

A New Tricyclo[6.3.1.0^{2,5}]dodecane Sesquiterpene from Cultures of the Basidiomycete *Campanella junghuhnii*

by Rong Liu^a), Zhong-Yu Zhou^a), Di Xu^a)^b), Fei Wang^a), and Ji-Kai Liu^{*a})

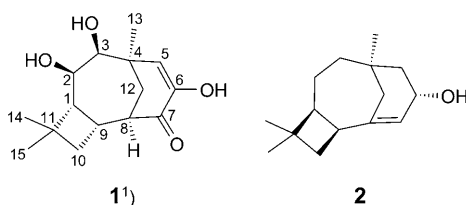
^a) State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204, P. R. China

(phone: +86-871-521-6327; fax: +86-871-515-0227; e-mail: jkliu@mail.kib.ac.cn)

^b) South China Agricultural University, Guangzhou 510642, P. R. China

A new sesquiterpene with a tricyclo[6.3.1.0^{2,5}]dodecane skeleton, 2,3,6-trihydroxycaryol-5-en-7-one (**1**), was isolated from the culture of the basidiomycete *Campanella junghuhnii*. The structure of **1** was elucidated on the basis of extensive spectroscopic analysis including IR, UV, MS, 1D- and 2D-NMR experiments.

Introduction. – *Campanella junghuhnii*, belonging to the family Marasmiaceae, is a small, thin, white mushroom, usually growing on bamboo stems [1]. So far, secondary metabolites produced by fungi of the genus *Campanella* have not been reported. As one part of our research for naturally occurring bioactive metabolites from higher fungi in China [2–6], we have carried out a chemical investigation on the cultures of *C. junghuhnii* which led to the isolation of a new sesquiterpene (**1**). Comparison of the NMR data of **1** with those of the cytotoxic sesquiterpene caryol-7-en-6-ol (**2**) [7], which was isolated from a New Zealand sponge of the genus *Eurypon*, implied that they share the same tricyclo[6.3.1.0^{2,5}]dodecane skeleton. This is the first report of the isolation of a sesquiterpene with this skeleton from a higher fungus. The structural elucidation of **1** was mainly performed with 1D- and 2D-NMR experiments.¹⁾



Results and Discussion. – The culture of *C. junghuhnii* (201) was filtered, and the filtrate was extracted four times with AcOEt. The organic layer was concentrated *in vacuo* to give a crude extract (40 g), which was subjected to repeated column chromatography to afford pure **1**.

¹⁾ Arbitrary numbering. For the systematic name, see *Exper. Part*.

Compound **1** was obtained as a colorless oil. The HR-ESI-MS exhibited a *quasi*-molecular ion peak at m/z 289.1420 ($[M + Na]^+$; calc. 289.1415), indicating the molecular formula $C_{15}H_{22}O_4$. The IR spectrum showed absorption bands for OH (3422 cm^{-1}), C=O (1734 cm^{-1}), and C=C (1667 cm^{-1}) functional groups. Based on the UV absorption maximum at 277 nm, and the C=O signal at $\delta(C)$ 198.2 (*s*, C(7)) and C=C signals at $\delta(C)$ 123.5 (*d*, C(5)¹) and $\delta(C)$ 146.3 (*s*, C(6)) in the ^{13}C -NMR spectrum (Table), it was concluded that the C=O group was present as a α,β -unsaturated ketone group. Broad-band decoupled ^{13}C -NMR and DEPT spectra disclosed the presence of three Me, two CH_2 , and six CH groups (thereof two O–CH groups at $\delta(C)$ 70.1 (*d*, C(2)) and $\delta(C)$ 81.6 (*d*, C(3))), and four quaternary C-atoms. Comparing the ^{13}C -NMR data of **1** with those of **2**, the five degrees of unsaturation of **1** required by the molecular formula could be accommodated by the presence of an enone group and of a tricyclic skeleton.

The ^1H , ^1H -COSY spectrum of **1** revealed two spin systems: the C(3)–C(2)–C(1)–C(9)–C(8)–C(12) and the C(9)–C(10) unit (see the formula for the arbitrary numbering system). C(2) and C(3) were both substituted by OH groups as deduced from the NMR signals (Table) at $\delta(\text{H})$ 4.18 (*dd*, $J = 11.6, 1.2$, H–C(2)), $\delta(C)$ 70.1 (*d*, C(2)), $\delta(\text{H})$ 3.68 (*d*, $J = 1.2$, H–C(3)), $\delta(C)$ 81.6 (*d*, C(3)) and the HMBC of H–C(2) with C(1), C(3), C(9), and C(11), and H–C(3) with C(1), C(2), C(4), C(5), and C(13) (Fig. 1). The HMBC spectrum showed also correlations from H–C(5), H–C(8), H–C(9), and H–C(12) to the ketone C=O group at $\delta(C)$ 198.2 (*s*, C(7)), which indicated that the ketone C=O group is located at C(7) and the C=C bond at C(5)/C(6). Furthermore, an oxygenated olefinic quaternary C-atom ($\delta(C)$ 146.3) assigned to C(6) was supported by the HMBCs from H–C(8) to C(6), and from H–C(12) and H–C(13) to C(5). The relative configuration of **1** was determined by a ROESY experiment. The ROESY correlations (Fig. 2) of H–C(3) and H–C(5) with α -Me(13), H–C(8) and Me(15) with H_α -C(10), H–C(2) with α -Me(15), H–C(1) and H–C(9) with H_β -C(12), and Me(14) with H_β -C(10) indicated that

