

Available online at www.sciencedirect.com



Chinese Chemical Letters 19 (2008) 1215-1217



## Two new lignans from Dipteronia dyeriana

Rong Guo a,b, Min Luo c, Chun Lin Long a,\*, Ma Lin Li c,\*, Zhi Qin Ouyang d, Yi Ping Zhou c, Yue Hu Wang a, Xing Yu Li a,b, Ya Na Sin a,b

\*Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650204, China

b Yunnan Agricultural University, Kunming 650201, China

c Yunnan Laboratory of Pharmacology for Natural Products, Kunming Medical College, Kunming 650031, China

d Yunnan Introduction & Propagation Center for Rare & Endangered Plants, Kunming 650032, China

Received 7 March 2008

## Abstract

A new sesquilignan, 7',8'-didehydroherpetotriol (1), and a new lignan glycoside, (+)-isolariciresinol-9'-O- $\alpha$ -L-rhamnopyranosyl-(1  $\rightarrow$  6)- $\beta$ -D-glucopyranoside (2), were isolated from the branches of *Dipteronia dyeriana*. Their structures were elucidated by spectroscopic methods and chemical evidence. Compound 1 possessed inhibitory activity against human leukaemia K562 cells with an IC<sub>50</sub> value of 39  $\mu$ mol/L.

© 2008 Chun Lin Long. Published by Elsevier B.V. on behalf of Chinese Chemical Society. All rights reserved.

Keywords: Aceraceae; Dipteronia dyeriana; Lignans; Didehydroherpetotriol; Leukaemia

Aceraceae comprises two genera, Acer and Dipteronia [1]. The plants of genus Acer contains various bioactive substances, such as triterpenoids with antitumor activity [2], stilbene glycosides with hepatoprotective and antioxidative activity [3,4], and diarylheptanoids as inhibitors of nitric oxide production [5]. To the best of our knowledge, there was not any report of chemical constituents and bioactivity on the genus Dipteronia. In our program to search antitumor agents from natural products, we have investigated Dipteronia dyeriana Henry collected from Pingbian County of Yunnan Province. In this paper, we report the structure elucidation of compounds 1 and 2 isolated from the branches of this plant, along with the inhibitory effects of compound 1 against human leukaemia K562 and human hepatoma HepG2 cells.

Compound 1 was obtained as pale yellow powder,  $[\alpha]_D^{24.8} + 54.0$  (c 0.50, MeOH), with UV (MeOH) absorption bands of 300 (4.11), 279 (4.15) and 234 (4.21) nm ( $\lambda_{max}$ : log  $\varepsilon$ ). Its molecular formula was assigned as  $C_{30}H_{30}O_9$  on the basis of the HR-ESI-MS m/z [M-H]<sup>-</sup> 533.1804 (calcd. 533.1811). In its IR (KBr) spectrum, absorption bands due to hydroxyl groups and aromatic ring were observed at 3406, 1608 and 1517 cm<sup>-1</sup>. The <sup>13</sup>C NMR spectrum (Table 1) of 1 exhibited 30 carbon signals, including 13 quaternary carbons, 11 methines, three methylenes and three methoxy carbons. Extensive analysis and comparison of the <sup>1</sup>H and <sup>13</sup>C NMR spectra of 1 with herpetotriol [6] suggested that compound 1 was a sesquilignan. However, instead of two dihydrobenzofuran segments in herpetotriol, there was one dihydrobenzofuran [ $\delta$  150.4 (C-4'), 134.2 (C-5'), 89.6 (C-7") and 55.1 (C-8")] and one benzofuran segment [ $\delta$  143.9

E-mail addresses: long@mail.kib.ac.cn (C.L. Long), limalinb@vip.163.com (M.L. Li).

1001-8417/\$ - see front matter © 2008 Chun Lin Long. Published by Elsevier B.V. on behalf of Chinese Chemical Society. All rights reserved. doi:10.1016/j.cclet.2008.06.051

<sup>\*</sup> Corresponding authors.

