

## Two new epimeric isopavine N-oxides from *Meconopsis horridula* var. *racemosa*

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### Abstract

Two new epimeric isopavine N-oxides, amuresinine N-oxide A (**1**) and B (**2**), were isolated from *Meconopsis horridula* var. *racemosa*. Their structures were elucidated by spectroscopic methods. © 2001 Elsevier Science B.V. All rights reserved.

**Keywords:** *Meconopsis horridula* var. *racemosa*; Amuresinine N-oxides

### 1. Introduction

*Meconopsis*, a relatively large genus in the family Papaveraceae, is mainly distributed discontinuously in both East Asia and West Europe. There are some species in the north-west Yunnan Province in China. *Meconopsis* are an important and traditional Tibetan plant medicine and have long been used by Tibetan people. Many isoquinoline alkaloids were isolated from plants of this genus and some of them have shown bioactivities such as anti-inflammatory and analgesic activity [1–3]. We describe here two new epimeric isopavine N-oxides from *Meconopsis horridula* var. *racemosa* (Maxim.) Prain.

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## 2. Experimental

### 2.1. Plant material

*M. horridula* var. *racemosa* whole plants were collected on mountains, at altitudes between 3350 and 4800 m, in Li jiang county and Di qin Tibetan Autonomy Prefecture, north-west Yunnan Province, China, from August to September 1998. A voucher specimen has been deposited in the Department of Ethnobotany, Kunming Institute of Botany, Chinese Academy of Sciences.

### 2.2. Extraction and isolation

Air-dried, powdered plant (1 kg) was extracted by hot 95% EtOH. The extract was evaporated, 5% HCl was added to pH 3–4, and CH<sub>3</sub>Cl extraction (I) was performed. The aqueous layer was added with 5% Na<sub>2</sub>CO<sub>3</sub> to pH 11–12 and then extracted with CH<sub>3</sub>Cl (II). The residue of the latter (400 mg) was Si-gel CC eluting with CHCl<sub>3</sub>/CH<sub>3</sub>OH gradient (from 20:1 to 8:1) to afford a mixture which was separated by HPLC (Bondapack C18) eluting with 1 ml/min CH<sub>3</sub>OH–(NH<sub>4</sub>)<sub>2</sub>CO<sub>3</sub> (0.01 M) 45:55 to afford **1** (5 mg) and **2** (7 mg).

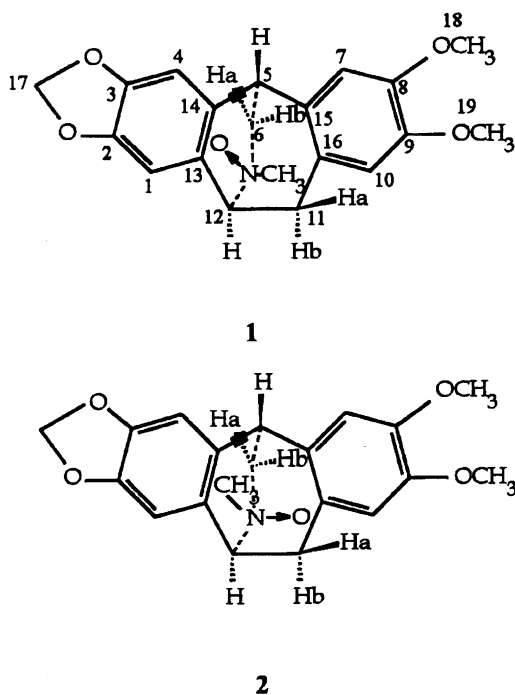
**Compound 1.**  $[\alpha]_D^{27}$  –101.03° (CHCl<sub>3</sub>, *c* 0.30); FAB-MS *m/z*: 356[M + H]<sup>+</sup> (100), 340(24), 295(22), 188(3); HR-FABMS *m/z*: 356.1547[M + H]<sup>+</sup> (calc. for C<sub>20</sub>H<sub>21</sub>NO<sub>5</sub> + H: 356.1498); EI-MS *m/z*: 355[M]<sup>+</sup> (2), 353[M–2]<sup>+</sup> (8), 339[M–16]<sup>+</sup> (39), 296(54), 295(58), 188(100); IR bands (KBr): 3012, 2927, 2853, 1609, 1518, 1486, 1457, 1376, 1352, 1252, 1235, 1201, 1117, 1037, 928, 871 cm<sup>–1</sup>; <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>): δ 6.92 (1H, *s*, H-1), 6.76 (1H, *s*, H-4), 3.75 (1H, *d*, *J* 5.38 Hz, H-5), 4.15 (1H, *dd*, *J* 12.67 and 5.38 Hz, H-6<sub>a</sub>), 4.24 (1H, *d*, *J* 12.67 Hz, H-6<sub>b</sub>), 6.61 (1H, *s*, H-7), 6.50 (1H, *s*, H-10), 3.15 (1H, *dd*, *J* 19.07 and 3.20 Hz, H-11<sub>a</sub>), 3.65 (1H, *dd*, *J* 19.07 and 4.22 Hz, H-11<sub>b</sub>), 4.68 (1H, *dd*, *J* 4.22 and 3.20 Hz, H-12), 5.93 (1H, *d*, *J* 1.41 Hz, H-17), 5.92 (1H, *d*, *J* 1.41 Hz, H-17), 3.86 (3H, *s*, H-18), 3.80 (3H, *s*, H-19), 3.52 (3H, *s*, NCH<sub>3</sub>); <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>): 108.86 (C-1), 147.11 (C-2), 147.84 (C-3), 106.39 (C-4), 44.19 (C-5), 78.33 (C-6), 110.81 (C-7), 147.67 (C-8), 148.65 (C-9), 113.55 (C-10), 34.89 (C-11), 78.47 (C-12), 127.22 (C-13), 131.22 (C-14), 132.58 (C-15), 122.55 (C-16), 101.16 (C-17), 56.00 (C-18), 55.95 (C-19), 59.77 (NCH<sub>3</sub>).

