

International Journal of Systematic and Evolutionary Microbiology
Stackebrandtia endophytica sp. nov., a novel actinobacterium isolated from
Tripterygium wilfordii
 --Manuscript Draft--

Manuscript Number:	IJS-D-14-00506R1
Full Title:	Stackebrandtia endophytica sp. nov., a novel actinobacterium isolated from Tripterygium wilfordii
Short Title:	Stackebrandtia endophytica sp. nov.
Article Type:	Standard
Section/Category:	New taxa - Actinobacteria
Corresponding Author:	Li-Xing Zhao Yunnan University CHINA
First Author:	Zi-Jun Xiong
Order of Authors:	Zi-Jun Xiong Cui-Ping Miao You-Kun Zheng Kai Liu Wen-Jun Li Wei-Hong Liu Li-Hua Xu Li-Xing Zhao
Manuscript Region of Origin:	CHINA
Abstract:	A novel endophytic actinobacterium, designated strain YIM 64602T, was isolated from healthy stems of Tripterygium wilfordii. It grew at 15-40 °C, pH 6.0-9.0 and in the presence of 0-3 % (w/v) NaCl. Phylogenetic analysis based on 16S rRNA gene sequence showed that strain YIM 64602T belongs to the genus Stackebrandtia. Whole-cell hydrolyzates of strain YIM 64602T contained amino acid of meso-diaminopimelic acid with sugars mannose, rhamnose and glucose, a trace of ribose. The major polar lipids were diphosphatidylglycerol, phosphatidylmethylethanolamine and phosphatidylethanolamine. MK-10(H6), MK-10(H4) and MK-11(H4) were the predominant components in the quinone system. The fatty-acid pattern was mainly composed of the saturated branched-chain acids iso-C16:0, anteiso-C17:0, iso-C15:0 and iso-C17:0. The DNA G+C content was 72.4 mol %. The 16S rRNA gene sequence analysis showed the highest pairwise sequence identity (96.0-98.5 %) with the members of the genus Stackebrandtia. Strain YIM 64602T displayed a DNA-DNA relatedness of 43.9 ± 0.4 % with the type strain Stackebrandtia albiflava YIM 45751T. Based on polyphasic evidence from this study, strain YIM 64602T (= BCRC 16954T = DSM 45928T) was considered to represent a novel species of the genus Stackebrandtia, for which the name Stackebrandtia endophytica is proposed.

1 ***Stackebrandtia endophytica* sp. nov., a novel actinobacterium isolated**

2 **from *Tripterygium wilfordii***

3 **Zi-Jun Xiong,^{1,2†} Cui-Ping Miao,^{1†} You-Kun Zheng,¹ Kai Liu,¹ Wen-Jun Li,¹**

4 **Wei-Hong Liu,³ Li-Hua Xu¹ and Li-Xing Zhao^{1*}**

5 **¹Key Laboratory of Microbial Diversity in Southwest China, Ministry of**
6 **Education and Laboratory for Conservation and Utilization of Bio-Resources,**
7 **Yunnan Institute of Microbiology, Yunnan University, Kunming 650091, P. R.**
8 **China.**

9 **²State Key Laboratory of Phytochemistry and Plant Resources in West China,**
10 **Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650201, P. R.**
11 **China.**

12 **³Department of Agriculture and Biological Sciences, Dali University, Dali 671003, P.**
13 **R. China.**

14 **Author for correspondence: Li-Xing Zhao**

15 **E-mail: zlx70@163.com**

16
17 **[†]These authors contributed equally to this work.**

18
19 **Running title: *Stackebrandtia endophytica* sp. nov.**

20
21 **Subject Category: New Taxa-Actinobacteria**

22
23 **The 16S rRNA gene sequence of strain YIM 64602^T has been deposited in GenBank**
24 **under the accession number KJ781245.**

