

## A new *ent*-kaurane diterpenoid from *Isodon phyllostachys*

Ai-Jun Hou, Hui Yang, Bei Jiang, Qin-Shi Zhao,  
Zhong-Wen Lin, Han-Dong Sun\*

Laboratory of Phytochemistry, Kunming Institute of Botany, Academia Sinica, Kunming 650204,  
Yunnan, PR China

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### Abstract

A new *ent*-kaurane diterpenoid, phyllostachysin C (**1**), together with five known compounds, sculponeatins B and C, nodosin, ursolic acid and 2 $\alpha$ -hydroxyursolic acid, were isolated from the leaves of *Isodon phyllostachys*. The structure of **1** was elucidated on the basis of its spectral properties. © 2000 Published by Elsevier Science B.V. All rights reserved.

*Keywords:* *Isodon phyllostachys*; *ent*-Kauranoids; Phyllostachysin C

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### 1. Introduction

*Isodon phyllostachys* (Diels) Hara (Lamiaceae), distributed widely in the north-west of Yunnan and south-west of Sichuan of China, has been used as antiphlogistic or an antibiotic agent in folk medicine. We reported previously two new *ent*-kauranoids, phyllostachysin A [1] and B [2], and further examination of the diterpenoids in this plant collected in Sichuan province led to the isolation of a new compound, phyllostachysin C (**1**), and five known isolates, sculponeatins B and C [3], nodosin [4], ursolic acid and 2 $\alpha$ -hydroxyursolic acid. We now present the isolation and characterization of **1**.

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\* Corresponding author. Tel.: +86-871-5150660; fax: +86-871-5216343.  
E-mail address: hdsun@mail.kib.ac.cn (H. Sun).

## 2. Experimental

### 2.1. Plant material

The leaves of *I. phyllostachys* were collected in Muli of Sichuan province in August 1987, and air-dried. The identity of plant material was verified by Prof. Xi-Wen Li, and a voucher specimen (870831-KIB) is deposited in the Herbarium of the Department of Taxonomy, Kunming Institute of Botany, Academia Sinica.

### 2.2. Extraction and isolation

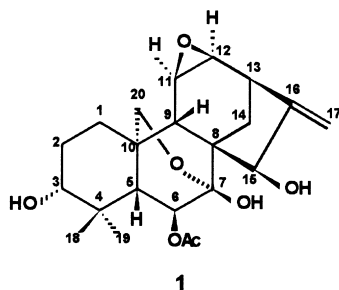
The dried and powdered leaves (2.2 kg) were extracted with Et<sub>2</sub>O and concentrated. The residue was dissolved in MeOH and decolorized by activated charcoal. After the removal of MeOH, the extract (92 g) was subjected to Si-gel CC eluting with petrol–chloroform and chloroform–acetone. Each fraction was further purified by recrystallization to yield **1** (550 mg), sculponeatins B (1.3 g) and C (3.3 g), nodosin (2.5 g), ursolic acid (115 mg) and 2 $\alpha$ -hydroxyursolic acid (50 mg).

*Phyllostachysin C* (**1**). C<sub>22</sub>H<sub>30</sub>O<sub>7</sub>, colorless needles, m.p. 203–205°C, [ $\alpha$ ]<sub>D</sub><sup>22</sup>: –102.0° (c 0.55, C<sub>5</sub>H<sub>5</sub>N); IRmax (KBr): 3550, 3520, 3330, 1740, 1655, 1225, 1100, 1055, 1038, 1020 cm<sup>-1</sup>; <sup>1</sup>H-NMR (400 MHz, Py-*d*<sub>5</sub>): 1.63 (1H, *m*, H-1 $\alpha$ ), 1.82 (1H, *overlap*, H-1 $\beta$ ), 1.86 (2H, *m*, H<sub>2</sub>-2 $\alpha$  and  $\beta$ ), 3.50 (1H, *br d*, *J* 9.2 Hz, H-3 $\beta$ ), 1.55 (1H, *d*, *J* 4.2 Hz, H-5 $\beta$ ), 5.76 (1H, *d*, *J* 4.2 Hz, H-6 $\alpha$ ), 2.57 (1H, *br s*, H-9 $\beta$ ), 3.19 (1H, *t*, *J* 4.3 Hz, H-11 $\alpha$ ), 3.22 (1H, *t*, *J* 4.3 Hz, H-12 $\alpha$ ), 2.96 (1H, *br s*, H-13 $\alpha$ ), 2.56 (1H, *d*, *J* 12.1 Hz, H-14 $\alpha$ ), 2.07 (1H, *dd*, *J* 4.6, 12.1 Hz, H-14 $\beta$ ), 5.01 (1H, *br s*, H-15 $\alpha$ ), 5.54 (1H, *br s*, H-17a), 5.25 (1H, *br s*, H-17b), 1.20 (3H, *s*, Me-18), 1.30 (3H, *s*, Me-19), 4.20 (1H, *ABd*, *J* 9.2 Hz, H-20a), 4.44 (1H, *ABd*, *J* 9.2 Hz, H-20b), 2.19 (3H, *s*, Me-OAc); <sup>13</sup>C-NMR (100 MHz, Py-*d*<sub>5</sub>): 28.4 (C-1), 27.9 (C-2), 77.2 (C-3), 40.1 (C-4), 58.4 (C-5), 74.4 (C-6), 96.4 (C-7), 51.4 (C-8), 41.1 (C-9), 37.5 (C-10), 50.8 (C-11), 53.6 (C-12), 38.3 (C-13), 26.6 (C-14), 74.7 (C-15), 153.4 (C-16), 109.5 (C-17), 27.8 (C-18), 15.9 (C-19), 67.9 (C-20), 169.2 (C-OAc), 21.5 (C-OAc); EI-MS (70 eV) *m/z*: 406 [M]<sup>+</sup> (1), 346 [M-AcOH]<sup>+</sup> (35), 328 [M-AcOH-H<sub>2</sub>O]<sup>+</sup> (8), 318 (3), 300 (4), 282 (10), 167 (100), 149 (67); HREI-MS *m/z*: 406.1986 (calcd for C<sub>22</sub>H<sub>30</sub>O<sub>7</sub>, 406.1992).

