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Two new steryl esters from the basidiomycete Tricholomopsis rutilans

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Abstract

Two new steryl esters with a polyhydroxylated ergostane-type nucleus, 3β , 5α -dihydroxy-(22E,24R)-ergosta-7,22-dien- 6β -yl oleate (**1**) and 3β , 5α -dihydroxy-(22E,24R)-ergosta-22-en-7-one- 6β -yl oleate (**2**), were isolated from the fruiting bodies of the basidiomycete *Tricholomopsis rutilans* along with three known sterols (**3**, **4**, and **5**). The structures of compounds **1** and **2** were established on the basis of spectroscopic means and chemical methods.

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Keywords: Tricholomopsis rutilans; Steryl ester; 3β , 5α -dihydroxy-(22E,24R)-ergosta-7,22-dien- 6β -yl oleate; 3β , 5α -dihydroxy-(22E,24R)-ergosta-22-en-7-one- 6β -yl oleate

1. Introduction

Tricholomopsis rutilans (Schaeff. Fr.) Singer is a widespread basidiomycete of the northern temperate zone, which fruits solitarily or in clusters on conifer stumps and logs and is occasionally found on wood chips [1]. A series of non-protein amino acids were previously reported from this fungus [2-5]. As a part of our search for naturally occurring secondary metabolites of higher fungi in the Yunnan Province [6–9], we have carried out a chemical investigation on this fungus and isolated two new steryl esters from its ethanol extract, 3β,5α-dihydroxy-(22E,24R)-ergosta-7,22-dien-6βyl oleate (1) and 3β , 5α -dihydroxy-(22E,24R)-ergosta-22en-7-one- 6β -yl oleate (2) (Fig. 1), along with three known sterols, (22E, 24R)-5 α , 8 α -epidioxyergosta-6, 22-dien-3β-ol (3), 3β-hydroxy-(22E,24R)-ergosta-5,8,22-trien-7one (4), and (22E, 24R)- $5\alpha, 6\alpha$ -epoxyergosta-8(14), 22-dien- 3β , 7α -diol (5). Previously, a few monohydroxylated steryl esters: ergosta-7,22-dien-3β-yl palmitate, ergosta-7,22-dien-3β-yl linoleate, 5α,8α-epidioxyergosta-6,22-dien-3B-yl linoleate, ergosta-7,24(28)-dien-3B-yl linoleate, and

ergosta-7-en-3β-yl linoleate, have been reported [10–12], but only one steryl ester with a polyhydroxylated ergostane-type nucleus, 3β,5α-dihydroxy-(22*E*,24*R*)-ergosta-7,22-dien-6βyl linoleate, has been isolated from the fungus *Catathelasma imperiale* [13]. This report deals with the isolation and the structural elucidation of these two new steryl esters (**1** and **2**).

2. Experimental

2.1. General

Melting points obtained on a XRC-1 apparatus are uncorrected. Optical rotations were measured on a Horiba SEPA-300 polarimeter. IR spectra were obtained with a Bio-Rad FTS-135 using KBr pellets. NMR spectra were recorded on Bruker AV-400 and Bruker DXR-500 spectrometers in CDCl₃ solvent with TMS as an internal standard. MS (EI, FAB) were recorded with a VG Autospec-3000 spectrometer. ESI and HR-ESI were recorded with an API QSTAR Pulsar 1 spectrometer. GC–MS was carried out on an Agilent 5973N instrument.

Silica gel (200–300 mesh) and pre-coated silica gel GF_{254} plates (Qingdao Marine Chemical Factory, PR China) were used for column chromatography and TLC, respectively.

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Fig. 1. Structures of 1 and 2.

Fractions were monitored by TLC, and spots were visualized by heating silica gel plates sprayed with 10% H₂SO₄ in ethanol.

2.2. Extraction and isolation

The fungus *T. rutilans* was collected on the Ailao Mountain of Yunnan Province, PR China, in July, 2003 and identified by Prof. *Zang Mu*, Kunming Institute of Botany, the Chinese Academy of Sciences. The voucher specimen was deposited in the Herbarium of the Kunming Institute of Botany, the Chinese Academy of Sciences.

