

## 岩芋中的一个新苯丙素苷<sup>\*</sup>

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**摘要:** 从岩芋 (*Remusatia vivipara*) 干燥的球茎中分离得到 10 个化合物, 其中一个为新的苯丙素苷, 经波谱学分析及酸水解的方法确定该新化合物的结构为 Caffeyl alcohol-3-*O*-*D*-glucopyranoside。已知化合物包括 3 个苯丙素类 (松柏苷, caffeyl alcohol 和松柏醇), 3 个新木脂素 [4, 7, 9, 9'-tetrahydroxy-3, 3'-dimethoxy-8-*O*-4'-neolignan-7-ene, (7*R*, 8*S*)-<sup>7</sup>-3, 3'-dimethoxy-4, 7, 9, 9'-tetrahydroxy-8-*O*-4'-neolignan-7-*O*-*D*-glucopyranoside, 以及 dehydrodiconiferyl alcohol-4'-*D*-glucoside], 1 个酰胺 [(2*E*, 4*E*)-*N*-isobutyl-2, 4-decadienamide], 1 个甾体皂苷 (methyl proto-taccaoside) 和 1 个三萜皂苷 (saxifragifolin B)。所有化合物均为首次从岩芋属植物中分离得到。

**关键词:** 岩芋; 天南星科; 苯丙素苷

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## A New Phenylpropanoid Glucoside from *Remusatia vivipara* (Araceae)

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**Abstract:** A new phenylpropanoid glucoside, caffeyl alcohol-3-*O*-*D*-glucopyranoside, together with nine known compounds, was isolated from the dry corms of *Remusatia vivipara* Schott. The structure of the new compound was determined by the spectroscopic method and acidic hydrolysis. The known compounds included three phenylpropanoids (coniferin, caffeyl alcohol and coniferyl alcohol), three neolignans [4, 7, 9, 9'-tetrahydroxy-3, 3'-dimethoxy-8-*O*-4'-neolignan-7-ene, (7*R*, 8*S*)-<sup>7</sup>-3, 3'-dimethoxy-4, 7, 9, 9'-tetrahydroxy-8-*O*-4'-neolignan-7-*O*-*D*-glucopyranoside, and dehydrodiconiferyl alcohol-4'-*D*-glucoside], an amide [(2*E*, 4*E*)-*N*-isobutyl-2, 4-decadienamide], a steroid saponin (methyl proto-taccaoside) and a triterpenoid saponin (saxifragifolin B). All compounds were isolated from the genus *Remusatia* for the first time.

**Key words:** *Remusatia vivipara*; Araceae; phenylpropanoid glucoside

*Remusatia vivipara* (Lodd.) Schott (Araceae) is a perennial herb mainly epiphytic on rocks and cliff-ledges in the subtropical forests of Asia such as SW China, Sri Lanka, Ne-

pal, India, Myanmar, Thailand, Vietnam and Indonesia, and Cameroon in West Africa (Li, 1979). Its corms are strongly poisonous but used externally to treat breast mastitis, absces-

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ses, acarasis and so on (Health Bureau of Yunnan Province, 1973). Up to now, the chemical constituents of the plant have not been reported yet. So a phytochemical investigation of the *R. vivipara* was carried out, which led to the isolation of a new phenylpropanoid glucoside (1) from the corms, together with nine known compounds: coniferin (2) (Sugiyama *et al.*, 1993), caffeyl alcohol (3) (Quideau and Ralph, 1992), coniferyl alcohol (4) (Quideau and Ralph, 1992), 4, 7, 9, 9-tetrahydroxy-3, 3-dimethoxy-8-*O*-neolignar-7-ene (5) (Lourith *et al.*, 2005), (7*R*, 8*S*)-7, 9, 9-tetrahydroxy-8-*O*-neolignar-7-*O*- $\beta$ -D-glucopyranoside (6) (Ma *et al.*, 2008), dehydroconiferyl alcohol-4- $\beta$ -D-glucoside (7) (Ye *et al.*, 2004), (2*E*, 4*E*)-*N*-isobutyl-2, 4-decadienamide (8) (Yasuda *et al.*, 1981), methyl proto-taccaoside (9) (Idaka *et al.*, 1991) and saxifragifolin B (10) (Waltho *et al.*, 1986). The structure elucidation of the new compound is reported.