## 3. Results and discussion

The ethereal extract from the leaves of *I. phyllostachys* was subjected to Si-gel column chromatography eluting with petrol–chloroform and chloroform–acetone. Each fraction was further purified repeatedly to yield phyllostachysin C (**1**) and five known compounds.

Phyllostachysin C (**1**) was obtained as colorless needles and had a molecular formula of C<sub>22</sub>H<sub>30</sub>O<sub>7</sub> deduced from HREI-MS (obsd 406.1986, calcd 406.1992) and the analysis of its <sup>13</sup>C-NMR spectrum. It contained an acetoxy group [<sup>1</sup>H-NMR:  $\delta$  2.19 (3H, *s*); <sup>13</sup>C-NMR:  $\delta$  169.2 (*s*) and 21.5 (*q*)], *exo*-methylene group [IRmax (KBr): 1655 cm<sup>-1</sup>; <sup>1</sup>H-NMR:  $\delta$  5.25 and 5.54 (each 1H, *br s*); <sup>13</sup>C-NMR:  $\delta$  109.5 (*t*)



and 153.4 (*s*), an oxygen-bearing methylene [ $^1\text{H-NMR}$ :  $\delta$  4.20 and 4.44 (each 1H, *ABd*,  $J = 9.2$  Hz);  $^{13}\text{C-NMR}$ :  $\delta$  67.9 (*t*)], and a ketalic group [ $^{13}\text{C-NMR}$ :  $\delta$  96.4 (*s*)]. Moreover, signals due to two tertiary methyl groups, three methylenes, eight methines including five oxygenated ones and three quaternary carbons were also observed in the  $^{13}\text{C-NMR}$  spectrum of **1**. These facts, with consideration of the structures of diterpenoids isolated so far from the *Isodon* genus [5,6], established the basic skeleton of **1** as 7 $\beta$ -hydroxy-7 $\alpha$ , 20-epoxy-*ent*-kaur-16-ene.

According to the  $^1\text{H}$ - $^1\text{H}$  COSY and HMQC experiments, all protons and related carbons were assigned, which provided information on the five oxygenated methines. The signals at  $\delta$  3.50 (1H, *br d*,  $J = 9.2$  Hz, H-3) and  $\delta$  77.2 (*d*, C-3), and the downfield shifts of C-2 and C-4 disclosed that a hydroxyl group was located at C-3. The acetoxy group was at C-6 through the signals of H-6 ( $\delta$  5.76, 1H, *d*,  $J = 4.2$  Hz) and C-6 ( $\delta$  74.4, *d*). The signals at  $\delta$  5.01 (1H, *s*, H-15) and  $\delta$  74.7 (*d*, C-15) were indicative of a 15-hydroxyl group. The signals at  $\delta$  3.19 (1H, *t*,  $J = 4.3$ ), 3.22 (1H, *t*,  $J = 4.3$  Hz),  $\delta$  50.8 (*d*) and 53.6 (*d*) were ascribable to H-11, H-12, C-11 and C-12, respectively, which showed the presence of a 11, 12-epoxide ring.

The following NOESY cross peaks: H-3 $\beta$  with H-1 $\beta$  and H-5 $\beta$ ; H-6 $\alpha$  with Me-18, 19; H-15 $\alpha$  with H-14 $\beta$ , indicated that 3-OH, 6-OAc and 15-OH possessed  $\alpha$ -,  $\beta$ - and  $\beta$ -orientation, respectively. The upfield shifts of C-19, 18 and C-9 owing to the  $\gamma$ -steric compression effects of 3 $\alpha$ - and 15 $\beta$ -OH also supported partial results above. Additionally, the  $\beta$ -configuration of the 11, 12-epoxide ring was decided by the NOE effects of H-11 $\alpha$  with H-14 $\alpha$  and H-1 $\alpha$ , and the peak form of H-12 (*t*,  $J = 4.3$  Hz). Thus, the structure of phyllostachysin C (**1**) was identified as shown.

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