

## 藏药船型乌头中的生物碱成分

黄圣卓<sup>1,3</sup>, 曹金鑫<sup>1,3</sup>, 蒋思萍<sup>2</sup>, 朱华结<sup>1\*</sup><sup>1</sup>中国科学院昆明植物研究所 植物化学与西部植物资源持续利用国家重点实验室, 昆明 650204;<sup>2</sup>西藏高原生物研究所 拉萨 850001; <sup>3</sup>中国科学院研究生院, 北京 100049

**摘要:** 为了研究藏药臭蚤草的活性成分, 从其总碱中分离并通过核磁共振和质谱等方法鉴定了 8 个生物碱, 包括 6 个二萜类生物碱 13-O-acetylhetisine (1)、2-acetyl-13-dehydro-11-epihetisine (2)、2-acetyl-13-dehydro-11-hetisine (3)、hetisinone (4)、尼奥林 (5) 和 foresticine (6), 以及异喹啉 (7) 和 dianthrime (8)。所有化合物均首次从该植物中分离得到。

**关键词:** 藏药; 船型乌头; 二萜生物碱

中图分类号: Q946.91; R284.2

文献标识码: A

Alkaloids of the Tibetan Medicinal Plant *Aconitum naviculare* StapfHUANG Sheng-zhuo<sup>1,3</sup>, CAO Jin-xin<sup>1,3</sup>, JIANG Si-ping<sup>2</sup>, ZHU Hua-jie<sup>1\*</sup><sup>1</sup>State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204, China; <sup>2</sup>Plateau Institute of Biology, Lhasa 850001, China;<sup>3</sup>Graduate School of the Chinese Academy of Sciences, Beijing 100049, China

**Abstract:** Six diterpenoid alkaloids 13-O-acetylhetisine (1), 2-acetyl-13-dehydro-11-epihetisine (2), 2-acetyl-13-dehydro-11-hetisine (3), hetisinone (4), neoline (5), and foresticine (6), together with isoquinoline (7) and dianthrime (8), were isolated from the Tibetan folk drug *Aconitum naviculare*. Their structures were elucidated based on <sup>1</sup>H and <sup>13</sup>C NMR and MS. This is the first report of the presence of compounds 1-8 in this plant.

**Key words:** Tibetan folk drug; *Aconitum naviculare*; diterpenoid alkaloids

## Introduction

The plant genus *Aconitum* (Ranunculaceae) has over 100 species distributed in the northern hemisphere. The whole plants of *Aconitum naviculare* Stapf are used in traditional Tibetan medicine system for the treatment of some inflammations like gastritis, hepatitis, nephritis<sup>[1]</sup>. Crude preparations from *Aconitum* species were popularly used in Asia, Alaska, and Europe<sup>[2]</sup> in folklore and traditional medicine for the treatment of traumatic injury<sup>[3]</sup>, as febrifuge and bitter tonic<sup>[4]</sup>, and as ingredients in intoxicating liquor<sup>[5]</sup>. However, little phytochemistry study has been reported on *A. naviculare*. In this paper, eight alkaloids were isolated from *A. naviculare*.

Received November 27, 2009; Accepted February 05, 2010

Foundation Item: This work was funded by the NSFC (30770235) 973 Program (2009CB522300) and Chinese Academy of Sciences (YZ-06-1).

\* Correspondence author. Tel: 86-871-5216179; E-mail: hjzhu@mail.kib.ac.cn

## Experimental Section

## General

Optical rotations determinations were carried on an OA AA-50 digital polarimeter. <sup>1</sup>H and <sup>13</sup>C NMR Spectra were recorded on Bruker AM-400 and DRX-500 spectrometers with CDCl<sub>3</sub> or CD<sub>3</sub>OD as solvent and TMS as standard. The MS data were recorded by a VG Auto-spec-3000 mass spectrometer; in *m/z* (rel. %). Column chromatography were carried out with SiO<sub>2</sub> (200-300 mesh, Qingdao Marine Chemistry Inc. CHCl<sub>3</sub>/MeOH) and Sephadex LH-20 (25-100 μm, Pharmacia Fine Chemical Co. Ltd. CHCl<sub>3</sub>/MeOH 1:1).

