

Two new *ent*-kaurane diterpenoids from *Isodon excisoides*

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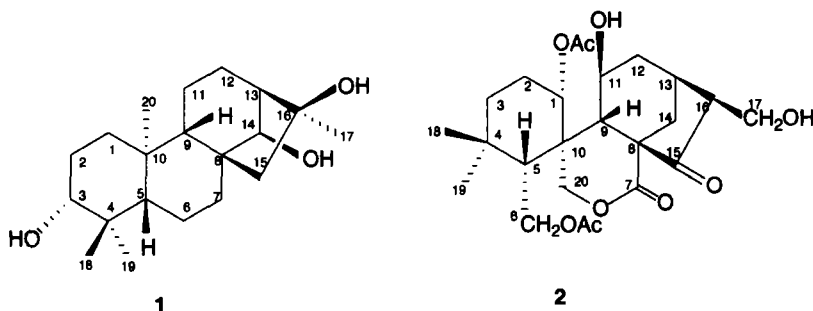
Abstract

Two new *ent*-kaurane diterpenoids taihangexcisoidesin A and B (1 and 2), were isolated from the EtOAc extract of the leaves of *Isodon excisoides*. Their structures were determined on the basis of spectroscopic methods.

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Isodon species are rich in *ent*-kaurane diterpenoids, some of which have antitumor, antibacterial, and antiinflammatory activities [1,2]. To search more bioactive compounds in genus *Isodon*, two new *ent*-kaurane diterpenoids taihangexcisoidesin A and B (1 and 2), were isolated from the EtOAc extract of the leaves of *Isodon excisoides* (Sun ex C.H. Hu) C.Y. Wu et H.W. Li, which was collected in Taihang Mountains, Henan province. This paper presents the structural elucidation of the new compounds.



Compound 1, obtained as colourless needles from MeOH, has a molecular formula $C_{20}H_{34}O_3$ based on its HR-ESI-MS (m/z 345.2391 $[M+Na]^+$, calcd. 345.2406) and the 1H and ^{13}C NMR data, suggesting four degrees of unsaturation. In its ^{13}C NMR (DEPT) spectrum showed the signals of four methyls, seven methylenes, five methines including two oxymethines [δ_C 78.1 (d), δ_H 3.44 (m, 1H) and δ_C 78.9 (d), δ_H 4.46 (d, 1H, $J = 8.4$ Hz)], and four quaternary carbons

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including one oxygenated [δ_{C} 79.2 (s)]. Considering the compounds isolated from the *Isodon* genus, compound **1** had a 20-non-oxygenated-*ent*-kaurane diterpene skeleton. The chemical shifts at δ_{H} 6.32 (d, 1H, $J = 6.4$ Hz), δ_{H} 5.98 (s, 1H) and 5.70 (br s, 1H) in the ^1H NMR spectrum and the absorption at 3351 and 3318 cm^{-1} in the IR spectrum suggested the existence of three hydroxyl groups. In the HMBC spectrum, the correlations were clearly observed among H-14 with C-8, C-9, C-15 and C-16, H-3 with C-1 and C-5, H₂-12, H₂-15 and H₃-17 with C-16 (Fig. 1). Meanwhile, according to the cross-peaks in the HMBC and ^1H - ^1H COSY spectra, three hydroxyl groups were obviously located at C-3, C-14 and C-16, respectively.

The relative configuration of the substituents were revealed by NOESY experiments (Fig. 2). In the NOESY spectrum, there were correlations of H-3 with H-1 β , H-5 β and Me-18, H-14 with H-6 α , H-12 α , H-13 α and Me-20. Thus, OH-3 and OH-14 have the α and β orientation, respectively. The β -orientation of OH-16 was suggested by the clear cross-peaks of H₃-17 with H-13 α in NOESY spectrum as shown in Fig. 2. Therefore, **1** was elucidated as 3 α , 14 β , 16 β -trihydroxy-*ent*-kaurane, and named taihangexcisoidesin A.

