

Three New Triterpenoids Containing Four-Membered Ring from the Fruiting Body of *Ganoderma sinense*

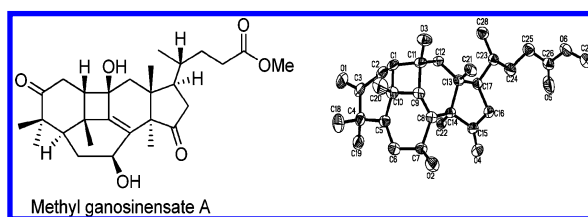
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ABSTRACT



Methyl ganosinensate A (**1**), ganosinensic acid A (**1a**), and ganosinensic acid B (**2**), three new triterpenoids with an unusual four-membered ring skeleton produced by a bond across C-1 to C-11, were isolated from the fruiting body of *Ganoderma sinense*. Their structures were established on the basis of extensive spectroscopic methods, including 1D and 2D NMR techniques, and methyl ganosinensate A was confirmed by X-ray crystallographic analysis.

The genus *Ganoderma* has been used as folk medicine since ancient time. Among them, the well-known species of *Ganoderma lucidum* has been widely used as a health supplement, especially in China, Japan, and Korea. Therefore, most research has focused on *G. lucidum*, on its chemical and biologically active constituents. The species is known to be prolific producers of lanostane-type triterpenoids, and over 100 such triterpenoids have been isolated and characterized,¹ some of which are antiandrogen,² antitumor,³ anti-HIV-1,⁴ and anti-inflammatory⁵ constituents.

In our previous research on this genus, we reported two novel 3,4-*seco*-trinorlanostane triterpenoids⁶ and a new 18(13→12 β) *abeo*-lanostadiene triterpenoid⁷ from *G. fornicatum*.

To further discover structurally diverse and biologically significant compounds from the *G. sinense*, we examined its fruiting body, which led to the isolation of three new triterpenoids, methyl ganosinensate A (**1**), ganosinensic acid A (**1a**), and ganosinensic acid B (**2**), with an unusual four-membered ring produced by linkage of C-1 with C-11. These

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compounds provide us a new insight into the biosynthesis of triterpenoids with the lanostane skeleton. Here we describe the structure elucidation and propose a biosynthetic route.

The dried and powdered fruiting body of *G. sinense* (dry weight 50 kg) was extracted with MeOH by refluxing and concentrated in vacuo to give a crude extract (5 kg), which was then partitioned between H₂O and EtOAc. The EtOAc fraction (1.5 kg) was repeatedly chromatographed on silica gel, RP-18, and Sephadex LH-20 (MeOH) to yield methyl ganosinensate A (**1**, 12 mg), mixed **1a**, and **2** (31 mg). Further purification with HPLC led to the isolation of ganosinensic acid B (**2**, 13 mg) and ganosinensic acid A (**1a**, 8 mg) (for details, see the Supporting Information).