## Plant Material

Whole plants of *A. naviculare* were collected from Tibet (China) in June and August 2006. The specimen identified by professor Jiang Si-ping and stored in Plateau Institute of Biology Lhasa.

## Extraction and Isolation

The dried material (20 kg) was extracted with methanol a crude methanol extract (2.1 kg) was obtained. It's solution was acidified with 0.1 M H<sub>2</sub>SO<sub>4</sub>. The acidic aqueous extract was then basified (pH 10) with 20% aqueous Na<sub>2</sub>CO<sub>3</sub> solution and then extracted with CHCl<sub>3</sub>. The combined chloroform solution was washed with H<sub>2</sub>O and dried with Na<sub>2</sub>SO<sub>4</sub> and the solvent was evaporated to afford a crude alkaloid extract (40 g), which was subjected to repeated column chromatography (CC) (SiO<sub>2</sub>; CHCl<sub>3</sub>/MeOH, 100:0 to 0:100). The fractions eluted by CHCl<sub>3</sub>/MeOH 80:20 (3.6 g) was further purified by repeated CC (1. SiO<sub>2</sub>, CHCl<sub>3</sub>/MeOH, 100:1; 2. Sephadex LH-20, CHCl<sub>3</sub>/MeOH 1:1) to afford compounds **1** (1.5 mg), **2** (26.7 mg), **3** (1.6 mg), **4** (13.7 mg), **5** (40.5 mg), **6** (12.0 mg), **7** (2.1 mg) and **8** (13.0 mg).

## Result and Discussion

**13-O-Acetylhetisine (1)** C<sub>22</sub>H<sub>27</sub>NO<sub>4</sub>, white powder, [α]<sub>D</sub><sup>26</sup> +20.0 (c 0.0015, MeOH), EI-MS (70 eV) *m/z* 369 [M]<sup>+</sup> (45), 352 (12), 340 (30), 326 (38), 310 (100), 282 (48), 253 (22), 160 (21), 144 (13), <sup>1</sup>H NMR (CD<sub>3</sub>OD, 400 MHz) δ: 5.02 (2H, d, *J* = 2.2 Hz), 4.78 (1H, s), 4.29 (1H, d, *J* = 9.0 Hz), 3.62 (1H, d, *J* = 1.6 Hz), 3.58 (1H, d, *J* = 1.7 Hz), 3.43 (1H, s), 2.72 (2H, m), 2.57 (1H, d, *J* = 4.9 Hz), 2.53 (1H, s), 2.46 (1H, d, *J* = 2.6 Hz), 2.30 ~ 2.06 (9H, overlap), 1.84-1.76 (3H, overlap), 1.21 ~ 1.17 (4H, overlap); <sup>13</sup>C NMR (CD<sub>3</sub>OD, 100 MHz) δ: 215.4 (s), 172.4 (s), 146.7 (s), 109.3 (t), 74.9 (d), 74.8 (d), 71.6 (d), 66.7 (d), 63.7 (t), 61.3 (d), 55.6 (d), 50.82 (s), 50.80 (d), 48.6 (d), 45.6 (s), 40.5 (t), 36.7 (s), 36.2 (t), 34.0 (t), 33.7 (t), 29.8 (q), 21.3 (q). Comparing the data to the reported results in literature [6], the structure is defined as 13-O-acetylhetisine.

