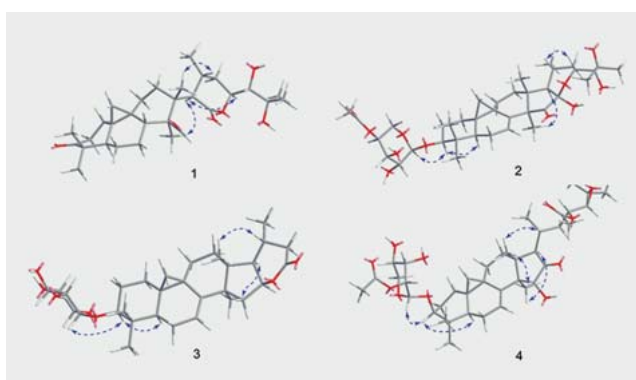
Position	1	5	Position	1	5
	$\delta_H^b$	$\delta_H^b$		$\delta_C^c$	$\delta_C^c$
1	1.81 <sup>a</sup> 1.51 <sup>a</sup>	1.82 <sup>a</sup> 1.53 <sup>a</sup>	1	32.2 t	32.2 t
2	2.73 m 2.37 brd	2.72 m 2.38 brd	2	37.4 t	37.4 t
3			3	215.2 s	215.1 s
4			4	50.3 s	50.2 s
5	1.82 <sup>a</sup>	1.81 <sup>a</sup>	5	45.4 d	45.4 d
6	1.36 <sup>a</sup> 1.01 <sup>a</sup>	1.38 <sup>a</sup> 1.01 <sup>a</sup>	6	19.2 t	19.2 t
7	1.42 <sup>a</sup> 1.16 m	1.39 <sup>a</sup> 1.16 m	7	22.0 t	22.0 t
8	2.47 m	2.44 dd (5.2 7.4)	8	39.2 d	38.9 d
9			9	22.1 s	22.0 s
10			10	25.7 s	25.7 s
11	1.64 <sup>a</sup> 1.54 <sup>a</sup>	1.64 <sup>a</sup> 1.54 <sup>a</sup>	11	27.2 t	27.1 t
12	1.62 <sup>a</sup> 1.41 <sup>a</sup>	1.62 <sup>a</sup> 1.41 m	12	32.7 t	32.7 t
13			13	47.9 s	47.5 s
14			14	55.2 s	55.3 s
15	9.97 s	9.94 s	15	208.0 s	207.6 s
16			16	175.5 s	175.3 s
17	2.76 d (5.5)	2.74 d (5.6)	17	55.8 d	55.5 d
18	1.56 s	1.56 s	18	18.4 q	18.7 q
19	0.80 d (4.7) 0.24 d (4.7)	0.80 d (4.7) 0.24 d (4.7)	19	21.9 t	21.9 t
20	2.05 <sup>a</sup>	2.06 <sup>a</sup>	20	28.8 d	28.8 d
21	1.02 d (6.4)	0.99 d (6.2)	21	25.7 q	25.1 q
22	2.57 m 1.82 <sup>a</sup>	2.07 <sup>a</sup> 1.90 m	22	35.0 t	37.0 t
23	5.02 d (10.9)	5.07 d (11.2)	23	80.9 d	78.9 d
24	4.00 s	3.76 s	24	79.1 d	80.2 d
25			25	72.2 s	72.7 s
26	1.59 s	1.67 s	26	27.7 q	26.2 q
27	1.62 s	1.73 s	27	28.0 q	29.5 q
28	1.53 s	1.51 s	28	14.8 q	14.6 q
29	1.01 s	1.01 s	29	20.6 q	20.6 q
30	1.08 s	1.08 s	30	22.6 q	22.5 q

<sup>a</sup> Signals overlapped. <sup>b</sup> Recorded at 600 MHz in Pyridine-*d*<sub>5</sub>. <sup>c</sup> Recorded at 150 MHz in Pyridine-*d*<sub>5</sub>.

