This article was downloaded by: [Kunming Institute of Botany] On: 21 March 2012, At: 19:03 Publisher: Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



# Journal of Asian Natural Products Research

Publication details, including instructions for authors and subscription information: <u>http://www.tandfonline.com/loi/ganp20</u>

# Pregnane glycosides from Dracaena cochinchinensis

Qing-An Zheng & Chong-Ren Yang

<sup>a</sup> State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Yunnan, 650204, Kunming, China

Available online: 17 Oct 2011

To cite this article: Qing-An Zheng & Chong-Ren Yang (2003): Pregnane glycosides from Dracaena cochinchinensis, Journal of Asian Natural Products Research, 5:4, 291-296

To link to this article: <u>http://dx.doi.org/10.1080/1028602031000112003</u>

# PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <u>http://www.tandfonline.com/page/terms-and-conditions</u>

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae, and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand, or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.



# PREGNANE GLYCOSIDES FROM DRACAENA COCHINCHINENSIS

#### QING-AN ZHENG and CHONG-REN YANG\*

State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204, Yunnan, China

(Received 16 December 2002; Revised 6 January 2003; In final form 19 February 2003)

Two new pregnane glycosides, named dracaenoside C and D, were isolated from the fresh stems of *Dracaena cochinchinensis*. Their structures were established as  $3\beta$ , $14\alpha$ -dihydroxypregna-5,16(17)-diene-20-one 3-O- $\alpha$ -L-rhamnopyranosyl(1 $\rightarrow$ 4)]- $\beta$ -D-glucopyranoside (1) and  $3\beta$ , $14\alpha$ -dihydroxypregna-5,16(17)-diene-20-one 3-O- $\alpha$ -L-rhamnopyranosyl(1 $\rightarrow$ 2)[ $\beta$ -D-glucopyranosyl(1 $\rightarrow$ 3)]- $\beta$ -D-glucopyranoside (2) by means of 2D NMR spectral and chemical methods.

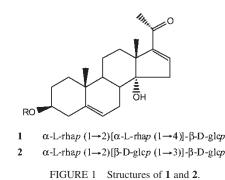
Keywords: Pregnane glycosides; Dracaenoside C; Dracaenoside D; Dracaena cochinchinensis; Agavaceae

# INTRODUCTION

Dragon's blood was used as a traditional Chinese medicine for more than 1500 years [1]. In China, the resin of *Dracaena cochinchinensis* S. C. Chen, named "longxuejie" was used as dragon's blood by local people for the treatment of diarrhea, diabetes, inflammation, bleeding, and as tonics. Chemical studies revealed that the resin contained a large amount of phenolic compounds, and some types of steroids, and aliphatic acids [2–6]. Steroidal saponins were also isolated from the fruits of *D. cochinchinensis* [7]. In a systematic chemical investigation on the resin and the original plants of dragon's blood we have reported two new C-22 steroidal lactone glycosides (dracaenoside A and B) from the fresh stems of this plant [8]. Further studies on the same plant led to the isolation of two new pregnane glycosides, named as dracaenoside C and D. Their structures were established as  $3\beta,14\alpha$ -dihydroxypregna-5,16(17)-diene-20-one  $3-O-\alpha$ -L-rhamnopyranosyl( $1\rightarrow 2$ )[ $\alpha$ -L-rhamnopyranosyl( $1\rightarrow 4$ ]- $\beta$ -D-glucopyranoside (dracaenoside C, 1) and  $3\beta,14\alpha$ -dihydroxypregna-5,16(17)-diene-20-one  $3-O-\alpha$ -L-rhamnopyranosyl( $1\rightarrow 3$ ]- $\beta$ -D-glucopyranoside (dracaenoside C, 1) we have spectral and chemical methods (Fig. 1).