Air-dried and crushed fruiting bodies of *T. rutilans* (2.5 kg) were extracted successively with ethanol four times at room temperature. The combined extraction was concentrated to dryness in vacuo to afford a syrup (180 g), which was subjected to silica gel column chromatography using petroleum ether/acetone (98:2, 95:5, and 90:10, v/v) to give fractions A (98:2, 3000 ml), B (95:5, 3000 ml), C (90:10, 3000 ml),

respectively. The residue (2 g) of fraction A was further purified by silica gel column chromatography using a gradient of petroleum ether/acetone (99:1, 98:2, v/v) to afford **3** (710 mg, petroleum ether/acetone 98:2, v/v). The residue (1 g) of fraction B was further purified by silica gel column chromatography using a gradient of petroleum ether/acetone (98:2, 97:3, v/v) to afford **1** (156 mg, petroleum ether/acetone 97:3, v/v). The residue (1.5 g) of fraction C was further purified by silica gel column chromatography using a gradient of petroleum ether/acetone (95:5, 94:6, 93:7, v/v) to afford **4** (148 mg, petroleum ester/acetone 95:5), **2** (21 mg, petroleum ester/acetone, 94:6), and **5** (103 mg, petroleum ester/acetone, 93:7).

2.2.1. 3β , 5α -Dihydroxy-(22E,24R)-ergosta-7,22dien- 6β -yl oleate (1)

Colorless, oily solid, $[\alpha]_D^{24} = -53.3^{\circ}$ (c 1.02, CHCl₃). TLC: (R_F 0.80, CHCl₃: MeOH 20:1). IR (KBr) ν : 3440, 2956, 2927, 2855, 1714, 1659, 1635, 1460, 1381, 1251, 1166, 1048, 1025, 970, 942 cm⁻¹. ¹H NMR and ¹³C NMR data are listed in Table 1. ESI-MS (pos.): 717 [M+Na]⁺, 395 [M+1 - C₁₈H₃₄O₂-H₂O]⁺, 377 [M+1 - C₁₈H₃₄O₂-2H₂O]⁺. HR-ESI-MS (pos.) calc. for C₄₆H₇₈O₄Na [M+Na]⁺: 717.5797; found: 717.5774.

2.2.2. Methanolysis of compound 1

A solution of **1** (25 mg) in methanol (10 ml) was treated with 0.1 M HCl (2 ml) at 75 °C for 18 h. The resultant mixture was extracted with petroleum ether to obtain the organic layer, which was dried (Na₂SO₄) and then analyzed by GC–MS.

2.2.3. 3β , 5α -Dihydroxy-(22E,24R)-ergosta-22en-7-one- 6β -yl oleate (**2**)

Colorless, oily solid, $[\alpha]_D^{26} = -63.7^\circ$ (c 0.46, CHCl₃). TLC: (R_F 0.45, CHCl₃: MeOH 20:1). IR (KBr) ν : 3426, 2956, 2926, 2855, 1731, 1639, 1461, 1381, 1237, 1164, 1034, 971, 944 cm ⁻¹. ¹H NMR and ¹³C NMR data are listed in Table 1. FAB-MS (pos.): 711 $[M+1]^+$, 429 $[M+1-C_{18}H_{34}O_2]^+$, 411 $[M+1-C_{18}H_{34}O_2-H_2O]^+$, 393 $[M+1-C_{18}H_{34}O_2-2H_2O]^+$. HR–ESI–MS (pos.) calc. for C₄₆H₇₈O₅Na $[M+Na]^+$: 733.5766; found: 733.5746.

3. Results and discussion

Compound **1** was obtained as a colorless, oily solid. The molecular formula of **1** was determined to be $C_{46}H_{78}O_4$ by positive ion HR–ESI–MS (calc. for $[M + Na]^+$: 717.5797; found: 717.5774). The ¹H NMR spectrum of **1** (Table 1) displayed singlets (δ 0.54, 1.00) for two tertiary methyl groups, doublets (δ 0.78, 0.79, 0.87, 0.98) for four secondary methyl groups, and a doublet (δ 5.18, J=5.0) typical for the 7-H of the trisubstituted olefinic proton of ergosta-7,22-dien-3 β ,5 α ,6 β -triol derivatives. A characteristic downfield doublet (δ 4.82, J=5.0) of 6 α -H caused by esterification

