



Two new diterpenoids and other constituents from *Isodon rubescens*

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ABSTRACT

Two new ent-kaurene diterpenoids, 15 α -acetoxyl-6,11 α -epoxy-6 α -hydroxy-20-oxo-6,7-seco-ent-kaur-16-en-1,7-olide (**1**), 15 α -hydroxy-20-oxo-6,7-seco-ent-kaur-16-en-1,7 α (6,11 α)-diolide (**2**), together with ten known compounds (**5–14**) were isolated from the leaves of *Isodon rubescens*. Their structures were elucidated mainly by various spectroscopic techniques and finally confirmed by single-crystal X-ray diffraction. Compounds **1**, **2**, **8** and **12** were evaluated for their cytotoxicities against EC-1, U87, A549, MCF-7 and Hela cell lines.

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1. Introduction

The genus *Isodon* (Labiateae) was widely distributed in China, the leaves of which have been shown to contain some ent-kaurene type diterpenoids with cytotoxic activities [1–3]. In a continuation of our research work on diterpenoids, we have recently investigated *Isodon rubescens* (Hemsl.) Hara collected from the northern Henan Province of China. From the AcOEt extract of the leaves, two new ent-kaurene diterpenoids (**1** and **2**) and ten known diterpenoid compounds (**5–14**) were isolated (Fig. 1). In addition, four of the compounds (**1**, **2**, **8** and **12**) were tested for their cytotoxicity towards cancer cells EC-1, U87, A549, MCF-7 and Hela. This report describes the structure determination of compounds **1** and **2**.

2. Experimental

2.1. General

Melting points were measured using an uncorrected X-4 Digital Display micromelting point apparatus. Optical rota-

tions were taken on a Perkin-Elmer 341 polarimeter. IR spectra were obtained on a Nicolet 170 SX FT-IR spectrometer. ¹H, ¹³C and 2D NMR spectra were measured on a Bruker AM-400 NMR spectrometer using TMS as the internal standard. HR-ESI-MS were recorded on a Waters HPLCQ-ToF HR-MS spectrometer. Column chromatography were carried out on silica gel (200–300 mesh) and TLC on silica gel (GF₂₅₄, 10–40 μ m), and both materials were supplied by Qingdao Marine Chemical Co. 5-Fluorouracil used as a positive control was supplied by Shanghai Xudong pharmaceutical Co. Ltd, (Shanghai, China). Spots were detected on TLC under UV or by heating after spraying with 5% H₂SO₄ in C₂H₅OH.

2.2. Plant material

The plant material, *I. rubescens* (Hemsl.) Hara was collected in Huixian of Henan province, P. R. China, in Aug. 2008, and identified by Professor Changshan Zhu, Henan Agriculture University. A voucher specimen (No. 20080706) is deposited in Pharmacy College, Xinxiang Medical University.

2.3. Extraction and isolation

The air-dried leaves of *I. rubescens* (15 kg) were pulverized and extracted four times with Me₂CO/H₂O (7:3 v/v) at room