**Compound 2.**  $[\alpha]_D^{27}$  –88.98° (CH<sub>3</sub>Cl, *c* 0.38); HR-FABMS *m/z*: 356.1489[M + H]<sup>+</sup> (calc. for C<sub>20</sub>H<sub>21</sub>NO<sub>5</sub> + H: 356.1498); EI-MS *m/z*: 355[M]<sup>+</sup> (2), 353[M–2]<sup>+</sup> (7), 339[M–16]<sup>+</sup> (19), 296(47), 295(100), 188(36); IR bands (KBr): 3010, 2923, 2853, 1609, 1517, 1485, 1486, 1457, 1374, 1353, 1253, 1233, 1210, 1118, 1036, 933, 870 cm<sup>–1</sup>; <sup>1</sup>H-NMR (500 MHz, CDCl<sub>3</sub>): δ 6.81 (1H, *s*, H-1), 6.75 (1H, *s*, H-4), 3.66 (1H, *d*, *J* 5.38 Hz, H-5), 3.98 (1H, *dd*, *J* 12.67 and 5.38 Hz, H-6<sub>a</sub>), 4.22 (1H, *d*, *J* 12.67 Hz, H-6<sub>b</sub>), 6.56 (1H, *s*, H-7), 6.58 (1H, *s*, H-10), 2.91 (1H, *dd*, *J* 19.07 and 3.20 Hz, H-11<sub>a</sub>), 4.73 (1H, *dd*, *J* 19.07 and 4.22 Hz, H-11<sub>b</sub>), 4.34 (1H, *dd*, *J* 4.22 and 3.20 Hz, H-12), 5.98 (1H, *d*, *J* 1.41 Hz, H-17), 5.91 (1H, *d*, *J* 1.41 Hz, H-17), 3.85 (3H, *s*, H-18), 3.78 (3H, *s*, H-19), 3.12 (3H, *s*, NCH<sub>3</sub>); <sup>13</sup>C-NMR (125 MHz, CDCl<sub>3</sub>): 108.14(C-1), 146.83(C-2), 147.34(C-3), 106.54(C-4), 45.00(C-5), 75.94(C-6), 110.14(C-

7), 146.19(C-8), 147.13(C-9), 113.83(C-10), 31.43(C-11), 78.48(C-12), 126.40(C-13), 132.21(C-14), 133.77(C-15), 125.14(C-16), 101.37(C-17), 56.03(C-18), 55.98(C-19), 62.38(NCH<sub>3</sub>).

### 3. Results and discussion

The two bases were isolated from the ethanol extract on silica column chromatography and preparative HPLC.



Compound **1** showed the quasi-molecular ion peak at  $m/z$  356[M + H]<sup>+</sup> in FABMS. In combination with <sup>13</sup>C and DEPT spectra, its molecular formula was deduced by HR-FABMS showing the molecular ion peak at  $m/z$  356.1547(calc. for C<sub>20</sub>H<sub>21</sub>NO<sub>5</sub> + H: 356.1498). <sup>13</sup>C and <sup>1</sup>H-NMR spectra showed signals characteristic of two 1,2,4,5 tetra-substituted aromatic rings, two aromatic methoxy groups and one double-oxygenated methene. In addition, a deshielded NCH<sub>3</sub> group at  $\delta$  3.52 was present. EI-MS showed a fragment pattern similar to amurensinine [4,5], which contains 16 m.u. less in its molecular weight. Compound **1** was therefore assigned the structure of amurensinine N-oxide. In fact, its EI-MS showed peaks at  $m/z$  339[M-16]<sup>+</sup>, 353[M-2]<sup>+</sup>, and 355[M]<sup>+</sup> similarly to (-)-argemonine N-oxide [6], as

expected for N-oxides [7]. In addition, compared with refractamine [8], the signals for  $\text{NCH}_3$ , H-6 and H-12 are deshielded by 0.7–1.3 p.p.m. and, compared with isothalisopavine [9], the signals for carbons  $\text{NCH}_3$ , C-6 and C-12 are deshielded by 14–20 p.p.m.

NOESY, showing that the proton signal of  $\text{NCH}_3$  was correlated with H-11<sub>b</sub> and H-12, suggested the relative configuration of  $\text{NCH}_3$ . The large negative optical rotation, typical of all isopavines so far isolated, suggested the assigned absolute configuration [9]. Though amurensinine has been isolated from other plants of the same genus (*M. horridula*, *M. napaulensis* and *M. sinuata*) [2,3], compound **1** is a new natural product, named amurensinine N-oxide A.

Compound **2** showed IR, EI-MS,  $^1\text{H}$  and  $^{13}\text{C}$ -NMR spectra similar to compound **1**. Compared to **1**, the signals for protons H-12, H-11<sub>a</sub>,  $\text{NCH}_3$  and H-6<sub>a</sub> were upfielded by 0.17–0.40 p.p.m. and H-11<sub>b</sub> was deshielded to 4.73 p.p.m. Meanwhile, the signals for carbons C-6 and C-11 were upfielded by 2.28 and 3.46 p.p.m., but  $\text{NCH}_3$  and C-16 were deshielded by 2.61 and 2.59 p.p.m., respectively. All the above evidence suggests compound **2** to be the epimer N-oxide of **1**. This was confirmed by NOESY which showed a correlation between  $\text{NCH}_3$  and H-12. Compound **2** was named amurensinine N-oxide B.

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