Methyl ganosinensate A (**1**)⁸ was isolated as colorless cubes. Its molecular formula, C₂₈H₄₀O₆, was established on the basis of HRESIMS. The ¹H NMR showed the presence of six methyl signals at δ_{H} 1.68 (s), 1.37 (s), 1.31 (s), 1.12 (s), 1.02 (s), and 0.94 (d, $J = 6.4$ Hz) and a methoxyl signal at δ_{H} 3.64 (s) (Table 1). The ¹³C and DEPT NMR spectra (Table 2) displayed 28 carbons, including 10 quaternary carbons (two sp² carbons at δ_{C} 136.4 and 154.7, one oxygenated sp³ carbon at δ_{C} 83.5, and three carbonyls at δ_{C} 216.5, 215.0, and 174.1), five methines (one oxymethine at δ_{C} 68.3), six methylenes, and seven methyls. The data suggested that **1** was close to the structure of lucidenic acid A,⁹ except for those of C-1 and C-11. Interestingly, the chemical shift of C-1 is no more than 40 ppm in a normal lanostane-type triterpenoid, while the chemical shift of C-1 in **1** is 57 ppm; the chemical shift of C-11 (83.5) is also quite different from the general data (δ_{C} 198–200). Furthermore, there was absence of one carbonyl and no adding of one unsaturated bond of **1**, but **1** and lucidenic acid A still retain the same degrees of unsaturation, and it could be deduced that **1** includes five rings instead of the usual four rings. This was further supported by HMBC correlations (Figure 1) observed from signals of H-1 (δ_{H} 2.77) to C-2 (δ_{C} 36.5), C-9 (δ_{C} 154.7), C-11 (δ_{C} 83.5), and C-19 (δ_{C} 18.8), as well as signals from H-12 (δ_{H} 2.13) to C-9, C-11, C-14 (δ_{C} 62.3), and C-18 (δ_{C} 17.7). These correlations allowed us to establish a four-membered ring E system, which could well explain the odd chemical shift of C-1 and C-11 and the degrees of unsaturation mentioned above. The five methyls were assigned by the key HMBC correlations observed from H-5 (δ_{H} 2.02) to C-28 (δ_{C} 27.1) and C-29 (δ_{C} 18.9), from H-12 to C-18, from H-1 to C-19, from H-30 (δ_{H} 1.31, s) to C-13, and from H-21 (δ_{H} 0.94, d, $J = 6.4$ Hz) to C-20 (δ_{C} 35.7) and C-17 (δ_{C} 46.8). Correlations from the signals H-28, H-29 to C-3 (δ_{C} 216.5); H-7 (δ_{H} 5.10) to C-8 (δ_{C} 136.4), C-9; H-30 to C-15 (δ_{C} 215.0), confirmed that the positions of two ketones and a hydroxyl groups were at C-3, C-15, and C-7, respectively. Comprehensive analysis of the ¹H–¹H COSY spectrum of **1** allowed the establishment of three

(8) Methyl ganosinensate A (**1**): colorless cubes (petroleum ether:acetone 10:1); mp 215–217 °C; [α]_D²⁵ = +171.3 (c 0.1, CH₃OH); negative FAB m/z 471 [M – H][–]; HRESIMS m/z [M + Na]⁺ 495.2728 (calcd for C₂₈H₄₀O₆Na, 495.2722); UV (CH₃OH) λ_{max} (log ϵ) 215 (3.55) nm; IR (KBr) ν_{max} 3400, 2930, 1717, 1695, 1368, 1257, 1156, 1043 cm^{–1}. NMR can be found in Table 1 and 2.

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Table 1. ¹H NMR Assignments of **1**, **1a**, and **2**^a

| no. | δ_{H} (J in Hz) mult | | |
|-------------|---------------------------------------|-------------------|--------------------|
| | 1 | 1a | 2 |
| 1 | 2.77 m | 2.76 m | 2.77 m |
| 2 α | 3.01 t (12.3) | 3.01 t (12.3) | 2.99 t (12.3) |
| 2 β | 2.50 dd (12.6, 7.5) | 2.49 (overlapped) | 2.41 (overlapped) |
| 5 | 2.02 m | 2.01 m | 2.01 m |
| 6 α | 2.33 m | 2.34 m | 2.35 m |
| 6 β | 2.02 m | 2.02 m | 2.04 m |
| 7 | 5.10 m | 5.11 m | 5.12 dd (8.5, 4.4) |
| 12 | 2.13 m | 2.15 m | 2.16 m |
| 16 α | 2.83 m | 2.87 m | 2.85 m |
| 16 β | 2.15 m | 2.24 m | 2.16 m |
| 17 | 2.10 m | 2.14 m | 2.26 m |
| 18 | 1.37 s | 1.38 s | 1.39 s |
| 19 | 1.68 s | 1.68 s | 1.67 s |
| 20 | 1.58 m | 1.70 m | 2.37 m |
| 21 | 0.94 d (6.4) | 0.99 d (6.4) | 1.11 (overlapped) |
| 22 | 1.81 m, 1.38 m | 2.62 m, 1.98 m | 2.55 m, 2.44 m |
| 23 | 2.43 m, 2.31 m | 2.50 m, 1.51 m | |
| 24 | | | 3.11 m, 2.62 m |
| 25 | | | 3.3 m |
| 27 | | | 1.35 d (7.5) |
| 28 | 1.02 s | 1.00 s | 1.02 s |
| 29 | 1.12 s | 1.12 s | 1.12 (overlapped) |
| 30 | 1.31 s | 1.30 s | 1.30 s |
| OMe | 3.64 s | | |