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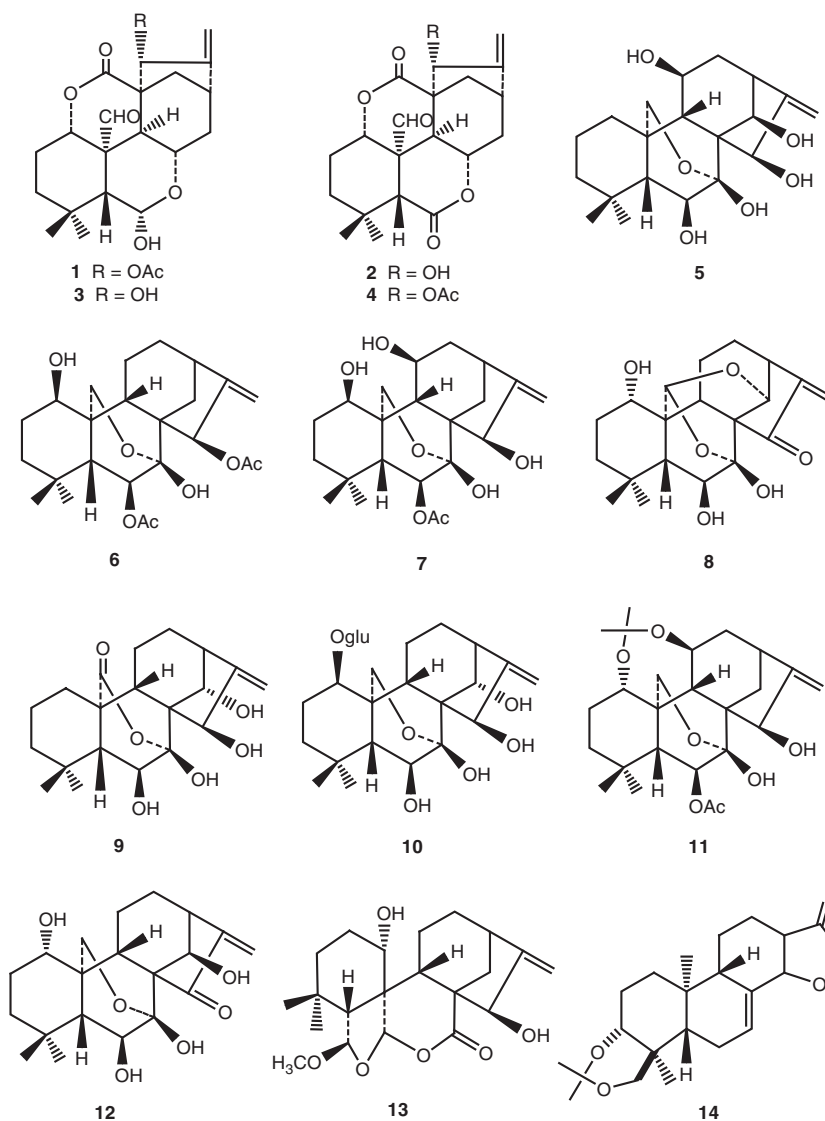


Fig. 1. The molecular structures of compounds 1–14.

temperature for 6 days and filtered. The filtrate was concentrated and partitioned successively between AcOEt and H₂O. The AcOEt extract (310 g) subjected to silica gel column (12×150 cm, 3000 g, 200–300 mesh) was gradually eluted with a system of CHCl₃—CH₃OH (1:0, 30:1, 20:1, 10:1, 5:1, 3:1) to give six factions according to their TLC analysis. From the faction 2 (CHCl₃—CH₃OH 30:1) compounds **1** (96 mg), **2** (29 mg) and **6** (56 mg) were obtained by repeated column chromatography over silica gel developing with CHCl₃—Me₂CO (20:1, 30:1) and followed by recrystallization. From the faction 3 (CHCl₃—CH₃OH 20:1) compounds **5** (14 mg), **7** (45 mg), **8** (60 mg), **9** (67 mg), **11** (14 mg), **12** (230 mg), **13** (110 mg) and **14** (10 mg) were isolated by further purified on silica gel column eluting with CHCl₃—MeOH (30:1) and CHCl₃—Me₂CO (15:1, 20:1). From the faction 5 (CHCl₃—CH₃OH 5:1) compound **10** (110 mg) was separated by repeated column

chromatography over silica gel eluted with CHCl₃—MeOH (8:1, 10:1) and CHCl₃—(Me)₂CHOH (10:1).