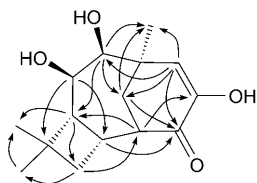


Fig. 1. Key HMBC data of **1**

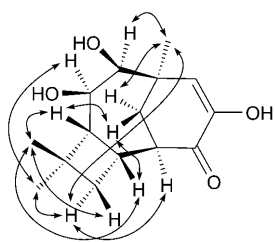


Fig. 2. Key ROESY correlations of **1**

H–C(1), H–C(2), H–C(3), H–C(5), H–C(8), H–C(9), Me(14), and Me(15) possessed β -, α -, α -, α -, α -, β -, β -, and α -orientations, respectively. On the basis of the above evidence, the structure of **1** was deduced as 2,3,6-trihydroxycaryol-5-en-7-one.

Table. NMR Spectral Data of **1** and **2**. Measured in CDCl₃; δ in ppm, J in Hz.

	1		2	
	δ (H) ^a	δ (C) ^b	δ (H) ^c	δ (C) ^d
H–C(1)	2.46 (<i>dd</i> , $J = 11.6, 11.6$)	47.6 (<i>d</i>)	1.82–1.89 (<i>m</i>)	49.9 (<i>d</i>)
H–C(2) or CH ₂ (2)	4.18 (<i>dd</i> , $J = 11.6, 1.2$)	70.1 (<i>d</i>)	1.58–1.65 (<i>m</i>), 1.42–1.50 (<i>m</i>)	22.2 (<i>t</i>)
H–C(3) or CH ₂ (3)	3.68 (<i>d</i> , $J = 1.2$)	81.6 (<i>d</i>)	1.20–1.29 (<i>m</i>), 1.12–1.19 (<i>m</i>)	37.9 (<i>t</i>)
C(4)		37.7 (<i>s</i>)		33.5 (<i>s</i>)
H–C(5) or CH ₂ (5)	5.84 (<i>s</i>)	123.5 (<i>d</i>)	1.96 (<i>dd</i> , $J = 13.0, 9.3$), 0.08–1.03 (<i>m</i>)	47.2 (<i>t</i>)
C(6) or H–C(6)		146.3 (<i>s</i>)	4.45–4.51 (<i>m</i>)	67.8 (<i>d</i>)
C(7) or H–C(7)		198.2 (<i>s</i>)	5.36–5.43 (<i>m</i>)	125.3 (<i>d</i>)
H–C(8) or C(8)	2.59–2.66 (<i>m</i>)	44.1 (<i>d</i>)		140.8 (<i>s</i>)
H–C(9)	2.80–2.89 (<i>m</i>)	32.3 (<i>d</i>)	3.21–3.30 (<i>m</i>)	38.3 (<i>d</i>)
CH ₂ (10)	1.72–1.78 (<i>m</i>), 1.57 (<i>dd</i> , $J = 12.3, 9.7$)	35.5 (<i>t</i>)	1.97 (<i>t</i> , $J = 10.7$), 1.67–1.74 (<i>m</i>)	36.0 (<i>t</i>)
C(11)		32.4 (<i>s</i>)		34.7 (<i>s</i>)
CH ₂ (12)	2.14 (<i>d</i> , $J = 5.3$), 1.82 (<i>dd</i> , $J = 6.5, 5.3$)	32.8 (<i>t</i>)	2.21–2.28 (<i>m</i>), 1.36 (<i>br. d</i> , $J = 12.5, 2.0$)	35.2 (<i>t</i>)
Me(13)	1.22 (<i>s</i>)	28.9 (<i>q</i>)	1.05 (<i>s</i>)	28.2 (<i>q</i>)
Me(14)	1.18 (<i>s</i>)	31.2 (<i>q</i>)	0.96 (<i>s</i>)	24.2 (<i>q</i>)
Me(15)	0.93 (<i>s</i>)	25.3 (<i>q</i>)	1.22 (<i>s</i>)	30.3 (<i>q</i>)

^a) Recorded at 400 MHz. ^b) Recorded at 125 MHz. ^c) Recorded at 300 MHz. ^d) Recorded at 75 MHz; multiplicities inferred from DEPT and HMQC experiments.