25

26 A novel endophytic actinobacterium, designated strain YIM 64602^T, was isolated
27 from healthy stems of *Tripterygium wilfordii*. It grew at 15-40 °C, pH 6.0-9.0 and in
28 the presence of 0-3 % (w/v) NaCl. Phylogenetic analysis based on 16S rRNA gene
29 sequence showed that strain YIM 64602^T belongs to the genus *Stackebrandtia*.
30 Whole-cell hydrolyzates of strain YIM 64602^T contained amino acid of
31 *meso*-diaminopimelic acid with sugars mannose, rhamnose and glucose, a trace of
32 ribose. The major polar lipids were diphosphatidylglycerol,
33 phosphatidylmethylethanolamine and phosphatidylethanolamine. MK-10(H₆),
34 MK-10(H₄) and MK-11(H₄) were the predominant components in the quinone
35 system. The fatty-acid pattern was mainly composed of the saturated
36 branched-chain acids iso-C_{16:0}, anteiso-C_{17:0}, iso-C_{15:0} and iso-C_{17:0}. The DNA G+C
37 content was 72.4 mol %. The 16S rRNA gene sequence analysis showed the highest
38 pairwise sequence identity (96.0-98.5 %) with the members of the genus
39 *Stackebrandtia*. Strain YIM 64602^T displayed a DNA-DNA relatedness of 43.9 ± 0.4 %
40 with the type strain *Stackebrandtia albiflava* YIM 45751^T. Based on polyphasic
41 evidence from this study, strain YIM 64602^T (= BCRC 16954^T = DSM 45928^T) was
42 considered to represent a novel species of the genus *Stackebrandtia*, for which the
43 name *Stackebrandtia endophytica* is proposed.

44

45 The family *Glycomycetaceae* contains three recognized genera: *Glycomyces* (Labeda *et*
46 *al.*, 1985; Labeda & Kroppenstedt, 2004), *Stackebrandtia* (Labeda & Kroppenstedt,
47 2005) and *Haloglycomyces* (Guan *et al.* 2009). Currently, the family comprises 14
48 members, and the genus *Stackebrandtia* contains 2 members, *Stackebrandtia*
49 *nassauensis* (Labeda & Kroppenstedt, 2005) and *Stackebrandtia albiflava* (Wang *et al.*,
50 2009) which were isolated from different soil samples. The two species in
51 *Stackebrandtia* are Gram-positive, strictly aerobic, filamentous actinomycetes. The
52 cell-wall peptidoglycan contains *meso*-diaminopimelic acid. The major polar lipids
53 consist of diphosphatidylglycerol (DPG), phosphatidylethanolamine (PE),
54 phosphatidylmethylethanolamine (PME) and phosphatedylglycerol (PG). The
55 predominant menaquinones are MK-10(H₄), MK-10(H₆), MK-11(H₄) and MK-11(H₆).

56 The major fatty acids are saturated, iso- and anteiso-branched fatty acids. The G +C
57 contents of the genomic DNA are 69-73 mol %. In the present study, we report another
58 novel species of this genus, strain YIM 64602^T, which was isolated from healthy stems
59 of *Tripterygium wilfordii*, a traditional Chinese medicinal plant.

60

61 The stems of *Tripterygium wilfordii* were collected in Yunnan Province, south-west
62 China. The samples were firstly washed in running water to remove soil particles and
63 sterilized by 5% sodium hypochlorite and 70% ethanol according to the established
64 procedure (Li *et al.*, 2008), then sliced into pieces, followed by plating on the
65 cellulose-asparagine agar [2.5 g cellulose, 2.0 g sodium pyruvate, 1.0 g asparagine, 0.5
66 g CaCl₂, 0.25 g KNO₃, 0.2 g MgSO₄·7H₂O, 0.2 g K₂HPO₄, 10 mg FeSO₄·7H₂O and 15 g
67 agar; pH 7.2,] containing nalidixic acid (25 mg L⁻¹), nystatin (75 mg L⁻¹) and potassium
68 dichromate (50 mg L⁻¹) to inhibit the growth of bacteria and fungi. The plates were
69 incubated at 28 °C for 4-6 weeks until the outgrowth of endophytic actinomycetes were
70 discerned. Strain YIM 64602^T was purified and maintained on ISP (International
71 Streptomyces Project) 2 (Shirling & Gottlieb, 1966) agar slants at 4 °C and as 20 % (v/v)
72 glycerol suspensions at -80 °C.