<sup>\*</sup>Corresponding author. Tel.: +86-871-5223424. Fax: +86-871-5150124. E-mail: cryang@public.km.yn.cn

ISSN 1028-6020 print/ISSN 1477-2213 online © 2003 Taylor & Francis Ltd DOI: 10.1080/1028602031000112003



### **RESULTS AND DISCUSSION**

Dracaenoside C was obtained as a white amorphous powder,  $[\alpha]_{p}^{21}$  -41.90 in MeOH. Negative ion FABMS (a quasi-molecular peak at m/z 783  $[M-H]^+$ ) combined with <sup>13</sup>C NMR and DEPT indicated the molecular formula  $C_{39}H_{60}O_{16}$ . Acid hydrolysis with 1 M HCl in dioxane (v/v, 1:1) gave rhamnose and glucose in a ratio of about 2:1, which were further determined to be L-rhamnose and D-glucose by GC-MS analysis of a trimethylsilylated derivative of the acid hydrolysate (retention time for L-rhamnose of 7.17 min, D-glucose of 11.43 min), when compared with standard sugars. The <sup>13</sup>C and <sup>1</sup>H NMR spectra also exhibited the presence of two  $\alpha$ -L-rhamnopyranosyl groups [anomeric protons at  $\delta$  5.84 (1H, brs) and  $\delta$  6.38 (1H, brs); secondary methyl groups at  $\delta$  1.61 (3H, d, J = 6.04 Hz) and 1.76 (3H, d, J = 6.32 Hz); anomeric carbons at  $\delta$  102.95 and 102.11] and a  $\beta$ -D-glucopyranosyl group [anomeric proton at  $\delta$  4.91 (1H, d, J = 6.0 Hz); anomeric carbon at  $\delta$  100.25]. Besides the sugar signals, 21 carbon peaks were observed in the <sup>13</sup>C NMR spectrum, which implied a  $C_{21}H_{30}O_3$  composition for the aglycone. Preliminary analysis of the <sup>1</sup>H and <sup>13</sup>C NMR of the aglycone part led to the assignment of three tertiary methyl groups at  $\delta$  1.12, 2.15, 1.14 (each 3H, s), two olefinic protons at  $\delta$  5.27 (1H, brd) and  $\delta$  6.72 (1H, brs), indicating dracaenoside C to be a pregnane type glycoside. In the <sup>13</sup>C NMR spectrum, a ketone signal at  $\delta$  196.79 ascribed to C-20 was deduced by the downfield shift of a tertiary methyl group ( $\delta 2.15$ ), and its upfield shift revealed an olefinic group at C-16 (17). Another olefinic group was ascribed to C-5 by the downfield shift of the tertiary methyl groups (C-19) at  $\delta$  19.14. The assignment of the carbinol groups at C-3, C-14 was suggested by the resonance at  $\delta$  77.83, 82.78, by comparison with literature data [8]. Analysis of the HMBC spectrum with <sup>1</sup>H-<sup>1</sup>H COSY and HMQC spectra led to the assignment of the aglycone carbons. The conformations of the ring junction, B/C trans, C/D trans, were ascertained by the ROESY correlations observed between H-8 and H-19, H-8 and H-11B, H-8 and H-18. The 3B orientation of the oxygen atom was confirmed by the ROESY observed between H-3 and H-1ax, H-3 and H-4eq (Fig. 2).

Assignment of the sugar moieties was performed by the <sup>1</sup>H-<sup>1</sup>H COSY and HMQC-TOCSY spectra combined with the HMBC spectrum. The respective linkage positions of the inner glucopyranosyl and the rhamnopyranosyl were at C-3 of the aglycone and C-2' and C-4' of the glucose moiety by observing the <sup>3</sup>J correlations of each anomeric proton signal with the corresponding carbon signal [ $\delta_{\rm H}$  4.91 to  $\delta_{\rm C}$  77.935 (C-3);  $\delta_{\rm H}$  5.84 to  $\delta_{\rm C}$  78.136 (C-2'), and  $\delta_{\rm H}$  6.38 to  $\delta_{\rm C}$  77.937 (C-4'), respectively].

All of the above results led to the full assignments of <sup>13</sup>C NMR data for dracaenoside C. The structure of this new glycoside was therefore elucidated as

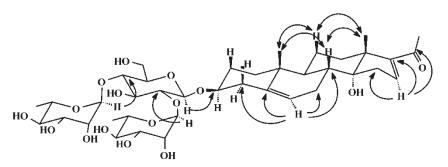


FIGURE 2 Selected HMBC and ROESY correlations of dracaenoside C ( $\rightarrow$  signal for HMBC;  $\leftarrow \rightarrow$  signal for ROESY).