Compound **2** was obtained from MeOH as colourless needles, showed the molecular ion peak at m/z 489.2083 $[\text{M}+\text{Na}]^+$ in its HR-ESI-MS, consistent with the molecular formula $\text{C}_{24}\text{H}_{34}\text{O}_9$ which was confirmed by its ^{13}C NMR spectrum presenting signals for 24 carbons of molecular formula including four carbons of two acetoxyl groups. On the basis of the characteristic lactone carbonyl signal at δ_{C} 170.3 (s) due to C-7 and noticeable oxygenated methylene signals [δ_{C} 67.6 (t), C-20; δ_{H} 5.08 and 4.79 (ABd, each 1H, $J = 12.4$ Hz), H-20a/b], compound **2** was inferred to be a 6,7-seco-7,20-olide *ent*-kauranoid. Comparison of the spectroscopic data of **2** with those of robdosin B [3] indicated that **2** was nearly identical with robdosin B except for C-16 and C-17. The exomethylene group at C-16 of robdosin B was replaced by a methine (δ_{C} 57.4, C-16; δ_{H} 3.11, m, 1H, H-16 α) and a hydroxymethyl group (δ_{C} 59.1, C-17; δ_{H} 4.39, m, 2H, H₂-17) in **2**, which was approved by the HMBC spectral evidence (Fig. 1). The 16 β -CH₂OH was determined by the HMBC correlations of H₂-17 with C-13 and C-15, and the NOEs of H-16 α with H-13 α , H₂-17 with H-12 β and H-9 β (Fig. 2), which was also confirmed by the obvious upshift of C-12 (δ_{C} 33.1 in **2**, δ_{C} 41.5 in robdosin B) caused by the steric compress effect between H₂-17 and H-12 β [4]. The β -orientation of H-1 was proved by the NOE of H-1 with H-5 β . Similarly, H-11 α was confirmed by the NOE among H-11 α with H-14 α . Thus, compound **2** was deduced as 16(*S*)-hydroxymethyl-11 β -hydroxy-1 α ,6-diacetoxy-6,7-seco-7,20-olide-*ent*-kaur-15-one, named taihangexcisoidesin B.

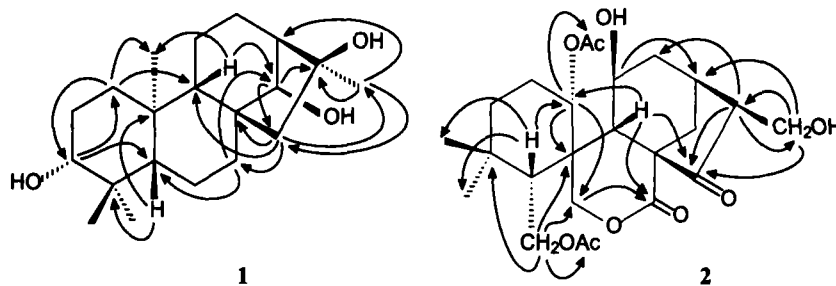


Fig. 1. Key HMBC correlations of compounds **1** and **2**.

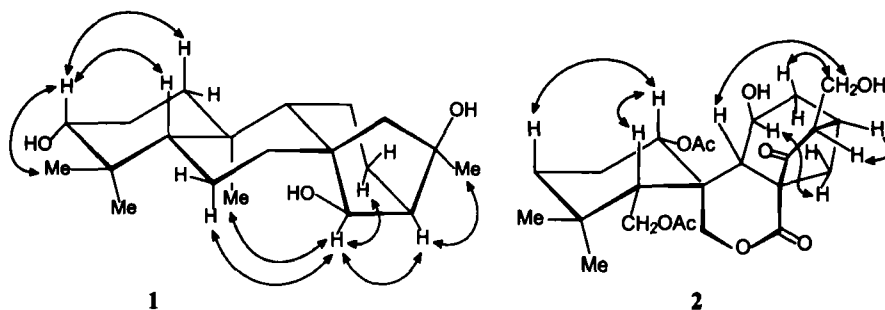


Fig. 2. Key NOESY correlations of compounds **1** and **2**.

Table 1
 ^{13}C (100 MHz) NMR spectral data of 1 and 2 in $\text{C}_5\text{D}_5\text{N}$ (δ in ppm).