Experimental Part

General. Column chromatography (CC): silica gel (SiO₂; 200–300 mesh, Qingdao Marine Chemical Inc., P. R. China) and Sephadex LH-20 (Amersham Biosciences, Uppsala, Sweden); TLC monitoring, visualization by heating the SiO₂ plates sprayed with 10% H₂SO₄ in EtOH. Optical rotations: Horiba SEPA-300 digital polarimeter. UV Spectra: Shimadzu UV-210 spectrometer; λ_{\max} (log ϵ) in nm. IR Spectra: Bruker Tensor-27 spectrometer; with KBr pellets; in cm^{–1}. ¹H- and ¹³C-NMR Spectra: Bruker AM-400 and DRX-500 spectrometers; δ in ppm, J in Hz. MS: VG Autospec-3000 and API QSTAR-Pulsar-1 spectrometer; in m/z (rel. int.).

Mushroom Material and Culture. The fungus *C. junghuhnii* was isolated from the tissue culture of its fruiting bodies collected at Gaoligong Mountains, Yunnan Province, P. R. China, in July 2006, and identified by Prof. Mu Zang, Kunming Institute of Botany, Chinese Academy of Sciences (CAS). The voucher specimen was deposited with the Herbarium of the Kunming Institute of Botany, CAS. Culture medium: potato (peeled) 200 g, glucose 20 g, KH₂PO₄ 3 g, MgSO₄ 1.5 g, citric acid 0.1 g, and thiamine hydrochloride 10 mg in 1 l of deionized H₂O. The pH was adjusted to 6.5 before autoclaving, and the fermentation was carried out on a shaker at 25° and 150 rpm for 20 d.

Extraction and Isolation. The whole culture of *C. junghuhnii* (201) was filtered, and the filtrate was extracted four times with AcOEt. The org. layer was concentrated *in vacuo* to give a crude extract (40 g),

and the residue was subjected to CC (SiO₂; CHCl₃/MeOH gradient system) to give ten fractions. The fraction (665 mg) eluted with CHCl₃/MeOH (95:5, v/v) was subjected to repeated CC (*Sephadex LH-20*; CHCl₃/MeOH 1:1) to produce three subfractions *Fr. 1* (259 mg), *Fr. 2* (165 mg), and *Fr. 3* (15 mg). *Fr. 2* was further purified by CC (SiO₂; petroleum ether/AcOEt 4:1) and (*Sephadex LH-20*, CHCl₃/MeOH 1:1) to afford pure compound **1** (10 mg).

2,3,6-Trihydroxycaryol-5-en-7-one (= (*1R,2R,5R,6R,7S,8R*)-*6,7,10-Trihydroxy-4,4,8-trimethyltricyclo[6.3.1.0^{2,5}]dodec-9-en-11-one*; **1**). Colorless oil. *R_f* (PE/acetone 2:1): 0.60. $[\alpha]_{\text{D}}^{25} = -52.1$ (*c* = 0.37, CHCl₃). UV (CHCl₃): 277 (3.65). IR (KBr): 3422, 2953, 2933, 1734, 1667, 1459, 1406, 1367, 1222, 1175, 1070, 1059, 1024, 984, 936. ¹H- and ¹³C-NMR (CDCl₃): *Table*. EI-MS: 266 (*M*⁺), 248 (*[M - H₂O]*⁺), 230 (*[M - 2 H₂O]*⁺). HR-ESI-MS: 289.1420 (*[M + Na]*⁺, C₁₅H₂₂NaO₄⁺; calc. 289.1415).

This project was supported by the *Chinese Academy of Sciences* (KSCX1-YW-R-24; KSCX2-YW-G-025) and the *National Basic Research Program of China* (2009CB522300).

REFERENCES

- [1] X. L. Mao, 'The Macrofungi in China', Henan Science and Technology Press, Henan, 2000, p. 168.
- [2] J. K. Liu, *Chem. Rev.* **2006**, *106*, 2209.
- [3] J. K. Liu, *Chem. Rev.* **2005**, *105*, 2723.
- [4] D.-Z. Liu, F. Wang, T.-G. Liao, J.-G. Tang, W. Steglich, H.-J. Zhu, J.-K. Liu, *Org. Lett.* **2006**, *8*, 5749.
- [5] X.-D. Qin, Z.-J. Dong, J.-K. Liu, L.-M. Yang, R.-R. Wang, Y.-T. Zheng, Y. Lu, Y.-S. Wu, Q.-T. Zheng, *Helv. Chim. Acta* **2006**, *89*, 127.
- [6] D.-Q. Luo, H.-J. Shao, H.-J. Zhu, J.-K. Liu, *Z. Naturforsch., C* **2005**, *60*, 50.
- [7] C. J. Barrow, J. W. Blunt, M. H. G. Munro, *Aust. J. Chem.* **1988**, *41*, 1755.

Received July 29, 2008