Table 1  $^{1}$  H NMR (500 MHz) and  $^{13}$  C NMR (100 MHz) data of 1 (CD3OD, TMS,  $\delta$  ppm)

No.	$\delta_{ m H}$	$\delta_{ m C}$	No.	$\delta_{ m H}$	$\delta_{\mathbf{C}}$
1		134.6	7′		155.7
2	6.93 (d, 1H, 1.0)	105.9	8′		115.1
3		146.4	9′	4.78 (br s, 2H)	55.4
4		143.9	1"		130.8
5		132.9	2"	6.95 (d, 1H, 2.0)	110.6
6	7.25 (d, 1H, 1.0)	111.3	3"		149.2
7	6.65 (br d, 1H, 16.0)	132.4	4"		147.7
8	6.31 (ddd, 1H, 16.0, 6.0, 6.0)	128.8	5"	6.75 (d, 1H, 8.0)	116.2
9	4.21 (dd, 2H, 6.0, 1.5)	63.8	6"	6.82 (dd, 1H, 8.0, 2.0)	119.8
1'		125.1	7"	5.56 (d. 1H. 6.5)	89.6
2'	7.39 (d, 1H, 1.0)	113.0	8"	3.56 (dd, 1H, 12.5, 6.5)	55.1
3'		145.7	9"	3.84 (m, 2H)	64.6
4'		150.4	3-OCH3	3.97 (s, 3H)	56.5
5′		134.2	3'-OCH3	3.89 (s, 3H)	56.8
6'	7.36 (d, 1H, 1.0)	117.5	3"-OCH₃	3.79 (s, 3H)	56.4

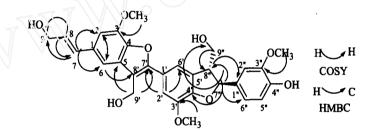


Fig. 1. The chemical structure and key <sup>1</sup>H-<sup>1</sup>H COSY and HMBC correlations for 1.

(C-4), 132.9 (C-5), 155.7 (C-7'), 115.1 (C-8'),] in 1. The planar structure of 1 was established by the HMBC correlations (Fig. 1). A trans-configuration between H-7" and H-8" was inferred from the coupling constant (J = 6.5 Hz) [7]. Thus, 1 was identified as 7',8'-didehydroherpetotriol. The absolute configuration of 1 has not been confirmed.

Table 2  $^{1}H$  NMR (400 MHz) and  $^{13}C$  NMR (100 MHz) data of 2 (CD3OD, TMS,  $\delta$  ppm)

No.	$\delta_{H}$	$\delta_{ m C}$	No.	$\delta_{ m H}$	$\delta_{\mathbf{C}}$
1		129.1	8′	1.85 (m, 1H)	45.8
2		134.4	9'	4.00 (dd, 1H, 9.6, 2.4)	69.2
				3.22 (m, 1H)	
3	6.17 (s, 1H)	117.4	5-OCH <sub>3</sub>	3.79 (s, 3H)	56.4
4		145.1	3'-OCH <sub>3</sub>	3.80 (s, 3H)	56.5
5		147.2	1"	4.07 (d, 1H, 8.0)	105.1
6	6.64 (s, 1H)	112.4	2"	3.19 (dd, 1H, 9.2, 8.0)	75.1
7	2.86 (dd, 1H, 16.0, 11.2)	33.9	3"	3.32 (m, 1H)	78.1
	2.79 (dd, 1H, 16.0, 5.2)				
8	2.08 (m, 1H)	39.4	4"	3.26 (m, 1H)	71.8
9	3.77 (m, 2H)	65.0	5"	3.35 (m, 1H)	76.8
1'		138.7	6"	3.93 (dd, 1H, 11.6, 1.6)	68.2
				3.57 (m, 1H)	
2'	6.78 (d, 1H, 2.0)	114.3	1′″	4.74 (d, 1H, 1.2)	102.3
3'		148.9	2′″	3.80 (m, 1H)	72.2
4'		145.8	3′″	3.64 (m, 1H)	72.3
5'	6.74 (d, 1H, 8.0)	116.1	4′″	3.34 (m, 1H)	74.0
6'	6.64 (dd, 1H, 8.0, 2.0)	123.2	5′″	3.63 (m, 1H)	69.8
7'	4.08 (d, 1H, 11.2)	47.9	6′″	1.22 (d, 3H, 6.0)	18.1

Fig. 2. The chemical structure and HMBC correlations for 2.