Position	δ_{C}		Position	δ_{C}	
	1	2		1	2
1	39.2 t	77.1 d	12	27.8 t	33.1 t
2	28.4 t	24.5 t	13	54.5 d	31.5 d
3	78.1 d	40.1 t	14	78.9 d	31.5 t
4	39.4 s	34.3 s	15	57.3 t	212.4 s
5	55.5 d	49.3 d	16	79.2 s	57.4 d
6	20.2 t	61.9 t	17	24.4 q	59.1 t
7	34.1 t	170.3 s	18	28.0 q	34.0 q
8	51.3 s	59.5 s	19	18.0 q	24.1 q
9	59.6 d	44.1 d	20	16.3 q	67.6 t
10	39.4 s	44.7 s	OCOCH_3		170.4 s, 170.5 s
11	18.0 t	65.9 d	OCOCH_3		21.5 q, 21.2 q

Compound 1: $\text{C}_{20}\text{H}_{34}\text{O}_3$, colourless needles, m.p. 237–239 °C, IR ν^{KBr} (cm^{-1}): 3351, 3318, 2946, 2864, 1453, 1066, 1040, 1028, 976, 934. ^1H -NMR ($\text{C}_5\text{D}_5\text{N}$, 400 MHz, δ (ppm)): 4.46 (d, 1H, $J = 8.4$ Hz, H-14 α), 3.44 (m, 1H, H-3 β), 2.78 (d, 1H, $J = 12.8$ Hz, H-7a), 2.32 (br s, 1H, H-13 α), 2.22 and 1.72 (d, each 1H, $J = 13.6$ Hz, H-15a/b), 1.88 (m, 2H, H₂-2), 1.74 (m, 1H, H-1a), 1.67 (m, 2H, H₂-12), 1.60 (m, 2H, H₂-11), 1.52 (s, 3H, Me-17), 1.45 (m, 2H, H₂-6), 1.14 (m, 1H, H-9 β), 1.13 (m, 1H, H-7b), 1.20, 0.98 and 0.97 (s, each 3H, 3 \times Me), 0.92 (m, 1H, H-1b), 0.85 (dd, 1H, $J = 11.6$, 2.0 Hz, H-5 β). HR-ESI-MS m/z : 345.2391 $[\text{M}+\text{Na}]^+$ (calcd. 345.2406). ^{13}C NMR data see Table 1.

Compound 2: $\text{C}_{24}\text{H}_{34}\text{O}_9$, colourless needles, m.p. 180–182 °C, $[\alpha]_{\text{D}}^{20} +53.8$ (c 0.02, MeOH), IR ν^{KBr} (cm^{-1}): 3572, 3471, 3322, 2993, 2944, 2837, 2822, 1726, 1708, 1640, 1409, 1375, 1301, 1262, 1233, 1126, 1054. ^1H -NMR ($\text{C}_5\text{D}_5\text{N}$, 400 MHz, δ (ppm)): 5.57 (dd, 1H, $J = 5.6$, 9.6 Hz, H-1 β), 5.08 and 4.79 (ABd, each 1H, $J = 12.4$ Hz, H-20a/b), 4.57 and 4.47 (dd, each 1H, $J = 12.8$, 4.0 Hz, H-6a/b), 4.41 (m, 1H, H-11 α), 4.39 (m, 2H, H₂-17), 3.29 (t, 1H, $J = 7.6$, 3.6 Hz, H-5 β), 3.11 (m, 1H, H-16 α), 3.06 (d, 1H, $J = 9.6$ Hz, H-9 β), 2.85 (br s, 1H, H-13 α), 2.62 (dd, 1H, $J = 12.8$, 4.0 Hz, H-14a), 2.32 (d, 1H, $J = 12.8$ Hz, H-14b), 2.24 (m, 2H, H₂-12), 2.17 and 1.98 (s, each 3H, 2 \times OAc), 1.92 (m, 2H, H₂-2), 1.33 (m, 2H, H₂-3), 0.91 and 0.87 (s, each 3H, 2 \times Me). HR-ESI-MS m/z : 489.2083 $[\text{M}+\text{Na}]^+$ (calcd. 489.2101). ^{13}C NMR data see Table 1.

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