Compound 4 was also obtained as a white powder, and its molecular formula was C<sub>39</sub>H<sub>58</sub>O<sub>11</sub> based on its HRTOF-ESIMS ion signal at *m/z* 725.3877 [M + Na]<sup>+</sup> (calcd for 725.3877), which corresponds to 11 degrees of unsaturation. The NMR spectrum of 4 clearly displayed the signals characteristic of a 9,19-cycloartane-type triterpene. Direct analysis of its NMR data (► **Table 2**) indicated that compound 4 resembles a 23-*O*-acetyl-7,8-didehy-



► **Fig. 2** Major HMBC (→) and <sup>1</sup>H-<sup>1</sup>H COSY (---) correlations of compounds 1–4.



► **Fig. 3** Key ROESY correlations of compounds 1–4.

droshengmanol-3-*O*- $\alpha$ -L-arabinopyranoside [22] except for the presence of one additional acetyl group. The location of the acetyl group at C-2' (► **Fig. 2**) was identified by the HMBC correlation between H-2' ( $\delta_H$  5.58, 1H, t, *J* = 8.3 Hz) and the carbonyl signal at  $\delta_C$  170.4. The similarity between the chemical shifts and coupling constants of C-23 and C-24 in compound 4 with those of 23-*O*-diacetyl-7,8-didehydroshengmanol-3-*O*- $\alpha$ -L-arabinopyranoside indicated the configurations of C-23 and C-24 were *R* and *S*, respectively. Finally, the structure of 4 was confirmed as 23-*O*-acetyl-7(8)-en-shengmanol-3-*O*-[2'-*O*-acetyl]- $\alpha$ -L-arabinopyranoside, as shown (► **Fig. 1**).

The new compounds (1–4) were evaluated for their cytotoxicities against HL-60, SMMC-7721, A549, MCF-7, and SW480 cell lines (► **Table 3**). Compounds 1 and 2 did not show cytotoxic activity with IC<sub>50</sub> value > 40  $\mu$ M. Compound 3 showed weak activity against A549 and MCF-7 cell lines with IC<sub>50</sub> value 27.75 and 22.45  $\mu$ M, respectively. Compound 4 also showed antitumor activity against the HL-60, A549, and MCF-7 cell lines with IC<sub>50</sub> value 26.54, 36.98, and 21.34  $\mu$ M, respectively.

The NMR, IR, UV, and HRTOF-ESIMS spectra of compounds 1–5, as well as the dose-response curves of cytotoxic activity, are available as Supporting Information.

► **Table 2** The NMR data of compounds 2–4 ( $\delta$  in ppm,  $J$  in Hz).