**2-Acetyl-13-dehydro-11-epihetisine (2)** C<sub>22</sub>H<sub>27</sub>NO<sub>4</sub>, white powder, [α]<sub>D</sub><sup>26</sup> + 48.7 (c 0.01335, MeOH), ESI-MS (pos.) *m/z* 370 [M + H]<sup>+</sup>, <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ: 5.03 (1H, d, *J* = 9.0 Hz), 4.83 (1H, s), 4.68 (1H, s), 4.05 (1H, d, *J* = 9.2 Hz), 3.77 (2H, s), 3.39 (1H, d, *J* = 13.9 Hz), 3.34 (1H, s), 2.99 (1H, d, *J* = 12.6 Hz), 2.50 ~ 2.03

(14H, overlap), 1.74 (1H, d, *J* = 12.6 Hz), 1.16 (3H, s), <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ: 210.1 (s), 171.1 (s), 142.9 (s), 109.6 (t), 75.1 (d), 69.6 (d), 69.1 (d), 65.6 (d), 61.4 (t), 57.4 (d), 54.9 (s), 52.1 (d), 49.8 (d), 49.1 (t), 47.2 (d), 44.2 (s), 42.7 (t), 41.3 (s), 34.0 (t), 32.5 (t), 28.0 (q), 21.1 (q). Comparing the data to the reported results in literature [7], the structure is defined as 2-acetyl-13-dehydro-11-epihetisine.

**2-Acetyl-13-dehydro-11-hetisine (3)** C<sub>22</sub>H<sub>27</sub>NO<sub>4</sub>, white powder, [α]<sub>D</sub><sup>26</sup> + 100.0 (c 0.0003, MeOH), ESI-MS (pos.) *m/z* 370 [M + H]<sup>+</sup> (60); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz) δ: 5.11 (1H, d, *J* = 9.5 Hz), 4.94 (1H, s), 4.77 (1H, s), 4.30 (1H, d, *J* = 9.0 Hz), 3.90 (2H, s), 3.48 (2H, d, *J* = 13.3 Hz), 3.10 (1H, *J* = 12.8 Hz), 2.65-2.14 (14H, overlap), 1.89 (1H, d, *J* = 12.4 Hz), 1.18 (3H, s); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125 MHz) δ: 211.6 (s), 170.0 (s), 143.4 (s), 109.7 (t), 74.1 (d), 73.1 (d), 70.3 (d), 65.3 (d), 63.5 (t), 59.7 (d), 55.4 (s), 54.2 (d), 49.8 (t), 48.9 (d), 47.8 (d), 44.9 (t), 44.4 (s), 42.3 (s), 35.1 (t), 33.1 (t), 28.5 (q), 21.0 (q). Comparing the data to the reported results in literature [7], the structure is defined as 2-acetyl-13-dehydro-11-hetisine.

**Hetisinone (4)** C<sub>20</sub>H<sub>25</sub>NO<sub>3</sub>, white powder. [α]<sub>D</sub><sup>26</sup> + 16.0 (c 0.00685, MeOH), ESI-MS (pos.) *m/z* 328 [M + 1]<sup>+</sup> (83), <sup>1</sup>H NMR (CD<sub>3</sub>OD, 400 MHz) δ: 4.71 (1H, s, br), 6.63 (1H, s, br), 4.28 (1H, m), 4.20 (s, br), 4.16 (1H, m), 3.69 (1H, m), 3.62 (1H, m), 3.20 (1H, m), 2.78 (1H, d, *J* = 6.7 Hz), 2.74 (1H, d, *J* = 4.0 Hz), 2.62 (2H, m), 2.30 ~ 2.39 (5H, overlap), 1.79 ~ 2.11 (4H, overlap), 1.32 (3H, s); <sup>13</sup>C NMR (CD<sub>3</sub>OD, 100 MHz) δ: 211.3 (s), 144.7 (s), 109.2 (t), 74.6 (d), 70.8 (d), 70.7 (d), 67.2 (d), 61.4 (t), 57.2 (d), 54.7 (d), 50.8 (d), 50.6 (d), 49.8 (d), 49.3 (t), 44.6 (t), 44.4 (s), 41.7 (s), 34.3 (t), 32.9 (t), 28.3 (q). Comparing the data to the reported results in literature [8-10], the structure is defined as hetisinone.