 $3\beta$ ,  $14\alpha$ -dihydroxypregna-5, 16(17)-diene-20-one 3-O- $\alpha$ -L-rhamnopyranosyl $(1 \rightarrow 2)[\alpha$ -L-rhamnopyranosyl $(1 \rightarrow 4)]$ - $\beta$ -D-glucopyranoside (1).

Dracaenoside D was obtained as a white amorphous powder,  $[\alpha]_{p}^{21} - 25.17$  in MeOH. Negative ion HR-FABMS gave a quasi-molecular peak at m/z 799.3772 corresponding to a molecular formula of C<sub>39</sub>H<sub>60</sub>O<sub>17</sub>. The <sup>13</sup>C and <sup>1</sup>H NMR spectra of dracaenoside D were closely related to those of dracaenoside C except for the sugar moieties, indicating that dracaenoside D is also a pregnane-type glycoside with an identical aglycone to that of dracaenoside C. The <sup>13</sup>C and <sup>1</sup>H NMR spectra showed the presence of a rhamnose and two glucose moieties in dracaenoside D, instead of the two rhamnose and a glucose in dracaenoside C. This was confirmed by acid hydrolysis of dracaenoside D which gave rhamnose and glucose in a ratio of 1:2. D-Glucose and L-rhamnose moieties were determined by GC-MS analysis, as for dracaenoside C. By comparison of the NMR data with those of dracaenoside C, the rhamnose moiety was deduced to be linked at the C-2' position of the inner glucose and the terminal glucose moiety was deduced to be linked at C-3' position of the inner glucose moiety by the significant downfield shift of C-3' to 89.536. These assignments of the sugar moieties were consistent with the reference data [9]. Consequently, dracaenoside D was determined as  $3\beta$ ,  $14\alpha$ -dihydroxypregna-5, 16(17)diene-20-one  $3-O-\alpha-L$ -rhamnopyranosyl $(1\rightarrow 2)$ [ $\beta$ -D-glucopyranosyl $(1\rightarrow 3)$ ]- $\beta$ -D-glucopyranoside (2).

Dracaenoside C and D are new naturally occurring C-21 pregnane glycosides.

# **EXPERIMENTAL**

#### **General Experimental Procedures**

NMR spectra were run on Bruker AM-400 (400 MHz for <sup>1</sup>H NMR and 100 MHz for <sup>13</sup>C NMR) and DRX-500 (500 MHz for 2D NMR) instruments with TMS as internal standard; IR spectra were measured on a Bio-Rad FTS-135 spectrometer with KBr pellets. FAB-MS spectra were recorded on a VG Auto Spec-300 spectrometer. UV spectra were obtained on a Shimadzu double-beam 210A spectrophotometer. GC-MS was run on Fisons MD-800 GC/MS instruments using a 30QC2/AC-5 fused silica capillary column (30 m × 0.25 mm) under the following condition: filament current: 4.2 A; column temperature:  $180^{\circ}C/260^{\circ}C$ , programmed increase of  $5^{\circ}C \text{ min}^{-1}$ ; carrier gas: He; head pressure: 12 psi; EI-MS: 70 eV; ion source temperature:  $250^{\circ}C$ . Silica gel (200–300 mesh and 10–40 µm), RP-18 (40–63 µm) and Sephadex LH-20 were used for column chromatography.

### **Plant Material**

The fresh stems of *D. cochinchinensis* were collected at Xishuangbanna, Yunnan, China. A voucher specimen is deposited at the State Key Laboratory of Phytochemistry and Plant Resources in the west China, Kunming Institute of Botany, Chinese Academy of Sciences.

### **Extraction and Isolation**

The fresh stems (25.0 kg) were chipped and extracted with MeOH at 60°C. After removing of the MeOH under reduced pressure, the viscous concentrate was partitioned between H<sub>2</sub>O and n-BuOH. Column chromatography of the n-BuOH layer on silica gel, eluting with a gradient mixture of CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O, and finally with MeOH, gave five fractions (I–V). Fraction III was chromatographed over ODS silica gel, eluting with MeOH–H<sub>2</sub>O, and silica gel with CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O to give dracaenoside C (30 mg) and dracaenoside D (16 mg).