Position	2		3		4	
	$\delta_{\text{H}}^b$	$\delta_{\text{C}}^c$	$\delta_{\text{H}}^b$	$\delta_{\text{C}}^c$	$\delta_{\text{H}}^b$	$\delta_{\text{C}}^c$
1	1.78 m 1.23 m	30.7 t	1.66 m 1.33 m	30.7 t	1.51 m 1.37 m	32.4 t
2	2.33 m 1.98 <sup>a</sup>	29.8 t	2.35 m 1.97 m	29.7 t	2.27 m 1.91 m	29.7 t
3	3.50 dd (11.6 4.0)	88.8 d	3.50 dd (11.7 4.1)	88.5 d	3.39 dd (11.4 4.1)	88.6 d
4		40.7 s		40.7 s		40.4 s
5	1.27 m	42.2 d	1.26 dd (12.5,5.1)	43.0 d	1.31 <sup>a</sup>	42.8 d
6	2.00 <sup>a</sup> 1.64 <sup>a</sup>	22.1 t	1.87 m 1.55 <sup>a</sup>	22.2 t	1.97 m 1.62 m	22.0 t
7	6.70 d (6.3)	117.3 d	5.10 d (6.8)	114.2 d	6.14 d (7.2)	115.3 d
8		142.6 s		149.2 s		147.6 s
9		21.8 s		21.3 s		21.8 s
10		28.8 s		28.7 s		28.7 s
11	2.12 m 1.04 m	25.1 t	2.06 m 1.15 m	25.3 t	2.20 m 1.76 m	25.5 t
12	1.91 m 1.45 m	31.6 t	1.63 m 1.55 <sup>a</sup>	32.2 t	1.80 m	33.8 t
13		40.7 s		44.4 s		41.2 s
14		54.2 s		50.5 s		49.8 s
15		212.5 s	2.23 m 1.93 m	42.9 t	4.61 s	81.2 d
16		96.6 s	4.88 <sup>a</sup>	81.1 d		220.8 s
17	2.17 m	57.8 d	1.92 <sup>a</sup>	54.9 d	2.33 d (7.8)	60.4 d
18	1.27 s	23.3 q	1.07 s	23.1 q	1.44 s	19.7 q
19	1.03 <sup>a</sup> 0.50 d (4.0)	28.1 t	1.07 <sup>a</sup> 0.47 d (3.9)	28.7 t	0.86 d (4.0) 0.50 d (4.0)	28.1 t
20	1.97 m	26.7 d	1.93 <sup>a</sup>	27.7 d	2.17 m	28.8 d
21	1.14 d (7.1)	23.4 q	0.99 d (5.9)	21.5 q	1.25 d (6.5)	20.1 q
22	2.45 m 1.69 m	33.0 t	2.46 dd (14.7 2.5) 2.28 dd (14.7 12.9)	39.1 t	2.91 m 1.74 m	37.8 t
23	4.66 dd (11.7 5.6)	77.0 d		174.3 s	5.45 m	72.3 d
24	3.62 brs	78.9 d			3.08 d (8.4)	65.6 d
25		73.3 s				58.9 s
26	1.61 s	26.8 q			1.28 s	25.1 q
27	1.62 s	29.0 q			1.45 s	19.2 q
28	1.64 s	25.0 q	1.06 s	27.1 q	1.30 s	22.1 q
29	1.35 s	26.0 q	1.37 s	26.1 q	1.13 s	25.8 q
30	1.06 s	14.5 q	1.08 s	14.6 q	1.02 s	14.4 q
1'	4.81 d (7.3)	107.9 d	4.88 <sup>a</sup>	107.9 d	4.85 d (7.9)	104.9 d
2'	4.42 t (9.0)	73.6 d	4.08 m	75.9 d	5.58 t (8.3)	75.9 d
3'	4.29 m	72.7	4.19 t (8.8)	79.0 d	4.22 <sup>a</sup>	76.6 d
4'	5.62 brs	72.7	4.27 m	71.6 d	4.23 <sup>a</sup>	71.7 d
5'	4.28 <sup>a</sup> 3.85 m	64.8 t	4.40 dd (11.2 5.2) 3.77 m	67.5 t	4.35 m 3.74 m	67.5 t
4'-OAc	2.01 s s	171.2 s 21.5 q				
23-OAc					2.06 s	171.0 s 21.3 q
2'-OAc					2.19 s	170.4 s 21.6 q

<sup>a</sup> Signals overlapped. <sup>b</sup> Recorded at 600 MHz in Pyridine-*d*<sub>5</sub>. <sup>c</sup> Recorded at 150 MHz in Pyridine-*d*<sub>5</sub>.

► **Table 3** Cytotoxic activities ( $IC_{50}$ ,  $\mu M$ ) of compounds 1–4 against five human cancer cell lines.<sup>a</sup>

Compounds	HL-60	SMMC-7721	A549	MCF-7	SW480
1	>40	>40	>40	>40	>40
2	>40	>40	>40	>40	>40
3	>40	>40	27.75 (26.44–29.06)	22.45 (21.25–23.65)	>40
4	26.54 (25.51–27.57)	>40	36.98 (35.88–38.08)	21.34 (20.37–22.31)	>40
DDP	1.24 (1.19–1.29)	7.14 (7.08–7.20)	6.30 (6.23–6.37)	17.65 (16.25–19.05)	13.50 (12.40–14.60)

<sup>a</sup> DDP (cisplatin) was used as a positive control. All data are present as the mean of  $IC_{50}$  values with lower and upper 95% CI from triplicate measurement ( $n = 3$ ).