73

74 Extraction of genomic DNA, PCR amplification and sequencing of the 16S rRNA gene
75 were carried out as described by Li *et al.* (2007) and Cui *et al.* (2001). The values for
76 sequence similarity among the closest strains were determined using the EzTaxon-e
77 server Database (<http://eztaxon-e.ezbiocloud.net/>; Kim *et al.*, 2012). Multiple alignments
78 with sequences of the most closely related actinobacteria were carried out using the
79 CLUSTAL_X 1.8 program (Thompson *et al.*, 1997). Phylogenetic trees were
80 constructed by the neighbour-joining (Saitou & Nei, 1987), maximum-parsimony (Fitch,
81 1971) and maximum-likelihood (Felsenstein, 1981) tree-making algorithms by using the
82 software packages MEGA version 5.05 (Tamura *et al.*, 2011). The stability of
83 relationships was assessed by performing bootstrap analyses with 1000 resamplings
84 (Felsenstein, 1985). The DNA-DNA hybridization was determined according to the
85 fluorometric micro-well method (Ezaki *et al.*, 1989; He *et al.*, 2005).

86

87 The almost complete (1521 bp) 16S rRNA gene sequence of strain YIM 64602^T was
88 determined and deposited in GenBank as KJ781245. The 16S rRNA gene sequence
89 showed the highest similarity with the members of the genus *Stackebrandtia*, family
90 *Glycomycetaceae*, especially with the type strains of *S. albiflava* YIM45751^T (98.5 %) and
91 *S. nassauensis* DSM 44728^T (96.0 %). Genomic relatedness of strain YIM 64602^T
92 to *S. albiflava* YIM 45751^T was $43.9 \pm 0.4\%$. A comparison of the sequences with the
93 type species of the related genera showed that the organism fell within the evolutionary
94 radiation occupied by the genus *Stackebrandtia* (Fig. 1). In the tree based on the
95 neighbour-joining algorithm, strain YIM 64602^T formed a coherent cluster with *S.*
96 *albiflava* YIM 45751^T and *S. nassauensis* DSM 44728^T; the branching order was
97 supported further by the bootstrap value 100% and 98%. The similar tree topology was
98 also obtained with the phylogenetic trees generated using maximum-parsimony and
99 maximum-likelihood algorithms (Figs. S1-2). These data supported the finding that
100 strain YIM 64602^T represents a different genomic species.

101

102 Biomass for chemical studies of strain YIM 64602^T was grown on ISP 2 agar plates for
103 7 days at 28 °C. The isomer of diaminopimelic acid and whole-cell sugars were analysed
104 according to the procedures developed by Hasegawa *et al.* (1983) and Tang *et al.* (2009).
105 Menaquinones were isolated according to Collins *et al.* (1977) and separated by HPLC
106 (Tamaoka *et al.*, 1983). Polar lipids were extracted and analysed by two-dimensional
107 TLC according to Embley & Wait (1994). Biomass for fatty acid analysis was obtained
108 by cultivation on tryptic soya agar (TSA) at 28 °C for 3 days. Cellular fatty acid analysis
109 was performed by using the Microbial Identification System (Sherlock Version 6.1;
110 MIDI database: TSBA6). The G + C DNA content of the strain YIM 64602^T was
111 determined by using the HPLC method (Mesbah *et al.*, 1989).