^a Measured at 500 MHz in C₅D₅N.

structural fragments, as drawn with bold lines in Figure 1. The structure of **1** was further confirmed by X-ray crystallographic analysis.¹⁰

The relative configuration of **1** was determined by the ROESY spectrum (Figure 1), in which the correlations of H-5/H-7 and H-1/H₃-19 indicated that 7-OH and H-1 was in β -orientation. From X-ray crystallographic analysis, H₃-19 and 11-OH have the same relative configuration, so 11-OH was in β -orientation. The conformation of **1** in solution as established by the ROESY spectrum was in good agreement with that in the solid state, as determined by X-ray study (Figure 2).

Ganosinensic acid A (**1a**),¹¹ obtained as colorless needles, was determined as C₂₇H₃₈O₆ on the basis of HRESIMS. Comparing signals of ¹³C spectra with **1**, the only difference is the disappearance of a methoxyl group. So **1a** is supposed

(10) Crystallographic data for **1**: C₂₈H₄₀O₆, $M = 472.60$, orthorhombic, space group $P2_12_12_1$, $a = 7.6814$ (1) Å, $b = 12.1284$ (1) Å, $c = 27.7022$ (3) Å, $V = 2580.82$ (5) Å³, $Z = 4$, $d = 1.216$ g/cm³, crystal dimensions 0.10 × 0.10 × 0.20 mm was used for measurements on a MAC DIP-2030K diffractometer with a graphite monochromator (ω - 2θ scans, $2\theta_{\text{max}} = 134.52^\circ$), Mo K α radiation. The total number of independent reflections measured was 3692, of which 3439 were observed ($|I| \geq 2\sigma(I)$). Final indices: $R_1 = 0.0406$, $wR_2 = 0.1111$ ($w = 1/\sigma(I)^2$), $S = 1.035$. The crystal structure of **1** was solved by direct method SHELXS-97 (Sheldrick, G. M. University of Gottingen: Gottingen, Germany, 1997) and the full-matrix least-squares calculations. Crystallographic data for the structure of **1** have been deposited in the Cambridge Crystallographic Data Centre (deposition number: CCDC 755772). Copies of these data can be obtained free of charge via the Internet at www.ccdc.cam.ac.uk/conts/retrieving.html (or from the Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB21EZ, U.K.; fax (+44) 1223–336–033; or deposit@ccdc.cam.ac.uk)

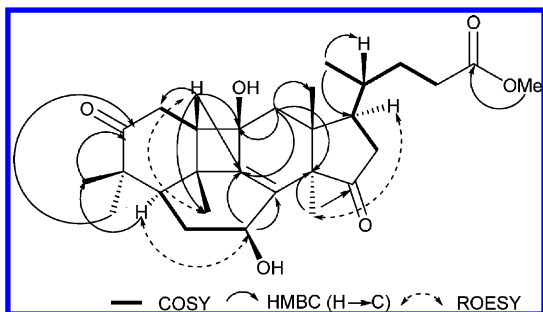


Figure 1. Key COSY, HMBC, and ROESY correlations of **1**.

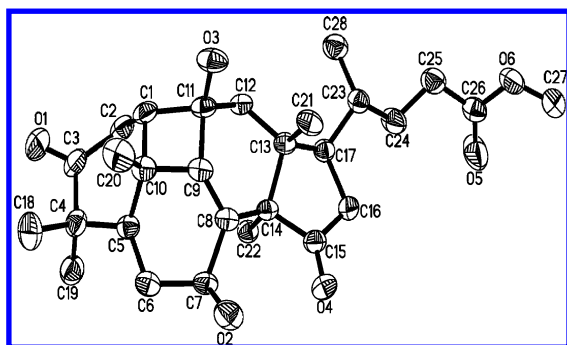


Figure 2. Single-crystal X-ray structure of **1**.

to possess a hydroxyl group at C-24, and this was confirmed by HMBC experiment.