Table 1 ^{1}H and ^{13}C NMR data (CDCl_3) for 1 and 2

Position	1	1	2	2
1	1.22 (m)	32.33 (CH ₂)	1.25 (m)	32.06 (CH ₂)
2	1.77 (m)	30.40 (CH ₂)	1.83 (m)	30.13 (CH ₂)
3	4.03 (m)	67.09 (CH)	4.09 (m)	66.85 (CH)
4	2.00 (m), 1.81 (m)	39.17 (CH ₂)	1.99 (m), 1.74 (m)	40.42 (CH ₂)
5	_	75.10 (C)	_	78.94 (C)
6	4.82 (brd, 5.0)	73.34 (CH)	4.55 (s)	82.47 (CH)
7	5.18 (brd, 5.0)	114.09 (CH)	_	207.44 (C)
8	_	145.49 (C)	2.63 (t, 11.1)	46.28 (CH)
9	1.97 (m)	43.06 (CH)	1.78 (m)	47.66 (CH)
10	_	37.11 (C)	_	39.30 (C)
11	1.55 (m)	21.88 (CH ₂)	1.47 (m)	21.63 (CH ₂)
12	1.65 (m), 1.28 (m)	39.24 (CH ₂)	1.64 (m), 1.15 (m)	38.63 (CH ₂)
13	_	43.64 (C)	_	42.39 (C)
14	1.86 (m)	54.76 (CH)	1.50 (m)	48.10 (CH)
15	1.24 (m)	22.79 (CH ₂)	1.24 (m)	24.62 (CH ₂)
16	1.67 (m)	27.83 (CH ₂)	1.64 (m)	28.46 (CH ₂)
17	1.24 (m)	55.88 (CH)	1.14 (m)	54.93 (CH)
18	0.54 (s)	12.19 (CH ₃)	0.65 (s)	12.31 (CH ₃)
19	1.00 (s)	18.10 (CH ₃)	1.30 (s)	17.36 (CH ₃)
20	1.97 (m)	40.33 (CH)	1.99 (m)	39.94 (CH)
21	0.98 (d, 6.6)	21.04 (CH ₃)	0.98 (d, 6.6)	21.04 (CH ₃)
22	5.12 (dd, 15.2, 7.9)	135.33 (CH)	5.16 (m)	135.61 (CH)
23	5.18 (dd, 15.2, 7.3)	132.04 (CH)	5.16 (m)	131.89 (CH)
24	1.80 (m)	42.78 (CH)	1.84 (m)	42.82 (CH)
25	1.46 (m)	33.00 (CH)	1.44 (m)	33.07 (CH)
26	0.78 (d, 7.2)	19.57 (CH ₃)	0.78 (d, 6.7)	19.63 (CH ₃)
27	0.79 (d, 7.4)	19.88 (CH ₃)	0.80 (d, 6.7)	19.95 (CH ₃)
28	0.87 (d, 6.8)	17.55 (CH ₃)	0.88 (d, 6.8)	17.64 (CH ₃)
1'	_	173.42 (C)	_	172.20 (C)
2'	2.26 (t, 7.6)	34.62 (CH ₂)	2.32 (t, 7.8)	34.23 (CH ₂)
3'	1.58 (m)	24.87 (CH ₂)	1.60 (m)	24.82 (CH ₂)
4'-7', 12'-15'	1.22–1.29 (m)	29.02–29.64 (CH ₂)	1.22–1.30 (m)	29.04-29.75 (CH ₂)
8'	1.96 (m)	27.13 (CH ₂)	2.00 (m)	27.17 (CH ₂)
9'	5.30 (m)	129.65 (CH)	5.32 (m)	129.66 (CH)
10′	5.30 (m)	129.88 (CH)	5.32 (m)	130.03 (CH)
11'	1.96 (m)	27.13 (CH ₂)	2.00 (m)	27.17 (CH ₂)
16′	1.22–1.29 (m)	31.84 (CH ₂)	1.22–1.30 (m)	31.89 (CH ₂)
17'	1.22–1.29 (m)	22.61 (CH ₂)	1.22–1.30 (m)	22.67 (CH ₂)
18'	0.84 (t, 7.0)	14.04 (CH ₃)	0.85 (t, 7.0)	14.10 (CH ₃)

Assignments made on the basis of ¹H, ¹H-COSY, HSQC, and HMBC experiments.