## Materials and Methods

### General experimental procedures

Optical rotations were recorded on a Horiba SEPA-300 polarimeter. UV spectra were acquired on a Shimadzu UV-2401A instrument. IR spectra were collected on Bruker Tensor 27 FTIR spectrometers with KBr pellets. NMR spectra were recorded on Bruker Avance III-600 spectrometers with tetramethylsilane as an internal standard at room temperature. HRTOF-ESIMS were recorded on an Agilent G6230 TOF spectrometer. TLC was performed on precoated TLC plates (200–250  $\mu m$  thickness, silica gel 60 F<sub>254</sub>, Qingdao Marine Chemical Inc.), and the spots were visualized by heating after spraying with 10% aqueous H<sub>2</sub>SO<sub>4</sub>. Semi-preparative HPLC was performed on an Agilent 1100 liquid chromatograph with a Zorbax SB-C18 (5  $\mu m$ , 9.4 mm  $\times$  250 mm, 3 mL/min) column.

### Plant material

The aerial parts of *A. heracleifolia* were collected from Yichun County, Heilongjiang Province, China, in September 2012 and were identified by Prof. Zongyu Wang, Kunming Institute of Botany, Chinese Academy of Sciences. A voucher specimen (ZDSQ20120901) has been deposited at the State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, China.

### Extraction and isolation

The air-dried and powdered aerial parts of *A. heracleifolia* (10 kg) were extracted with MeOH (3  $\times$  30 L  $\times$  24 h) at room temperature to give a residue (265 g) after concentration under vacuum at 50 °C. The extract was subjected to silica gel CC (200–300 mesh, 3 kg, 20  $\times$  150 cm) and eluted with CHCl<sub>3</sub>-MeOH [100:0 (2 L), 80:1 (5 L), 50:1 (5 L), 20:1 (8 L), and 10:1 (8 L)] to afford fractions A (3 g), B (5 g), C (2.5 g), D (120 g), and E (10 g). Fraction B was divided into four sub-fractions (B.1–B.4) by RP-18 CC (20–45  $\mu m$ , 250 g, 5  $\times$  50 cm) eluted with MeOH-H<sub>2</sub>O (gradient from 50:50 to 100:0, 20 L). Fraction B.3 (2.5 g) was subjected to repeated silica gel columns (200–300 mesh, 50 g, 5  $\times$  25 cm) eluting with CHCl<sub>3</sub>-Me<sub>2</sub>CO (gradient from 20:1 to 5:1, 5 L) and then purified by semi-preparative HPLC (eluting with CH<sub>3</sub>CN-H<sub>2</sub>O, gradient from 50:50 to 75:25, 3.0 mL/min, 40 min, 210 nm) to give compounds 1 (2 mg), 5 (4 mg), 8 (4 mg), and 9 (2 mg). Com-

pounds 2 (9 mg), 3 (4 mg), 4 (8 mg), 6 (10 mg), and 7 (7 mg) were isolated from fraction C by silica gel CC (200–300 mesh, 50 g, 5  $\times$  25 cm) eluted with CHCl<sub>3</sub>-Me<sub>2</sub>CO (15:1 to 5:1, 5 L) followed by repeated semi-preparative HPLC (eluted with CH<sub>3</sub>CN-H<sub>2</sub>O, gradient from 50:50 to 75:25, 3.0 mL/min, 40 min, 210 nm).

### Physicochemical properties of 1–4

*Cimihaclein E* (1): white powder;  $[\alpha]_D^{25} = -69.25$  ( $c$  0.08, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 204 (3.89) nm; IR (KBr)  $\nu_{max}$  3440, 2942, 2871, 1700, 1632, 1457, 1380, 1032, 977  $cm^{-1}$ ; <sup>1</sup>H and <sup>13</sup>C NMR data (C<sub>5</sub>D<sub>5</sub>N) see ► **Table 1**; HRTOF-ESIMS  $m/z$  525.3180 [M + Na]<sup>+</sup> (calcd for C<sub>30</sub>H<sub>46</sub>O<sub>6</sub>Na, 525.3192).