112

113 Strain YIM 64602^T shared consistent chemotaxonomic characteristics with *S. albiflava*
114 YIM 45751^T. The strain YIM 64602^T contained *meso*-diaminopimelic acid (*meso*-DAP)
115 as the diagnostic diamino acid in the peptidoglycan and sugars in whole-cell

116 hydrolysates contained mannose, rhamnose, glucose, and with a trace of ribose. Strain
117 YIM 64602^T is distinguished from the type strain of *S. albiflava* YIM 45751^T by the
118 absence of galactose and xylose (Wang *et al.*, 2009). In this study, *S. albiflava* YIM
119 45751^T, *S. nassauensis* DSM 44728^T and *Glycomyces harbinensis* DSM 46494^T were
120 reanalyzed as described by Tang *et al.* (2009). All of them were found to contain
121 mannose, galactose, rhamnose, glucose and ribose (Fig. S5). The predominant
122 menaquinones of YIM 64602^T were MK-10(H₆), MK-10(H₄) and MK-11(H₄). The polar
123 lipids consisted of DPG, PE and PME, and with some PG, phosphatidylinositol (PI);
124 phosphatidylinositol mannosides (PIM), unknown phospholipids (PL) and unidentified
125 polar lipid (UL) as minor components (Fig. S3). The major cellular fatty acid
126 compositions (>10 %) of strain YIM 64602^T showed the presence of iso-C_{16:0} (20.29 %),
127 anteiso-C_{17:0} (18.48 %), iso-C_{15:0} (11.37 %) and iso-C_{17:0} (10.87 %). Detailed cellular
128 fatty acid composition of strains YIM 64602^T and *S. albiflava* YIM 45751^T were
129 presented in Table S1. The DNA G+C content was 72.4 mol %. The chemotaxonomic
130 data for the new isolate matched genus *Stackebrandtia*, and also differentiated from
131 them by absence of galactose in the whole-cell hydrolysate, absence of MK-10(H₆) in the
132 predominant menaquinone (Table 1).

133

134 Aerial spore-mass colour, substrate mycelium pigmentation and coloration of the
135 diffusible pigments of strain YIM 64602^T were recorded on ISP 2, 3, 4 and 5 media and
136 Czapek's agar. Colours were determined by using colour chips from the ISCC-NBS
137 colour charts (standard samples, no. 2106) (Kelly, 1964). Morphological properties were
138 examined using a light microscopy (BH 2; Olympus) and scanning electron microscopy
139 (Quanta 200; FEI) after 14-21 days incubation on ISP 2 medium at 28 °C. Growth was
140 tested at 4, 10, 15, 20, 28, 30, 35, 40, 45 and 50 °C on ISP 2 medium by incubating the
141 cultures for 14 days. The ability of the strain to grow at different pH (pH 4, 5, 6, 7, 8, 9,
142 10 and 11, using the buffer system described by Xu *et al.*, 2005) and NaCl
143 concentrations (0, 1, 2, 3, 4, 5, 6, 7, 8, 9 and 10 %, w/v) was examined at 28 °C after 14
144 days. Anaerobic cultivation was performed on ISP 2 using the OxoidAnaeroGen system
145 (Miller *et al.*, 1995). Carbon source utilization, catalase, oxidase and gelatinase activities,

146 hydrolysis of starch, Tween 20, Tween 40, Tween 60 and Tween 80, nitrate reduction,
147 urease and H₂S production were determined using standard methods (Gerhardt *et al.*,
148 1994; Lanyi, 1987; MacFaddin, 2000).

149

150 Strain YIM 64602^T was a Gram- positive actinobacterium and it can grow well on ISP 2
151 and ISP 4 media, formed yellow-white to white substrate mycelia and yellow to white
152 aerial mycelia. Yellow diffusible pigments were only produced on the ISP 2 medium.
153 Substrate mycelia showed extensive branching without fragmenting (Fig. S4). It could
154 not grow under anaerobic conditions. The isolates grew over the temperature range
155 15-40°C, pH range 6.0-9.0 and NaCl concentration range 0-3% (w/v). Optimal growth
156 was observed at 28 °C and at pH 7.0 without NaCl. Other physiological characteristics
157 are given in Table 1 and in the species description.