## Results and Discussion

Compound 1 was obtained as white amorphous powder. Negative HRESIMS analysis of the compound exhibited a quasimolecular ion peak at  $m/z$  327. 1083 [ $M-H$ ]<sup>-</sup> (calcd, 327. 1079), corresponding to the molecular formula C<sub>15</sub> H<sub>20</sub> O<sub>8</sub> for 1. Its IR spectrum showed absorption bands for hydroxy groups (3441 cm<sup>-1</sup>) and a phenyl ring (1631 cm<sup>-1</sup>, 1611 cm<sup>-1</sup> and 1515 cm<sup>-1</sup>). In the <sup>13</sup>C NMR spectrum of 1 (Table 1), 15 carbon signals including two oxygenated methylenes [ $\delta_c$  63. 8 (C-9), 62. 4 (C-6)], ten methines [ $\delta_c$  131. 5 (C-7), 127. 5 (C-8), 123. 4 (C-6), 117. 0 (C-5), 116. 5 (C-2), 104. 4 (C-1), 78. 4 (C-5), 77. 6 (C-3), 74. 9 (C-2), 71. 4 (C-4)] and three quaternary carbons [ $\delta_c$  148. 1 (C-4), 146. 9 (C-3), 130. 8 (C-1)] were observed. The <sup>1</sup>H NMR spectrum of 1 (Table 1) displayed signals for an ABX benzene ring [ $\delta_H$  7. 31 (1H, d,  $J = 1. 6$  Hz, H-2), 6. 95 (1H,

dd,  $J = 8. 2, 1. 6$  Hz, H-6) and 6. 77 (1H, d,  $J = 8. 2$  Hz, H-5)], a *trans*-allylic moiety [ $\delta_H$  6. 48 (1H, d,  $J = 15. 9$  Hz, H-7), 6. 20 (1H, dt,  $J = 15. 9, 6. 0$  Hz, H-8) and 4. 18 (2H, dd,  $J = 6. 0, 1. 1$  Hz, H-9)]. In addition, its <sup>1</sup>H NMR spectrum showed an anomeric proton signal at 4. 75 (1H, d,  $J = 7. 1$  Hz, H-1), indicating the presence of a sugar moiety in 1. Acid hydrolysis of 1 yielded a D-glucose and determined by TLC and its optical rotation ( $[\alpha]_D^{26} = 50. 3$  (c 0. 055, H<sub>2</sub>O)). The coupling constant ( $J = 7. 1$  Hz) of H-1 and H-2 deduced the glucose to be a  $\beta$ -anomeric configuration.

Table 1 <sup>1</sup>H (500 MHz) and <sup>13</sup>C (100 MHz) NMR Data of 1 in CD<sub>3</sub>OD ( $\delta$  in ppm,  $J$  in Hz)

No.	c	H
1	130. 8 (s)	
2	116. 5 (d)	7. 31 (d, $J = 1. 6$ )
3	146. 9 (s)	
4	148. 1 (s)	
5	117. 0 (d)	6. 77 (d, $J = 8. 2$ )
6	123. 4 (d)	6. 95 (dd, $J = 8. 2, 1. 6$ )
7	131. 5 (d)	6. 48 (d, $J = 15. 9$ )
8	127. 5 (d)	6. 20 (dt, $J = 15. 9, 6. 0$ )
9	63. 8 (t)	4. 18 (dd, $J = 6. 0, 1. 1$ )
1	104. 4 (d)	4. 75 (d, $J = 7. 1$ )
2	74. 9 (d)	3. 49 (m)
3	77. 6 (d)	3. 47 (m)
4	71. 4 (d)	3. 39 (m)
5	78. 4 (d)	3. 43 (m)
6	62. 4 (t)	3. 91 (dd, $J = 12. 1, 2. 2$ ) 3. 71 (dd, $J = 12. 1, 5. 5$ )