*Dracaenoside C.* (1) C<sub>39</sub>H<sub>60</sub>O<sub>16</sub>, is an amorphous white powder,  $[\alpha]_{\rm p}^{21}$  – 41.90 (MeOH, *c* 0.2); Negative ion FAB-MS *m/z* 783[M–H]<sup>+</sup>; UV (MeOH) λ<sub>max</sub> (nm): (log ε): 237; IR (KBr) ν<sub>max</sub> (cm<sup>-1</sup>): 3433 (br, OH), 2933 (CH), 1652 (C=O), 1374, 1041, 887; <sup>13</sup>C NMR: see Table I; <sup>1</sup>H NMR see Table II.

Acid hydrolysis of dracaenoside C. A solution of dracaenoside C (5 mg) in 1 M HCl (dioxane-H<sub>2</sub>O, 1:1, 1 ml) was heated at 100°C for 2 h. After removing the solution under reduced pressure, the residue was suspended with H<sub>2</sub>O and extracted with chloroform 3 times. The monosaccharide fraction was dried and dissolved in pyridine (5 ml). Then trimethylchlorosilane (0.5 ml) was added and the mixture was kept at room temperature for 30 min. After drying under reduced pressure, the reaction mixture was extracted with ether (0.5 ml), and the ether layer was analyzed by GC-MS. The retention time of L-rhamnose and D-glucose were 7.17 and 11.43 min, respectively.

*Dracaenoside D.*  $C_{39}H_{60}O_{17}$ , is an amorphous white powder,  $[\alpha]_{D}^{21} - 25.17$  (MeOH, *c* 0.4). Negative ion FAB-MS: m/z 799  $[M-H]^+$ ; 637  $(M^+-162-H)$ . Negative ion

TABLE I <sup>13</sup>C NMR spectral data of dracaenoside C (1) and D (2) (100 MHz, in pyridine-d<sub>5</sub>)

	С	D		С	D
1	37.61	37.60		Glc (inner)	Glc (inner)
2	30.24	30.12	1'	100.25	99.88
3	77.94	78.53	2'	78.14	77.91
4	39.10	38.80	3′	76.96	89.54
5	140.89	140.82	4′	77.94	69.59
6	122.37	122.41	5'	78.00	78.74
7	26.79	26.79	6′	61.29	62.45
8	34.13	34.13		Rha	Rha
9	43.69	43.65	1″	102.11	102.28
10	37.68	37.60	2″	72.59	72.53
11	19.87	19.84	3″	72.79	72.84
12	26.79	26.79	4″	74.17	74.15
13	52.40	52.39	5″	69.61	69.59
14	82.78	82.76	6"	18.72	18.76
15	41.79	41.77		Rha	Glc
16	143.22	143.16	1///	102.95	104.58
17	153.52	153.53	2′′′	72.59	74.99
18	20.51	20.49	3′′′	72.91	77.14
19	19.14	19.12	4‴	73.97	71.52
20	196.79	196.76	5′′′	70.46	77.67
21	27.13	27.12	6'''	18.57	62.45

TABLE II	<sup>1</sup> H NMR spectral data of	dracaenoside C (1)	) and D $(2)$ (400 MHz,	in pyridine- $d_5$ , J in Hz)