Ganosinensic acid B (**2**)¹² was determined as $C_{30}H_{42}O_7$ on the basis of a combination of FABMS and HRESIMS. Inspection of spectral data of **2** revealed the presence of the same four-membered ring as in **1**, including the characteristic data of C-1 (δ_C 57.0, d) and C-11 (δ_C 83.5, s). This analysis suggested that **2** was also a triterpenoid with a four-membered ring. In the ^{13}C NMR spectrum, most of the signals in **2** were superimposable over those of **1**, except for the side chain moiety, where the former possesses an eight-carbon group and the latter possesses a five-carbon group. The ^{13}C NMR signals of the side chain of **2** were in good agreement with those of ganoderic acid C.⁹ This was further supported by HMBC correlations observed from signals of H-27 (δ_H 1.11, d, $J = 8.2$ Hz) to C-24, C-25, and C-26, signals from H-25 (δ_H 2.37) to C-23, C-24, C-26, and C-27, as well as a proton spin system deduced from 1H – 1H COSY correlations, H-24/H-25/H-27.

(11) Ganosinensic acid A (**1a**): colorless needles (petroleum ether/acetone 3:1); mp 206–208°; $[\alpha]_D^{25} = +167.2$ (c 0.1, CH_3OH); negative FAB m/z 457 $[M - H]^-$; HRESIMS m/z 481.2564 $[M + Na]^+$ (calcd for $C_{27}H_{38}O_6Na$, 481.2566); UV (CH_3OH) λ_{max} (log ϵ) 214 (3.68) nm; IR (KBr) ν_{max} 3350, 2950, 1720, 1675, 1380, 1270, 1160, 1050 cm^{-1} . NMR can be found in Table 1 and 2.

(12) Ganosinensic acid B (**2**): colorless powders; $[\alpha]_D^{25} = +140.8$ (c 0.1, CH_3OH); negative FAB m/z 513 $[M - H]^-$; HRESIMS m/z 537.2824 $[M + Na]^+$ (calcd for $C_{30}H_{42}O_7Na$, 537.2828); UV (CH_3OH) λ_{max} (log ϵ) 216 (3.96) nm; IR (KBr) ν_{max} 3410, 2947, 1740, 1685, 1370, 1222, 1161, 1038 cm^{-1} . NMR can be found in Tables 1 and 2.

Table 2. ^{13}C NMR and DEPT Assignments of **1**, **1a**, and **2**^b

| | 1 | 1a | 2 |
|------------------|----------|-----------|----------|
| 1 | 57.0 d | 57.0 d | 57.0 d |
| 2 | 36.5 t | 36.5 t | 36.5 t |
| 3 | 216.5 s | 216.5 s | 216.4 s |
| 4 | 47.0 s | 47.0 s | 47.1 s |
| 5 | 55.4 d | 55.4 d | 55.5 d |
| 6 | 29.4 t | 29.4 t | 29.4 t |
| 7 | 68.3 d | 68.4 d | 68.3 d |
| 8 | 136.4 s | 136.3 s | 136.3 s |
| 9 | 154.7 s | 154.7 s | 154.8 s |
| 10 | 48.6 s | 48.6 s | 48.6 s |
| 11 | 83.5 s | 83.6 s | 83.5 s |
| 12 | 37.4 t | 37.4 t | 37.4 t |
| 13 | 45.6 s | 45.6 s | 45.7 s |
| 14 | 62.3 s | 62.4 s | 62.4 s |
| 15 | 215.0 s | 215.3 s | 214.8 s |
| 16 | 41.0 t | 41.1 t | 41.0 t |
| 17 | 46.8 d | 46.9 d | 46.6 d |
| 18 | 17.7 q | 17.7 q | 17.7 q |
| 19 | 18.8 q | 18.8 q | 18.8 q |
| 20 | 35.7 d | 35.9 d | 32.6 d |
| 21 | 18.6 q | 18.7 q | 20.3 q |
| 22 | 31.4 t | 31.9 t | 49.8 t |
| 23 | 31.1 t | 31.8 t | 208.9 t |
| 24 | 174.1 s | 176.1 s | 46.9 t |
| 25 | | | 35.7 t |
| 26 | | | 178.4 s |
| 27 | | | 17.7 q |
| 28 | 27.1 q | 27.1 q | 27.1 q |
| 29 | 18.9 q | 18.9 q | 18.9 q |
| 30 | 20.6 q | 20.7 q | 20.6 q |
| OCH ₃ | 51.4 q | | |