In the HMBC spectrum (Fig 1), H-7 showed correlations to C-2 and C-6, and H-8 showed to C-1, meanwhile H-2 and H-6 also showed correlations to C-1. These correlations indicated that the propenol moiety was placed at C-1. The assignments of C-3 and C-4 were ascertained by the HMBC correlation from H-5 to C-1 and C-3, H-6 to C-2 and C-4, H-2 to C-4 and C-6. The linkage of the  $\beta$ -D-glucose substituent to C-3 was established by the correlation from H-1 to C-3. Therefore, 1 was elucidated as caffeyl alcohol-3-*O*- $\beta$ -D-glucopyranoside.

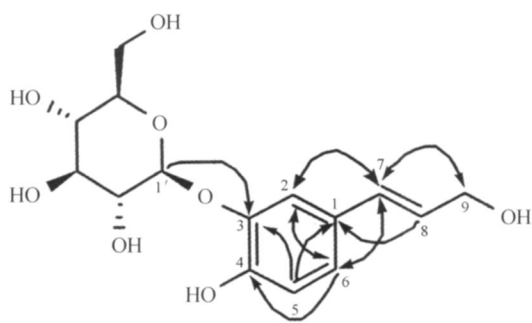


Fig. 1 Key HMBC correlations of 1

## Experimental

**General** MCI gel CHP 20P (75-150  $\mu\text{m}$ , Mitsubishi Chemical Corporation, Tokyo), silica gel G (300-400 mesh, Qingdao Makall Group Co., Ltd),  $\text{C}_{18}$  silica gel (40-75  $\mu\text{m}$ , Fuji Silysia Chemical Ltd.), silica gel H (10-40  $\mu\text{m}$ ), and Sephadex LH-20 (GE Healthcare Bio-Sciences AB) were used for column chromatography, and silica gel GF<sub>254</sub> (Qingdao), for preparative TLC as pre-coated plates. The TLC spots were visualized under UV light and by dipping into 5%  $\text{H}_2\text{SO}_4$  in alcohol, followed by heating. Semipreparative HPLC was carried out on an Agilent 1200 series pump equipped with a diode array detector and a Zorbax SB- $\text{C}_{18}$  column (5.0  $\mu\text{m}$ , 9.4  $\times$  250 mm). 1D and 2D NMR spectra were obtained on BRUKER AM-400 and DRX-500 spectrometers with TMS as internal standard. MS analyses were performed on a VG Auto Spec-3000 mass spectrometer. Optical rotations were measured with a Horiba SEPA-300 polarimeter. UV spectra were determined on a Shimadzu double-beam 210A spectrometer. IR spectra were measured on a Bio-Rad FTS-135 infrared spectrophotometer with KBr disks.

**Plant material** The corms of *R. vivipara* were collected in October 2008 from Luxi City, Yunnan Province, People's Republic of China and identified by Dr. Hu Guang-Wan (Kunming Institute of Botany, Chinese Academy of Sciences). A voucher specimen (No. LX005) was deposited at the Laboratory of Ethnobotany, Kunming Institute of Botany.