2.4. Spectroscopic data

15 α -acetoxy-6,11 α -epoxy-6 α -hydroxy-20-oxo-6,7-seco-ent-kaur-16-en-1,7-olide (**1**). Colorless crystals from Me₂CO; m.p. 281–283 °C, $[\alpha]_D^{20}$ –173.1 (c 0.54, Me₂CO); IR (KBr): ν^{\max} = 3516, 2987, 2967, 2940, 2874, 1743, 1715, 1700, 1660, 1373, 1181, and 919 cm⁻¹; HR-ESI-MS: m/z 427.1735 [M + Na]⁺, requires m/z 427.1733 for C₂₂H₂₈O₇Na. ¹H- and ¹³C-NMR data: see Table 1.

15 α -hydroxy-20-oxo-6,7-seco-ent-kaur-16-en-1,7 α (6,11 α)-diolide (**2**). Colorless needles from Me₂CO; m.p. 260–262 °C, $[\alpha]_D^{23}$ –135.7 (c 0.28, Me₂CO); IR (KBr): ν^{\max} = 3375, 1754, 1720, 1700, 1365, 1320, 1077, and

Table 1¹H (400 MHz) and ¹³C (100 MHz) NMR data of compounds **1** and **2** in pyridine-d₅ (δ in ppm).

Position	1 δ _H	1 δ _C	2 δ _H	2 δ _C
1	4.55 (dd, J = 4.0, 6.8 Hz)	77.4 d	4.70 (dd, J = 3.2, 11.2 Hz)	76.8 d
2	1.71, 1.99 m	25.8 t	2.02, 2.36 m	25.1 t
3	1.42, 2.77 m	40.7 t	1.43 m	39.6 t
4		33.5 s		33.0 s
5	1.91 s	50.1 d	2.84 s	50.8 d
6	5.85 s	92.3 d		169.7 s
7		172.1 s		173.7 s
8		48.7 s		50.8 s
9	3.27 (d, J = 11.6 Hz)	35.0 d	3.14 (d, J = 11.6 Hz)	38.9 d
10		50.6 s		50.4 s
11	4.29 (dt, J = 8.8, 11.2 Hz)	62.6 d	5.02 (dt, J = 8.8, 11.2 Hz)	70.2 d
12	1.67, 2.71 m	39.5 t	1.71, 2.65 m	37.7 t
13	2.78 m	37.7 d	2.81 m	37.2 d
14	1.14, 2.21 (d, J = 12.0 Hz)	32.3 t	1.82, 2.10 m	32.0 t
15	6.66 s	80.8 d	5.47 s	81.6 d
16		153.7 s		157.3 s
17	5.03, 5.11 brs	111.0 t	5.20, 5.44 brs	110.6 t
18	0.95 s	30.3 q	1.07 s	22.2 q
19	1.15 s	22.1 q	1.14 s	30.2 q
20	10.68 s	202.8 d	10.07 s	199.5 d
21		169.9 s		
22	1.96 s	20.5 q		

898 cm⁻¹; HR-ESI-MS: *m/z* 383.1473 [M + Na]⁺, requires *m/z* 383.1471 for C₂₀H₂₄O₆Na. ¹H- and ¹³C-NMR data: see Table 1.

2.5. Cytotoxicity assay

Cytotoxic activities of the isolated compounds **1**, **2**, **8** and **12** were evaluated using the standard MTT colorimetric method [4]. The cell lines used in this experiment were EC-1, U87, A549, MCF-7 and HeLa, and 5-fluorouracil was used as the reference compound.