suggested that the oleate moiety was clearly located at the 6β position of the sterol nucleus [13]. These data were compatible with those reported for cerevisterol ((22E, 24R)-ergosta-7,22-dien-3 β ,5 α ,6 β -triol) and its derivatives [14,15,19]. Furthermore, the ¹H NMR spectrum of **1** also showed the characteristic signals at $\delta 0.84$ (t, J = 7.0) for a terminal methyl group, at $\delta 2.26$ (t, J = 7.6) for a methylene group in α -position to an ester function, at δ 5.30 (m) for two olefinic protons, suggesting a disubstituted C=C bond, and other signals at δ 1.96, 1.58, 1.22–1.29 (overlapped) all related to a monounsaturated long-chain fatty-acid ester moiety. Following methanolysis, 1 yielded the unsaturated fatty-acid methyl ester, which was identified as methyl oleate ((9Z)-octadec-9-enoic acid methyl ester) by GC-MS analysis, and this was consistent with the ESI-MS of 1, which exhibited characteristic fragment ion peaks at m/z 395 $[M+1-C_{18}H_{34}O_2-H_2O]^+$ and 377 $[M+1-C_{18}H_{34}O_2-2H_2O]^+$. Supporting evidence was also obtained from the ¹³C NMR spectrum (Table 1), which

had typical signals at δ 173.42 (C-1'), 34.62 (C-2'), 24.87 (C-3'), 27.13 (C-8' and C-11'), 129.65 (C-9'), and 129.88 (C-10'), for the oleate moiety with (*Z*)-configuration. The characteristic upfield values for both the allylic C-atoms C-8' and C-11' were typical for *cis*-olefins as compared to *trans*-olefins [16]. The linked position of the oleate moiety was also reasonably confirmed by the HMBC spectrum (Fig. 2), which clearly displayed the strong correlation peak between 6α -H (δ 4.82) and C-1' (δ 173.42). The stereochemistry of the side chain was determined by comparison of the ¹H and ¹³C NMR data of **1** with those of (22*E*,24*R*)-methyl- Δ^{22} -sterol side chain [17]. From all of these data mentioned above, the structure of **1** was, therefore, determined to be 3β , 5α -dihydroxy-(22*E*,24*R*)-ergosta-7,22-dien-6 β -yl oleate.

Compound 2 was also obtained as a colorless, oily solid. A quasi-molecular ion peak at m/z 711 $[M+1]^+$ and a series of characteristic fragment ion peaks at m/z



Fig. 2. Selected HMBC correlations of 1 and 2.

 $429 [M+1-C_{18}H_{34}O_2]^+, 411[M+1-C_{18}H_{34}O_2-H_2O]^+,$ 393 $[M+1-C_{18}H_{34}O_2-2H_2O]^+$ were given in positive ion FAB-MS. The positive ion HR-ESI-MS of 2 exhibited a molecular formula of $C_{46}H_{78}O_5$ (calc. for $[M + Na]^+$: 733.5766; found: 733.5746), which was consistent with the analysis of its ¹³C NMR spectrum. The ¹H NMR and ¹³C NMR (Table 1) spectra of 2 showed very similar signals compared with those of 1, indicating 2 also possessed the same steryl ester skeleton as 1, and the only difference between 2 and **1** was that the Δ^7 C=C group of **1** was replaced by a keto group in 2. The presence of the 7-keto function also led to the appearance of the 8 β -H signal further downfield at δ 2.63 (t, J = 11.1) in the ¹H NMR spectrum 2. By comparing this data of the sterol nucleus with that in the literature [18], 2 was inferred to be a derivative of 3β , 5α , 6β -trihydroxy-(22E,24R)-ergosta-22-dien-7-one, and the oleate moiety was also assigned to the 6^β position. From all of these data mentioned above, the structure of 2 was, therefore, determined to be 3β , 5α -dihydroxy-(22E,24R)-ergosta-22-en-7-one-6 β yl oleate.

Comparison of the physicochemical properties with the reported data allowed for the identification of compounds **3**, **4**, and **5**, isolated from the same fungus, as $(22E,24R)-5\alpha,8\alpha$ -epidioxyergosta-6,22-dien-3\beta-ol (3)

[19], 3β -hydroxy-(22*E*,24*R*)-ergosta-5,8,22-trien-7-one (**4**) [18], (22*E*,24*R*)-5 α ,6 α -epoxyergosta-8(14),22-dien-3 β ,7 α -diol (**5**) [19], respectively.

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