**Extraction and isolation** The air-dried, powdered corms of *R. vivipara* (1.0 kg) was exhaustively extracted with 95% EtOH (3 times) and then with MeOH (twice) under reflux. The crude extract (68 g) was suspended in  $\text{H}_2\text{O}$  and then partitioned successively with EtOAc and *n*-BuOH to obtain two fractions: EtOAc (A, 13 g), and *n*-BuOH (B, 6 g). Fr. B was subjected to column chroma-

tography over MCI gel CHP 20P with MeOH- $\text{H}_2\text{O}$  (a gradient elution of increasing concentration) and gained five fractions (B<sub>1</sub>-B<sub>5</sub>). Each fraction was further purified by repeated column chromatography ( $\text{C}_{18}$  silica gel, Sephadex LH-20, silica gel) and then semipreparative HPLC to obtain compounds. From Fr. B<sub>1</sub>, compounds 1 (4.2 mg) and 2 (6.1 mg) were obtained. Fr. B<sub>2</sub> gave compound 6 (4.3 mg). Fr. B<sub>4</sub> afforded compounds 9 (30.7 mg) and 10 (15.2 mg). Fr. A was subjected to reversed-phase column chromatography over  $\text{C}_{18}$  silica gel eluting with a gradient of increasing MeOH in  $\text{H}_2\text{O}$  (5%-95%) and obtained five fractions (A<sub>1</sub>-A<sub>5</sub>). Subsequently, Fr. A<sub>2</sub> and A<sub>4</sub> were chromatographed on Sephadex LH-20 column, repeated silica gel column and then followed by semipreparative HPLC, to give compounds 3 (7.5 mg), 4 (4.6 mg), 5 (5.6 mg), and 7 (4.0 mg) from Fr. A<sub>2</sub> and 8 (3.5 mg) from Fr. A<sub>4</sub>.

**Acid hydrolysis of 1**: A solution of 1 (4.0 mg) in 2 mol  $\text{L}^{-1}$  HCl (2 ml) was heated at 90  $^\circ\text{C}$  for 4 h. After evaporating the acidic solution, the reaction mixture was subjected to column chromatography over silica gel ( $\text{CHCl}_3$ -MeOH- $\text{H}_2\text{O}$ , 3 : 1 : 0.05) to yield a D-glucose (1.1 mg), which was detected by TLC comparing with the authentic sample and its optical rotation,  $[\alpha]_{\text{D}}^{26.3} 50.3$  (*c* 0.055,  $\text{H}_2\text{O}$ ).

**Caffeoyl alcohol-3-O- $\beta$ -D-glucopyranoside (1)**: White amorphous powder;  $[\alpha]_{\text{D}}^{25.5} 35.2$  (*c* 0.125, MeOH); UV (MeOH)  $\lambda_{\text{max}}^{\text{MeOH}}$  (log  $\epsilon$ ) 263 (3.54) nm; IR  $\lambda_{\text{max}}^{\text{KBr}}$   $\text{cm}^{-1}$ : 3441, 1631, 1610, 1515, 1283, 1074;  $^1\text{H}$  and  $^{13}\text{C}$  NMR (see Table 1); ESIMS  $m/z$  363 [ $\text{M} - \text{Cl}$ ] $^-$ , 327 [ $\text{M} - \text{H}$ ] $^-$ ; HRESIMS  $m/z$  327.1083 [ $\text{M} - \text{H}$ ] $^-$  (calcd for  $\text{C}_{15}\text{H}_{19}\text{O}_8$ , 327.1079).

**Caffeoyl alcohol (3)**: White amorphous powder (MeOH);  $^1\text{H}$  NMR ( $\text{CD}_3\text{OD}$ , 400 MHz)  $\delta$ : 6.86 (1H, *d*,  $J = 1.7$  Hz, H-2), 6.68 (1H, *d*,  $J = 8.1$  Hz, H-5), 6.71 (1H, *dd*,  $J = 8.1, 1.7$  Hz, H-6), 6.42 (1H, *d*,  $J = 15.8$  Hz, H-7), 6.10 (1H, *dt*,  $J = 15.8, 6.0$  Hz, H-8), 4.16 (2H, *dd*,  $J = 6.0, 1.3$  Hz, H-9); ESIMS  $m/z$  165 [ $\text{M} - \text{H}$ ] $^-$ . This compound was determined according to its MS spectrum and by comparing its  $^1\text{H}$  NMR data with compounds 1 and 4. Its  $^1\text{H}$  NMR data were reported for the first time.

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