158

159 In view of the combination of morphological, physiological, chemotaxonomic and
160 genotypic data (Table 1) discussed here, such as Gram-positive, strictly aerobic,
161 filamentous characters, containing *meso*-DAP, the major polar lipids are DPG, PE and
162 PME, the predominant menaquinones are MK-10(H₆), MK-10(H₄) and MK-11(H₄), the
163 fatty-acid pattern was mainly composed of the saturated branched-chain acids, it is
164 evident that strain YIM 64602^T belongs to the genus *Stackebrandtia*. However, a few
165 characteristics that are unique to strain YIM 64602^T differentiate it from *S. albiflava*
166 YIM 45751^T and *S. nassauensis* DSM 44728^T (Table 1). YIM 64602^T and *S.*
167 *nassauensis* DSM 44728^T can utilize trehalose, while *S. albiflava* YIM 45751^T can't;
168 *S. nassauensis* DSM 44728^T and *S. albiflava* YIM 45751^T can utilize raffinose, fructose
169 and glucose, hydrolyse gelatin, while YIM 64602^T can't. They can also be
170 differentiated based on the growth temperature, pH and NaCl tolerance. Based on the
171 phenotypic, chemotaxonomic and genotypic data presented above, we propose that
172 strain YIM 64602^T represents a novel species within the genus *Stackebrandtia*, and the
173 name *Stackebrandtia endophytica* sp. nov. is proposed.

174

175 **Description of *Stackebrandtia endophytica* sp. nov.**

176 *Stackebrandtia endophytica*(en.do.phy'ti.ca. Gr. pref. *endo* within; Gr. n. *phyton* plant; L.
177 fem. suff.-*ica* adjectival suffix used with the sense of belonging to; N.L. fem. adj.
178 *endophytica* within plant, endophytic, pertaining to the isolation from plant tissues).

179

180 Good growth occurs on ISP 2 and ISP 4 (produces white to yellowish white substrate
181 mycelia and aerial mycelia). Weakly grow is observed on ISP 3, ISP 4, ISP 5 and
182 Czapek's sucrose agar. Yellow soluble pigments are produced on the ISP 2 media.
183 Grows over the temperature range 15-40°C, pH range 6.0-9.0 and NaCl concentration
184 range 0-3% (w/v). Catalase-positive and oxidase-negative, nitrate is reduced to nitrite.
185 H₂S is not produced. The strain can degrade starch, Tweens 20, 60, 80 and urea, but not
186 gelatin or Tweens 40. Utilizes trehalose, D-galactose, sucrose and xylose as sole carbon;
187 arabinose, D-mannose, L-sorbose, succinic acid, inositol, raffinose, cellobiose, D-
188 fructose, maltose or glucose are not utilized.

189

190 The type strain is YIM 64602^T (=BCRC 16954^T=DSM 45928^T), isolated from
191 surface-sterilized stems of *Tripterygium wilfordii*, collected in Yunnan Province,
192 south-west China.

193

194

195 **Acknowledgements**

196 This research was supported by the National Natural Science Foundation of China (No.
197 U0932601, 81102806).

198

199 **References**

- 200 **Collins, M. D., Pirouz, T., Goodfellow, M. & Minnikin, D. E. (1977).** Distribution of
201 menaquinones in actinomycetes and corynebacteria. *J Gen Microbiol* **100**, 221-230.
- 202 **Cui, X. L., Mao, P. H., Zeng, M., Li, W. J., Zhang, L. P., Xu, L. H. & Jiang, C. L.**
203 **(2001).** *Streptomonospora salina* gen. nov., sp. nov., a new member of the family
204 *Nocardiopsaceae*. *Int J Syst Evol Microbiol* **51**, 357–363.

205 **Embley, T. M. & Wait, R. (1994).** Structural lipids of eubacteria. *Chemical Methods in*
206 *Prokaryotic Systematics*, 121-161. Edited by M. Goodfellow & A. G. O'Donnell.
207 Chichester: Wiley.

208 **Ezaki, T., Hashimoto, Y. & Yabuuchi, E. (1989).** Fluorometric deoxyribonucleic
209 acid-deoxyribonucleic acid hybridization in microdilution wells as an alternative to
210 membrane filter hybridization in which radioisotopes are used to determine genetic
211 relatedness among bacterial strains. *Int J Syst Bacteriol* **39**, 224-229.