3. Results and discussion

Compound **1** was obtained as colorless crystals, showed the quasi-molecular ion peak at *m/z* 427.1735 [M + Na]⁺ (calcd. 427.1733) in its HR-ESI-MS, corresponding to the molecular formula C₂₂H₂₈O₇. The IR (KBr) spectrum exhibited the presence of carbonyl groups (1700, 1715 and 1743 cm⁻¹) and a hydroxyl group (3516 cm⁻¹). The fact that no absorption was recorded in the UV spectrum indicated the absence of any conjugated systems in the molecule. A singlet at δ 1.96 (3H, s) in the ¹H NMR spectrum and two signals at δ 169.9 (s) and 20.5 (q) in the ¹³C NMR spectrum exhibited the presence of an acetoxy group. Comparison of other NMR data with those of known compound 6α,15α-dihydroxy-20-oxo-6,7-seco-6,11α-epoxy-1,7-olide-*ent*-kaur-16-ene (**3**) [5], indicated that **1** had the similar skeleton of 6,11α-epoxy-20-oxo-6,7-seco-*ent*-kaur-16-en-1,7-olide as **3**. The only difference between **1** and **3** was that **1** had one more acetoxy group. However, some differences were also observed. The signals at δ 5.85 (1H, s, H-15β) and δ 80.2 (d, C-15) in **3** were shifted downfield to δ 6.66 (1H, s, H-15β) and δ 80.8 (d, C-15) in **1**, respectively. Moreover, the signals at δ 49.8 (s, C-8) and δ 158.8 (s, C-16) in **3** were shifted upfield to δ 48.7 (s, C-8) and δ 153.7 (s, C-16) in **1**. These changes of chemical shift value suggested that the position of acetoxy group should be at C-15. The HMBC correlations of H-22 (δ 1.96, s) and H-15 (δ 6.66, s) with C-21 (δ

169.9 s) further confirmed the above assignment. The relative configuration of **1** was determined by the analysis of a NOESY experiment. In the NOESY spectrum, there were correlations of H-11 (δ 4.29 dt) with H-5 (δ 1.91 d) and H-6 (δ 5.85 s); H-15 (δ 6.66 s) with H-14 (δ 2.21 m) but no correlation with H-9 (δ 3.27 d). Thus, OH-6 and AcO-15 have the α orientation, respectively. On the basis of the above evidence, the structure of compound **1** was determined to be 15α-acetoxy-6,11α-epoxy-6α-hydroxy-20-oxo-6,7-seco-*ent*-kaur-16-en-1,7-olide, and named rube-crystal A (Fig. 1).

The single crystal X-ray diffraction analysis confirms the molecular structure of **1** proposed by spectroscopic methods. The crystal data were as follows: Colorless block crystal from Me₂CO, crystal size: 0.50 × 0.43 × 0.40 mm; crystal data: a = 10.669(3), b = 12.546(3), c = 14.710(3) Å, V = 1969.1(8) Å³, space group P2₁2₁2₁, Z = 4, D_{calc} = 1.364 g/cm³, λ = 0.71073 Å, μ(MoKα) = 0.10 mm⁻¹, F(000) = 864, and T = 103(2) K; a total of 15561 reflections were collected on a AFC10/Saturn724+ with a rotating anode tube. Refinement with 2552 reflections (2399 with I > 2σ) led to final R, R(all), GOF values of 0.0354, 0.0390, and 0.998. (Fig. 2). The crystal structure has been deposited at the Cambridge Crystallographic Data Centre and allocated the deposition number CCDC 814320.

Compound **2** was obtained as colorless crystal. The molecular formula of C₂₀H₂₄O₆ was deduced from the HR-ESI-MS (*m/z* 383.1473 [M + Na]⁺). Its IR spectrum showed the absorption bands for carbonyl groups (1754, 1720 and 1700 cm⁻¹) and a hydroxyl group (3375 cm⁻¹). The ¹³C NMR and DEPT spectra of **2** showed 20 carbon signals which were indicated to be composed of two lactonic carbonyl carbons, one aldehyde group, four quaternary carbon (including an olefinic one), six methines including three oxygenated ones, five methylenes including one olefinic one, and two methyls, which suggested **2** as an *ent*-kauranoid, combined with the consideration of the similar structure of Glaucoalactone (**4**) [6] previously isolated from this genus. The difference between these two compounds was