**Neoline (5)** C<sub>24</sub>H<sub>39</sub>NO<sub>6</sub>, yellow powder. [α]<sub>D</sub><sup>26</sup> + 5.68 (c 0.01935, MeOH), ESI-MS (pos.) *m/z* 438 [M + 1]<sup>+</sup> (65). <sup>1</sup>H NMR (CD<sub>3</sub>OD, 400 MHz) δ:

4.22 (1H, d,  $J = 6.5$  Hz), 4.21 (1H, m), 3.82 (s, br), 3.58 (1H, d,  $J = 7.9$  Hz), 3.39 (1H, m), 3.36 (3H, s), 3.33 (3H, s), 3.30 (3H, s), 3.29 (2H, s), 3.27 (1H, m), 2.97 (1H, d,  $J = 11.1$  Hz), 2.89 (1H, s), 2.85 (1H, m), 2.75 (1H, m), 2.57 (1H, d,  $J = 11.1$  Hz), 2.25 (1H, d,  $J = 6.5$  Hz), 2.20 (2H, d,  $J = 7.1$  Hz), 2.20 (1H, m), 2.08 (1H, s), 1.95 (1H, m), 1.65 (1H, m), 1.63 (1H, m), 1.59 (2H, m), 1.27 (1H, s), 1.14 (3H, d,  $J = 7.1$  Hz); <sup>13</sup>C NMR (CD<sub>3</sub>OD, 100 MHz)  $\delta$ : 84.3 (d), 80.8 (t), 79.5 (d), 76.3 (d), 75.2 (s), 73.2 (d), 64.2 (d), 59.4 (q), 58.6 (t), 58.3 (q), 56.5 (q), 54.4 (d), 50.8 (s), 49.8 (d), 48.5 (d), 45.3 (d), 45.2 (d), 42.5 (t), 41.8 (d), 39.2 (s), 31.4 (t), 30.0 (t), 29.8 (t), 12.6 (q). Comparing the data to the reported results in literature <sup>[11]</sup>, the structure is defined as neonile.

**Forensicine (6)** C<sub>24</sub>H<sub>39</sub>NO<sub>6</sub>, yellow powder.  $[\alpha]_D^{26} + 18.5$  (c 0.00755, MeOH), ESI-MS (pos.)  $m/z$  [M + 1]<sup>+</sup> (67), <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$ : 4.19 (1H, m), 4.15 (1H, d,  $J = 6.5$  Hz), 3.65 (1H, s), 3.62 (1H, d,  $J = 8.0$  Hz), 3.34 (1H, m), 3.32 (3H, s), 3.31 (3H, s), 3.30 (3H, s), 3.24 (1H, d,  $J = 8.0$  Hz), 2.67 (2H, m), 2.51 (1H, m), 2.45 (1H, m), 2.26 ~ 2.35 (3H, overlap), 2.16 (2H, m), 2.03 (3H, m), 1.83 ~ 1.87 (2H, overlap), 1.56 (1H, m), 1.43-1.55 (3H, m), 1.11 (3H, t,  $J = 7.0$  Hz). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125 MHz)  $\delta$ : 83.1 (d), 81.8 (d), 80.2 (t), 75.9 (d), 74.2 (s), 72.2 (d), 63.8 (d), 59.2 (q), 57.9 (q), 57.0 (t), 56.3 (q), 52.1 (d), 49.4 (d), 48.2 (d), 48.2 (t), 44.8 (d), 44.0 (d), 42.8 (t), 40.3 (d), 38.1 (s), 29.8 (t), 29.4 (t), 29.3 (t), 13.0 (q). Comparing the data to the reported results in literature <sup>[12]</sup>, the structure is defined as forensicine.

**Isoquinoline (7)** C<sub>9</sub>H<sub>7</sub>N, light yellow oil. ESI + MS  $m/z$  130 [M + 1]<sup>+</sup> (78). <sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz)  $\delta$ : 9.12 (1H, s), 8.41 (1H, d,  $J = 8.2$  Hz), 7.86 (1H, m), 7.71 (1H, m), 7.56 (1H, m), 7.50 (2H, m). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 125 MHz)  $\delta$ : 152.2 (d), 142.4 (d), 135.8 (d), 130.3 (d), 128.4 (d), 127.3 (d), 126.9 (d), 126.1 (d), 120.1 (d). Comparing the data to the reported results in literature

<sup>[13]</sup>, the structure is defined as isoquinoline.