212 **Felsenstein, J. (1981).** Evolutionary trees from DNA sequences: a maximum likelihood
213 approach. *J Mol Evol* **17**, 368-376.

214 **Felsenstein, J. (1985).** Confidence limits on phylogenies: an approach using the
215 bootstrap. *Evolution* **39**, 783-791.

216 **Fitch, W. M. (1971).** Toward defining the course of evolution: minimum change for a
217 specific tree topology. *Syst Zool* **20**, 406-416.

218 **Gerhardt, P., Murray, R. G. E., Wood, W. A. & Krieg, N. R. (editors) (1994).**
219 *Methods for General and Molecular Bacteriology*. Washington, DC: American Society
220 for Microbiology.

221 **Guan, T. W., Tang, S. K., Wu, J. Y., Zhi, X. Y., Xu, L. H., Zhang, L. L. & Li, W. J.**
222 **(2009).** *Haloglycomyces albus* gen. nov., sp. nov., a halophilic, filamentous
223 actinomycete of the family *Glycomycetaceae*. *Int J Syst Evol Microbiol* **59**, 1297-1301.

224 **Hasegawa, T., Takizawa, M. & Tanida, S. (1983).** A rapid analysis for chemical
225 grouping of aerobic actinomycetes. *J Gen Appl Microbiol* **29**, 319-322.

226 **He, L., Li, W., Huang, Y., Wang, L. M., Liu, Z. H., Lanoot, B. J., Vancanneyt, M. &**
227 **Swings, J. (2005).** *Streptomyces jietaisiensis* sp. nov., isolated from soil in northern
228 China. *Int J Syst Evol Microbiol* **55**, 1939-1944.

229 **Kelly, K. L. (1964).** Inter-Society Color Council–National Bureau of Standards Color
230 Name Charts Illustrated with Centroid Colors. Washington, DC: US Government
231 Printing Office.

232 **Kim, O. S., Cho, Y. J., Lee, K., Yoon, S. H., Kim, M., Na, H., Park, S. C., Jeon, Y.**
233 **S., Lee, J. H., Yi, H., Won, S., Chun, J. (2012).** Introducing EzTaxon-e: a

234 prokaryotic 16S rRNA Gene sequence database with phylotypes that represent
235 uncultured species. *Int J Syst Evol Microbiol* **62**, 716-721.

236 **Labeda, D. P. & Kroppenstedt, R. M. (2004)**. Emended description of the genus
237 *Glycomyces* and description of *Glycomyces algeriensis* sp. nov., *Glycomyces*
238 *arizonensis* sp. nov. and *Glycomyces lechevalierae* sp. nov. *Int J Syst Evol Microbiol* **54**,
239 2343–2346.

240 **Labeda, D. P. & Kroppenstedt, R. M. (2005)**. *Stackebrandtia nassauensis* gen. nov., sp.
241 nov. and emended description of the family *Glycomycetaceae*. *Int J Syst Evol Microbiol*
242 **55**, 1687-1691.

243 **Labeda, D. P., Testa, R. T., Lechevalier, M. P. & Lechevalier, H. A. (1985)**.
244 *Glycomyces*, a new genus of the *Actinomycetales*. *Int J Syst Bacteriol* **35**, 417-421.

245 **Lanyi, B. (1987)**. Classical and rapid identification methods for medically important
246 bacteria. *Methods Microbiol* **19**, 1-67.

247 **Li, J., Zhao, G. Z., Chen, H. H., Wang, H. B., Qin, S., Zhu, W. Y., Xu, L.H., Jiang,**
248 **C. L. & Li, W. J. (2008)**. Antitumour and antimicrobial activities of endophytic
249 streptomycetes from pharmaceutical plants in rainforest. *Lett Appl Microbiol* **47**,
250 574-580.