^b Measured at 100 MHz in C_5D_5N .

The relative stereochemistry of **2** was constructed from the ROESY spectrum (Figure 3), together with 1D NMR data comparison with those of **1** (Tables 1 and 2). The configurations of H-7 and H-1 were deduced to be α -H and β -H, respectively.

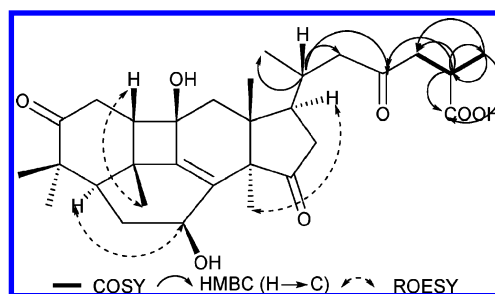
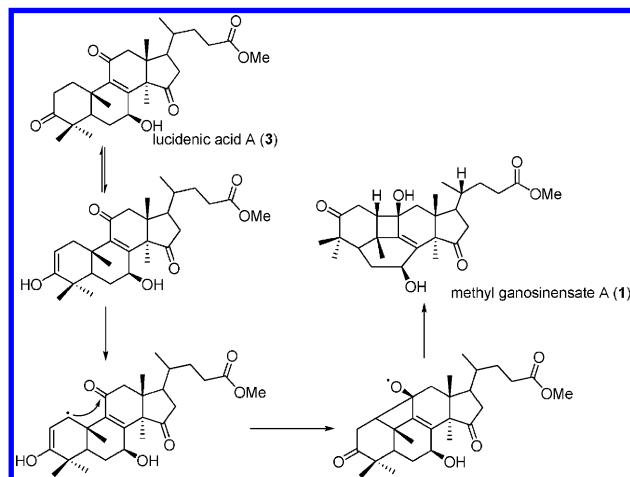


Figure 3. Key COSY, HMBC, and ROESY correlations of **2**.

According to the literature, methyl ganosinensate A (**1**), ganosinensic acid A (**1a**), and ganosinensic acid B (**2**) have a complex triterpenoid skeleton without precedent among known natural products. Especially, the four-membered ring

Scheme 1. Proposed Biogenetic Pathway for **1**



framework produced by a bond across C-1 to C-11 provides us a new insight into the biosynthesis of lanostane-type triterpenoid. The co-occurrence of the three triterpenoids and normal lanostane-type triterpenoids within the same plant raises the possibility that triterpenoids with a four-membered ring result from a further modification of an existing metabolite. Because the core skeleton of the three triterpenoids is still preserved in lucidenic acid A (**3**), and **3** is widely distributed in the genus *Ganoderma*, it is concluded that **3**

should occur early in Scheme 1 as the biogenetic origin. Herein, we propose a new possible biosynthetic route of methyl ganosinensate A (**1**) in Scheme 1.

Considering some triterpenoids are known to have anti-tumor activity,^{13,14} the antitumor activities of compounds **1**, **1a**, and **2** were evaluated with the HL-60, SMMC-7721, A-549, MCF-7, and SW480 cell lines; however, neither of them showed significant inhibitory activity ($IC_{50} > 40 \mu\text{g/mL}$ for the five cell lines).

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Supporting Information Available: Experimental procedures, 1D and 2D NMR spectra and crystallographic data of methyl ganosinensate A (**1**), ganosinensic acid A (**1a**), and B (**2**). This material is available free of charge via the Internet at <http://pubs.acs.org>.

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