**Dianthramide B (8)** *N*-salicyl-4-hydroxyanthranilic acid methyl ester, C<sub>15</sub>H<sub>13</sub>NO<sub>5</sub>, white powder. EI-MS (70 eV)  $m/z$  287 [M]<sup>+</sup> (16), 255 ((9), M-MeOH), 167 ((100), M-34ArCOO), 135 ((50), M-ArCOO-MeOH), 121 ((44), ArCOOH); FAB-MS: 286 [M - H]<sup>-</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$ : 12.3 (1H, s, -OH), 11.9 (1H, s, -OH), 8.68 (1H, d,  $J = 9.5$  Hz), 7.79 (1H, d,  $J = 8.5$  Hz), 7.56 (1H, d,  $J = 2.9$  Hz), 7.44 (1H, s), 7.12 (1H, dd,  $J = 9.0$ , 2.9 Hz), 6.99 (2H, m), 3.98 (3H, s); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$ : 170.9 (d), 171.2 (s), 166 (s), 163.4 (s), 144.8 (s), 136.5 (d), 135.2 (d), 129.2 (d), 121.3 (d), 120.2 (d), 118.2 (d), 113.1 (d), 109.4 (d), 109.3 (s), 53.6 (q). Comparing the data to the reported results in literature <sup>[14,15]</sup>, the structure is defined as dianthramide B (*N*-salicyl-4-hydroxyanthranilic acid methyl ester).

**Acknowledgements** This work was funded by NSFC (30770235 and 30873141) and Chinese Academy of Sciences (YZ-06-1), 973 Program (2009CB522304) and Key State Laboratory of Phytochemistry and Plant Resources in West China of Kunming Institute of Botany (P2008-ZZ17).

## Reference

- Editorial board of Flora of China. Flora of China. Beijing: Science Press, 1997, 27: 186-187.
- Wink M, Le Q, Philip W. In Alkaloids: Biochemistry Ecology and Medical Applications. New York: Plenum Press, 1998: 11-44.
- Yue J, Xu J, Zhao Q *et al.* Diterpenoid alkaloids from *Aconitum leucostomum*. *J Nat Prod*, 1996, 59: 277.
- Chopra RN, Chopra IC *et al.* Chopra's Indigenous Drugs of India. Calcutta: U. N. Dhur and Sons Pty Ltd, 1958: 54-57.
- Baillie LC, Batsanov A *et al.* Synthesis of the A/E/F sections of conaconitine, napelline and related diterpenoid alkaloids of the aconitine group. *J Chem Soc, Perkin Trans 1*, 1998, 20: 3471.
- Benn M, Richardson JF *et al.* Hetisine 13-O-acetate, a new diterpenoid alkaloid from *Delphinium nuttallianum* Pritz. *Heterocycles*, 1986, 24: 1605-1607.

(下转第 660 页)