251 **Li, W. J., Xu, P., Schumann, P., Zhang, Y. Q., Pukall, R., Xu, L. H., Stackebrandt,**
252 **E. & Jiang, C. L. (2007)**. *Georgenia ruanii* sp. nov., a novel actinobacterium isolated
253 from forest soil in Yunnan (China), and emended description of the genus *Georgenia*.
254 *Int J Syst Evol Microbiol* **57**, 1424-1428.

255 **MacFaddin, J. F. (2000)**. *Biochemical Tests for the Identification of Medical Bacteria*,
256 3rd edn. Baltimore, MD: Williams & Wilkins.

257 **Mesbah, M., Premachandran, U. & Whitman, W. B. (1989)**. Precise measurement of
258 the G+C content of deoxyribonucleic acid by high performance liquid chromatography.
259 *Int J Syst Bacteriol* **39**, 159-167.

260 **Miller, P. H., Wiggs, L. S. & Miller, J. M. (1995)**. Evaluation of AnaeroGen system for
261 growth of anaerobic bacteria. *J Clin Microbiol* **33**, 2388-2391.

262 **Saitou, N. & Nei, M. (1987)**. The neighbor-joining method: a new method for
263 reconstructing phylogenetic trees. *Mol Biol Evol* **4**, 406-425.

264 **Shirling, E. B. & Gottlieb, D. (1966).** Methods for characterization of *Streptomyces*
265 species. *Int J Syst Bacteriol* **16**, 313-340.

266 **Tamaoka, J., Katayama-Fujimura, Y. & Kuraishi, H. (1983).** Analysis of bacterial
267 menaquinone mixtures by high performance liquid chromatography. *J Appl Bacteriol* **54**,
268 31-36.

269 **Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M. & Kumar, S. (2011).**
270 MEGA5: molecular evolutionary genetics analysis using maximum likelihood,
271 evolutionary distance, and maximum parsimony methods. *Mol Biol Evol* **28**, 2731-2739.

272 **Tang, S. K., Wang, Y., Chen, Y., Lou, K., Cao, L. L., Xu, L. H. & Li, W. J. (2009).**
273 *Zhihengliuella alba* sp. nov., and emended description of the genus *Zhihengliuella*. *Int J*
274 *Syst Evol Microbiol* **59**, 2025-2031.

275 **Thompson, J. D., Gibson, T. J., Plewniak, F., Jeanmougin, F. & Higgins, D. G.**
276 **(1997).** The CLUSTAL_X windows interface: flexible strategies for multiple sequence
277 alignment aided by quality analysis tools. *Nucleic Acids Res* **25**, 4876-4882.

278 **Wang, Y. X., Zhi, X. Y., Zhang, Y. Q., Cui, X. L., Xu, L. H. & Li, W. J. (2009).**
279 *Stackebrandtia albiflava* sp. nov. and emended description of the genus *Stackebrandtia*.
280 *Int J Syst Evol Microbiol* **59**, 574-577.

281 **Xu, P., Li, W. J., Tang, S. K., Zhang, Y. Q., Chen, G. Z., Chen, H. H., Xu, L. H. &**
282 **Jiang, C. L. (2005).** *Naxibacter alkalitolerans* gen. nov., sp. nov., a novel member of the
283 family ‘*Oxalobacteraceae*’ isolated from China. *Int J Syst Evol Microbiol* **55**,
284 1149-1153.

285

286

287

288

289 **Fig. 1.** Neighbour-joining tree based on 16S rRNA gene sequences showing the
290 relationships of strain YIM 64602^T and the type species of the related genera. Bootstrap
291 values (>50 %) based on 1000 replicates are shown at the branch nodes. Asterisks
292 indicate that the corresponding branches were also recovered in trees generated with the
293 maximum-parsimony and maximum-likelihood methods. *Dietzia maris* ATCC 35013^T
294 (X79290) was used as the outgroup. Bar, 0.01 substitutions per nucleotide position.