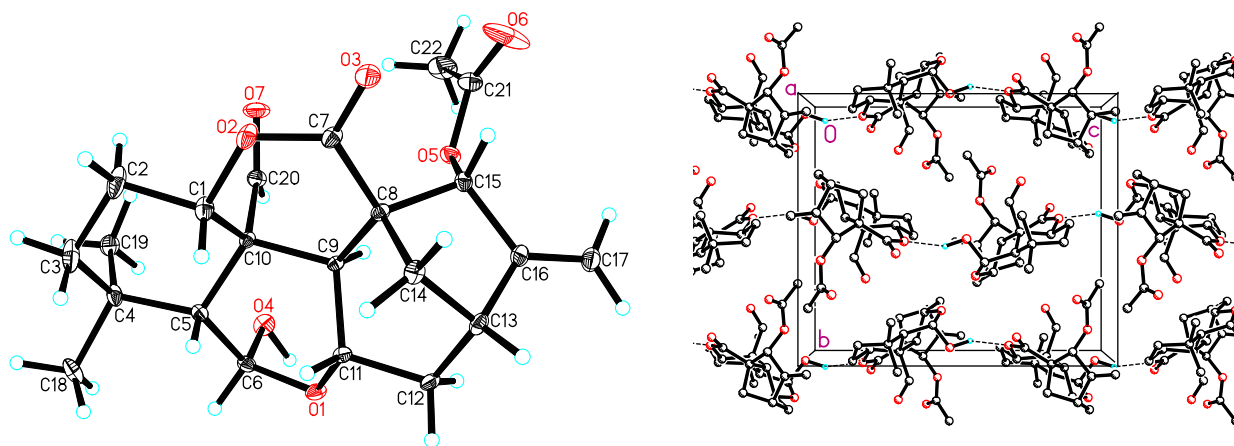


Fig. 2. The crystal structure of compound 1.

the group at C-15 position. There was a hydroxy group at C-15 in **2** instead of an acetoxy group in **4**. In the NOESY spectrum, H-15 (δ 5.47 s) showed correlation with H-14 (δ 1.82 m) but no correlation with H-9 (δ 3.14 d), it showed that the OH-15 was in α -orientation. Therefore, the structure of **2** was determined to be 15 α -hydroxy-20-oxo-6,7-seco-*ent*-kaur-16-en-1,7 α (6,11 α)-diolide, and named rubescrystal B (Fig. 1).

Its structural assignment including its relative stereochemistry was finally confirmed by X-ray analysis. The crystal data were as follows: Colorless crystal from Me₂CO, crystal size: 0.55 \times 0.21 \times 0.09 mm; crystal data: $a = 9.453(7)$, $b = 6.693(5)$, $c = 16.805(13)$ Å, $V = 1049.5(14)$ Å³, space group P2₁, $Z = 2$, $D_{\text{calc}} = 1.324$ g/cm³, $\lambda = 0.71073$ Å, $\mu(\text{MoK}\alpha) = 0.097$ mm⁻¹, $F(000) = 448$, and $T = 163(2)$ K; a total of 8093 reflections were collected on a AFC10/Saturn724+ with a rotating anode

tube. Refinement with 2552 reflections (1975 with $I > 2\sigma(I)$) led to final R , $R(\text{all})$, GOF values of 0.0527, 0.0650, and 1.001. (Fig. 3). The crystal structure has been deposited at the Cambridge Crystallographic Data Centre and allocated the deposition number CCDC 814321.

The known compounds **5–14** were identified by comparison of their NMR data with literature values. They were identified as Rubescensin C (**5**) [7], Trichokaurin (**6**) [8], Maoyecrystal F (**7**) [9], Ponicidin (**8**) [10], Rabdoternin A (**9**) [11], 1 α -O- β -D-glucopyranosyl enmenol (**10**) [12], Acetonide (**11**) [13], Oridonin (**12**) [14], Rabdonervosin B (**13**) [15], Acetonide of rubescensin J (**14**) [16].