*Cimihaclein F* (2): white powder;  $[\alpha]_D^{25} = -10.49$  ( $c$  0.15, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 204 (3.78), 268 (3.07) nm; IR (KBr)  $\nu_{max}$  3440, 2964, 2872, 1734, 1632, 1456, 1383, 1055, 1001  $cm^{-1}$ ; <sup>1</sup>H and <sup>13</sup>C NMR data (C<sub>5</sub>D<sub>5</sub>N) see ► **Table 2**; HRTOF-ESIMS  $m/z$  699.3715 [M + Na]<sup>+</sup> (calcd for C<sub>37</sub>H<sub>56</sub>O<sub>11</sub>Na, 699.3720).

*Cimihaclein G* (3): white powder;  $[\alpha]_D^{25} = -80.57$  ( $c$  0.07, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 203 (4.00) nm; IR (KBr)  $\nu_{max}$  3440, 2964, 2869, 1721, 1631, 1455, 1382, 1045, 969  $cm^{-1}$ ; <sup>1</sup>H and <sup>13</sup>C NMR data (C<sub>5</sub>D<sub>5</sub>N) see ► **Table 2**; HRTOF-ESIMS  $m/z$  553.3138 [M + Na]<sup>+</sup> (calcd for C<sub>31</sub>H<sub>46</sub>O<sub>7</sub>Na, 553.3141).

23-O-Acetyl-7(8)-en-shengmanol-3-O-[2'-O-acetyl]- $\alpha$ -L-arabinopyranoside (4): white powder;  $[\alpha]_D^{25} = -49.33$  ( $c$  0.08, MeOH); UV (MeOH)  $\lambda_{max}$  (log  $\epsilon$ ) 204 (3.96) nm; IR (KBr)  $\nu_{max}$  3434, 2926, 2853, 1738, 1631, 1459, 1379, 1044, 989  $cm^{-1}$ ; <sup>1</sup>H and <sup>13</sup>C NMR data (C<sub>5</sub>D<sub>5</sub>N) see ► **Table 2**; HRTOF-ESIMS  $m/z$  725.3877 [M + Na]<sup>+</sup> (calcd for C<sub>39</sub>H<sub>58</sub>O<sub>11</sub>Na, 725.3877)

### Hydrolysis and identification of the sugar moieties in compounds 2 and 4

Compounds 2 and 4 (2.5 mg of each) were dissolved in methanol (3 mL) and refluxed with 1.0 N HCl (2 mL) at 90 °C for 2 h. After neutralizing with 1.0 N NaOH, the reaction mixtures were extracted with CHCl<sub>3</sub>, and the aqueous layers were concentrated under reduced pressure to give the monosaccharides, which had  $R_f$  values (EtOAc/CHCl<sub>3</sub>/MeOH/H<sub>2</sub>O, 3:2:2:1) and specific rotations ( $[\alpha]_D^{20} + 82.78$ ,  $c$  0.05, MeOH) that were consistent with those of L-arabinopyranose (Sigma-Aldrich).

## Biological assays

Cytotoxic activity was investigated using five human cancer cell lines, human leukemia HL-60, hepatocellular carcinoma SMMC-7721, lung cancer A549, breast cancer MCF-7, and colon cancer SW480 (Cell Bank of Chinese Academy of Sciences). Cells were cultured at 37 °C in a humidified atmosphere of 5% CO<sub>2</sub> in RPMI-1640 medium (HyClone) supplemented with 10% (v/v) FBS (HyClone) and dispersed in identical 96-well plates. Compounds were dissolved in DMSO and serially diluted in saline to give final DMSO concentrations below 1%. Each tumor cell line was exposed to the test compounds at concentrations of 0.064, 0.32, 1.6, 8, and 40 μM for 48 h with cisplatin (DPP; Sigma, > 98%) as the positive control; cell viability was determined by MTT cytotoxicity assay by measuring the absorbance at 570 nm with a microplate reader (Bio-Rad 680) [23]. Three independent trials were conducted for each compound (n = 3). The IC<sub>50</sub> values and 95% confidence interval (CI) were estimated using GraphPad Prism 6.

## Supporting Information

The NMR, IR, UV, and HRTOF-ESIMS spectra of compounds 1–5, as well as the dose-response curves of cytotoxic activity, are available as Supporting Information.

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## Conflict of Interest

The authors declare no conflicts of interest.

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