295

Table 1. Differential characteristics of strain YIM 64602^T and related species

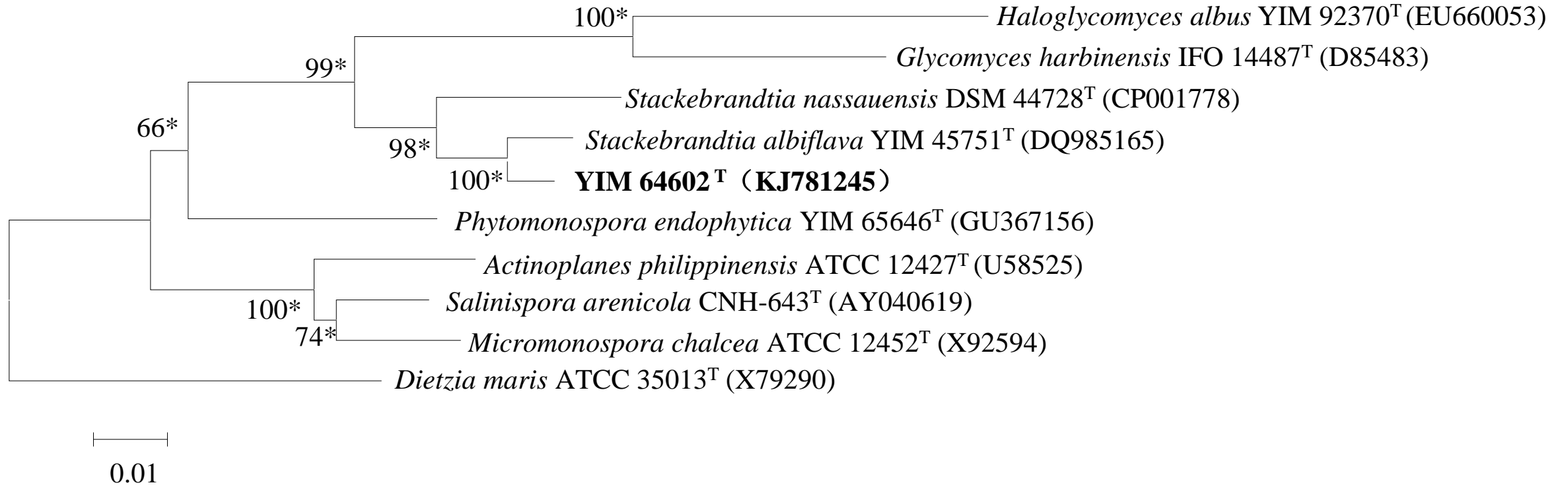
Character	1	2	3 [†]
Temperature (°C)	15-40	20-37	15-37
Growth on ISP 4	+	-	ND
Soluble pigment	+	-	ND
NaCl(%)	3	-	4-9
pH	6-9	6-8	ND
Gelatinase	-	+	+
Utilization of:			
Trehalose	+	-	+
Raffinose	-	+	+
Fructose	-	+	+
Glucose	-	+	+
Predominant menaquinones	MK-10(H ₆), MK-10(H ₄) and MK-11(H ₄)	MK-10(H ₆), MK-10(H ₄), MK-11(H ₆) and MK-11(H ₄)	MK-10(H ₆), MK-10(H ₄), MK-11(H ₆) and MK-11(H ₄)
major fatty acids	iso -C _{16:0} , anteiso -C _{17:0} , iso -C _{15:0} and iso -C _{17:0}	anteiso -C _{17:0} , iso -C _{15:0} and iso -C _{17:0}	anteiso -C _{17:0} , 2-hydroxy- anteiso -C _{17:0} , iso -C _{17:0} , iso -C _{16:0} and iso -C _{15:0}
G + C content (mol %)	72.4	69.4*	72.4

297 Taxa: 1, YIM 67072^T; 2, *S. albiflava* YIM 45751^T; 3, *S. nassauensis* DSM 44728^T.

298 *Data from Wang *et al.* (2009). †Data from Wang *et al.* (2009) and Labeda &

299 Kroppenstedt, (2005). +, Positive; -, negative; ND, no data.

300



As is on our Paper ahead of Press page 11: **Fig. 1.**

Supplementary Material Files

[Click here to download Supplementary Material Files: Supplementary materials-revised version.pdf](#)