From the anti-tumor activity data, it was found that compounds **8** and **12** have a significant inhibition on cells EC-1, U87, A549, MCF-7 and Hela, compound **2** has inhibition on cells

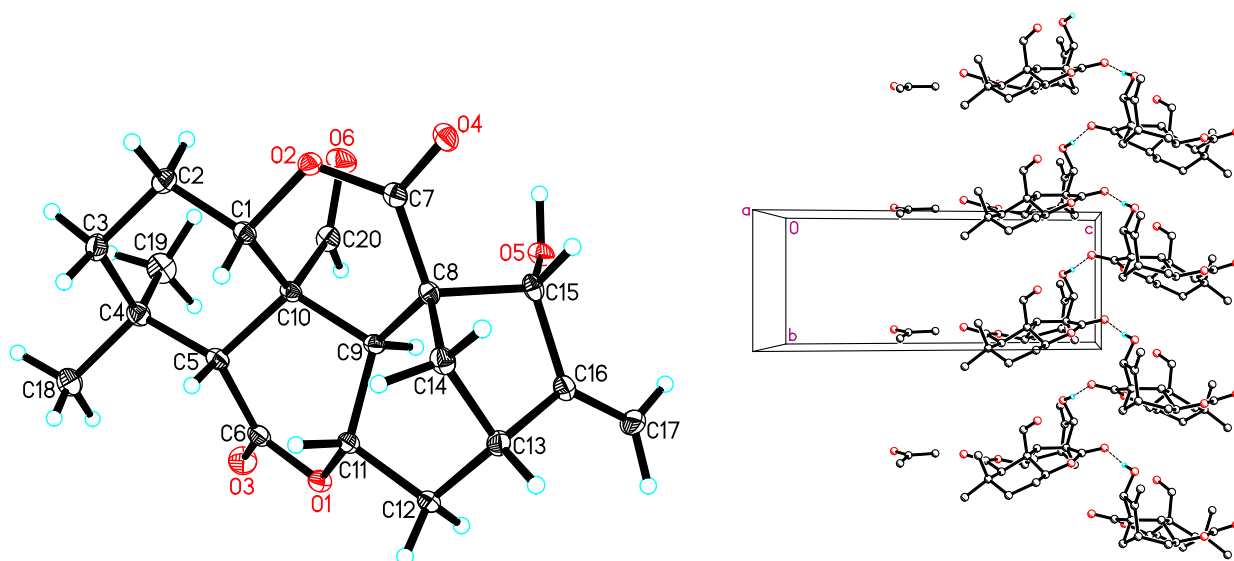


Fig. 3. The crystal structure of compound 2.

Table 2In vitro cytotoxicity data of compounds **1**, **2**, **8** and **12** by the MTT assay.

Compound	Cell lines/IC ₅₀ (μM)					
	EC-1	MCF-7	Hela	K562	A549	U87
1	678.19 ± 39.25	563.90 ± 47.38	365.17 ± 13.48	238.16 ± 15.07	784.39 ± 25.97	487.25 ± 9.01
2	37.69 ± 3.62	79.62 ± 6.33	80.07 ± 6.32	197.35 ± 2.10	462.13 ± 39.06	180.09 ± 11.44
8	43.87 ± 3.05	16.85 ± 2.63	28.14 ± 1.05	5.12 ± 0.46	73.33 ± 5.84	32.19 ± 4.07
12	66.25 ± 3.17	40.78 ± 3.26	37.59 ± 2.00	9.78 ± 1.78	188.75 ± 6.52	80.79 ± 3.40
5-fluorouracil	5.90 ± 0.49	21.08 ± 1.07	300.7 ± 11.28	108.66 ± 5.76	120.36 ± 3.79	28.41 ± 4.55

EC-1, MCF-7 and Hela, respectively; and compound **1** exhibited no significant inhibitory effects against selected cancer cells with IC₅₀ values over 100 μM. Cytotoxicity data: see Table 2.

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