	С	D	
1	1ax,1.00, m; 1eq,1.70, m	1ax, 0.98, m; 1eq, 1.66, m	
2	2ax,1.82, m; 2eq, 2.03, m	2eq, 2.03, brd; 2ax, 1.76, m	
3	3.83, m	3.90, m	
4	4eq2.80, dd, J = 3.6, 9.6 Hz;	4eq, 2.80, m; 4ax, 2.76, m	
	4ax, 2.74, dd, $J = 11.0$ , 11.8 Hz		
6	5.27, brd, $J = 4.0  \text{Hz}$	5.45, brd, $J = 2.8$ Hz	
7	1.89, m; 2.55, m	1.85, m; 2.55, m	
8	1.98, m	1.91, m	
9	2.00, m	1.93, m	
11	1.62, m	1.62, m	
12	12ax, 2.59, m; 12eq, 2.45, m	12eq, 2.45, m; 12ax, 2.59, m	
15	15ax, 2.54, brd, $J = 14.84$ Hz;	15ax, 2.54, brd, $J = 15.0$ Hz	
	15eq, 2.51, brd, $J = 14.84$ Hz	15eq, 2.51, brd, J = 15.0 Hz	
16	6.72, brs	6.71, brs	
18	1.14, s	1.14, s	
19	1.12, s	1.13, s	
21	2.15, s	2.15, s	
	Glc (inner)	Glc (inner)	
1'	4.96, d, J = 6.4  Hz	4.90, d, $J = 6.4 \mathrm{Hz}$	
2'	4.38, m		
- 3'	4.19, m		
4'	4.20, m		
5′	3.62, brs		
6'	4.06, dd, J = 12.4, 3 Hz		
0			
	Rha	Rha	
1″	6.38, brs	6.38, brs	
2"	4.83, brs	4.82, brs	
3″	4.63, dd, $J = 9.30$ , 3.2 Hz	4.57, dd, $J = 9.32$ , 3 Hz	
4″	4.35, m	4.32, m	
5″	4.91, m	4.82, m	
6″	1.76, d, $J = 6.16 \mathrm{Hz}$	1.76, J = 6.16  Hz	
	Rha	Glc	
1///	5.84, brs	5.08, d, $J = 7.46 \mathrm{Hz}$	
2'''	4.67, brs		
3‴	4.53, dd, $J = 9.32$ , 3 Hz		
4‴	4.32, m		
5‴	4.90, m		
6'''	1.61, d, J = 6.04  Hz		

HR-FABMS: m/z 799.3772 ([M–H]<sup>+</sup>; calcd. 799.3752). UV (MeOH)  $\lambda_{max}$  (nm) (log  $\varepsilon$ ): 238. IR (KBr)  $\nu_{max}$  (cm<sup>-1</sup>): 3430 (br, OH), 2932, 1652 (C=O), 1373, 1048, 888. <sup>13</sup>C NMR: see Table I; <sup>1</sup>H NMR: see Table II.

Acid hydrolysis of dracaenoside D. Dracaenoside D (5 mg) was subjected to acid hydrolysis as described for dracaenoside C to give a mixture of monosaccharides. The monosaccharides were identified as D-glucose and L-rhamnose by direct TLC comparison with authentic samples and GC-MS analysis of the corresponding trimethylsilylated derivatives. The retention times of L-rhamnose and D-glucose were 7.17 and 11.43 min, respectively.

## Acknowledgements

The authors are grateful to the members of the analytical group in our institute for measurements of all spectra.

## Q-A. ZHENG AND C-R. YANG

#### References

- [1] Cai, X.T. and Xu, Z.F. (1979), Acta Bot. Yunnan 1, 1-9.
- [2] Wang, J.L., Li, X.C., Jiang, D.F. and Yang, C.Y. (1995), Acta Bot. Yunnan 17, 336-340.
- [3] Zhou, Z.H., Wang, J.L. and Yang, C.R. (1999), Chin. Trad. Herb Drugs 30, 801-804.
- [4] Zhou, Z.H., Wang, J.L. and Yang, C.R. (2001), Chin. Trad. Herb. Drugs 32, 484-486.
- [5] Zhou, Z.H., Wang, J.L. and Yang, C.R. (2001), Acta Pharm. Sin. 36, 200–204.
- [6] Lu, W.J., Wang, X.F., Chen, J.Y., Lu, Y., Wu, N., Kang, W.J. and Zheng, Q.T. (1998), Acta Pharm. Sin. 33, 755-758.
- [7] Yang, C.R. and Wang, Z. (1986), Acta Bot. Yunnan 8, 355-358.
- [8] Zheng, Q.A. and Yang, C.R. (2003), Chin. Chem. Lett., In press.
- [9] Agrawal, P.K., Jain, D.C., Gupta, R.K. and Thakur, R.S. (1985), Phytochemistry 24, 2479-2496.