## 参考文献

- 1 Lu D( 卢丹 ), Li YP( 李亚平 ). Research progress about chemical constituents and pharmacology effects of *Syringa*. *J Changchun Coll Tradit Chin Med* ( 长春中医学院学报 ), 2001, 17 ( 4 ): 58-60.
  - 2 Zhang SJ( 张树军 ), Zhang JF( 张军锋 ), Wang JL( 王金兰 ). Chemical constituents in stem bark of *Syringa Oblata*. *Chin Tradit Herb Drugs* ( 中草药 ) 2006, 37: 1624-1626.
  - 3 Zhang JF( 张军锋 ), Ji H( 焦华 ), Wang JL( 王金兰 ) *et al.* Chemical constituents in barks of *Syringa Oblata* Lindl. *Nat Prod Res Dev* ( 天然产物研究与开发 ) 2007, 8: 617-619.
  - 4 Masaco K, Yamauchi Y. Studies the constituents of *Syringa species*. *Yakugakuzasshi*, 1987, 107: 350-354.
  - 5 Asaka Y, Kamikawa T, Tokoroyama T. Structure and absolute configuration of *Syringa vulgaris*. *Tetrahedron*, 1970, 26: 2365-2367.
  - 6 Maillard M, Adewunmi CO, Hostettman K. A triterpene glycoside from the fruits of *Tetrapleura tetraptera*. *Phytochemistry*, 1992, 31: 1321-1323.
  - 7 Kriwacki RW, Pitner TP. Current aspects of practical two dimensional ( 2D ) nuclear magnetic resonance ( NMR ) spectroscopy: applications to structure elucidation. *Pharm Res*, 1989, 6: 531-554.
  - 8 Yu DQ( 于德权 ), Yang JS( 杨峻山 ), Xie JX( 谢晶曦 ). Handbook of Analytical Chemistry ( 分析化学手册 ). Beijing: Chemical Industry Press, 1989.
  - 9 Furukawa M, Takagi N, Ikeda T *et al.* Two novel long-chain alkanolic acid esters of lupeol from *Alecrim-propolis*. *Chem Pharm Bull* 2002, 50: 439.
  - 10 Li J( 李军 ), Zhang SX( 张淑霞 ), Guo HQ( 郭华强 ) *et al.* Extraction and derermination of hydroxy phenlethanol compound in *Syringa Oblata* lindl bark. *Chem Adhesion* ( 化学与黏合 ) 2008, 30: 37-39.
  - 11 Catola V, Bighelli A, Rezzi S *et al.* Composition and chemical variability of the triterpene fraction of dichloromethane extracts of cork ( *Quercus suber* L ). *Ind Crop Prod* 2002, 15: 15-22.
- 
- ( 上接第 657 页 )
- 7 Jiang Q, Glinski JA, *et al.* Heterocyclic derivatives, Part 3. Rearrangement of 11-acetyl-2, 13-didehydrohetisine and 13-dehydro-2, 11-diacetylhetisine. *Heterocycles*, 1988, 27: 925-932.
  - 8 Aplin RT, Benn MH, *et al.* Structure of hetisinone. *Can J Chem*, 1968, 46: 2635-2636.
  - 9 Jones PG. Crystal structure of hetisinone hydrate, C<sub>20</sub>H<sub>27</sub>NO<sub>4</sub>. *Z. Kristallogr*, 1993, 208: 344-346.
  - 10 Pelletier SW, Glinski JA, *et al.* The diterpenoid alkaloids of *Delphinium tatsienense* Franch. *Heterocycles*, 1983, 20: 1347-1354.
  - 11 Pelletier SW, Djarmati Z. Carbon-13 nuclear magnetic resonance: aconitine-type diterpenoid alkaloids from *Aconitum* and *Delphinium* species. *J Am Chem Soc*, 1976, 98: 2626-2636.
  - 12 Pelletier SW, Ying CS, *et al.* The structures of forestine and foresticine, two new C<sub>19</sub>-diterpenoid alkaloids from *Aconitum forrestii* Stapf. *J Nat Prod*, 1984, 47: 474-477.
  - 13 Johns SR, Willing RI. Carbon-13 N. M. R spectra of quinoline and methylquinolines. The magnitude of the vicinal ( peri ) 3J<sub>CCCH</sub> coupling constants. *Aust J Chem*, 1976, 29: 1617-1622.
  - 14 Niemann GJ, Liem J, *et al.* The amide-type phytoalexin activity of carnation extracts is partly due to an artifact. *Phytochemistry*, 1991, 30: 3923-3927.
  - 15 Ponchet M, Martin-Tanguy J, *et al.* Dianthramides A and B, two *N*-benzoylanthranilic acid derivatives from elicited tissues of *Dianthus caryophyllus*. *Phytochemistry*, 1984, 23: 1901-1903.