Compound 2 was isolated as white powder,  $[\alpha]_D^{24.7} + 3.9$  (c 0.43, MeOH). UV (MeOH) absorption band was at  $\lambda_{\text{max}}$  (log  $\varepsilon$ ): 283 (3.77) nm. The FABMS spectrum of compound 2 showed a molecular ion peak at m/z 667 [M-H]<sup>-</sup>, besides significant fragment peaks at m/z 521 [M-(rhamnose-OH)]<sup>-</sup>. Its molecular formula was determined as  $C_{32}H_{44}O_{15}$  on the basis of the HR-ESI-MS m/z [M-H]<sup>-</sup> 667.2586 (calcd. 667.2601). In its IR (KBr) spectrum, absorption bands due to hydroxyl groups and aromatic double bonds were observed at 3424, 1627 and 1513 cm<sup>-1</sup>. The <sup>13</sup>C NMR spectrum (Table 2) of 2 showed 32 carbon signals, including two methoxyl, one methyl, four methylenes, eighteen methines and seven quaternary carbons. There was a (+) or (-) isolariciresinol moiety in 2 by comparison of the <sup>1</sup>H and <sup>13</sup>C NMR spectra of 2 with those in the literatures [8,9]. The spectral data of  $\delta_H$  4.07 (d, 1H, J = 8.0 Hz) and  $\delta_C$  105.1 (d), 78.1 (d), 76.8 (d), 75.1 (d), 71.8 (d), 68.2 (t) showed the presence of a  $\beta$ -glucose moiety [10]. In addition, the remaining six carbon signals [ $\delta$  102.3 (d), 74.0 (d), 72.3 (d), 72.2 (d), 69.8 (d), 18.1 (q)] and an anomeric proton resonance at  $\delta$  5.49 (d, 1H, J = 1.2 Hz) was the characteristic of  $\alpha$ -rhamnoside [10]. Acidic hydrolysis of 2 in 2 mol/L HCl water solution [11] gave a rhamnose, a  $\beta$ -D-glucose ([ $\alpha$ ]<sub>D</sub><sup>20.6</sup> + 83.3 (c 0.060, H<sub>2</sub>O)), and a (+)-isolariciresinol ([ $\alpha$ ]<sub>D</sub><sup>20.7</sup> + 25.6 (c 0.065, Me<sub>2</sub>CO)). And the location of the glucose and rhamnose were established on the basis of HMBC correlations from  $\delta$  4.07 (d, 1H, H-1") to  $\delta$  69.2 (C-9") and  $\delta$  4.74 (d, 1H, H-1") to  $\delta$  68.2 (C-6") (Fig. 2). Thus, the structure of 2 was elucidated as (+)-isolariciresinol-9'-O- $\alpha$ -L-rhamnopyranosyl-(1  $\rightarrow$  6)- $\beta$ -D-glucopyranoside.

The bioassay results showed that 1 inhibited the growth of K562 and HepG2 cells with an IC<sub>50</sub> of 39 and 312  $\mu$ mol/L, respectively [12,13].

## Acknowledgments

This work was financially supported by grants from the Ministry of Science and Technology of China (2005DKA21006) and the Knowledge Innovation Project of Chinese Academy of Sciences.

## References

- [1] T.Z. Xu, Acta Botanica Yunnanica 18 (1996) 43.
- [2] S.M. Kupchan, M. Takasugi, R.M. Smith, P.S. Steyn, J. Org. Chem. 36 (1971) 1972.
- [3] H. Yang, S.H. Sung, Y.C. Kim, J. Nat. Prod. 68 (2005) 101.
- [4] H. Yang, M.K. Lee, Y.C. Kim, J. Agric. Food Chem. 53 (2005) 4182.
- [5] T. Morikawa, J. Tao, I. Toguchida, H. Matsuda, M. Yoshikawa, J. Nat. Prod. 66 (2003) 86.
- [6] J. Favre-Bonvin, M. Kaouadji, A.-M. Mariotte, Tetrahedron Lett. 43 (1978) 4111.
- [7] Y.Z. Wang, H. Chen, X.K. Zheng, W.S. Feng, Chin. Chem. Lett. 18 (2007) 1224.
- [8] T. Popoff, O. Theander, Acta Chem. Scand. B 31 (1977) 329.
- [9] L.N. Lundgren, T. Popoff, O. Theander, Phytochemistry 20 (1981) 1967.
- [10] Y.H. Wang, J.H. Wang, H.P. He, H. Zhou, X.W. Yang, C.S. Li, X.J. Hao, J. Asian Nat. Prod. Res. 10 (2008) 25.
- [11] H.J. Kim, E.R. Woo, H. Park, J. Nat. Prod. 57 (1994) 581.
- [12] K. Likhitwitayawuid, C.K. Angerhofer, G.A. Cordell, J.M. Pezzuto, N. Ruangrungsi, J. Nat. Prod. 56 (1993) 30.
- [13] E.K. Seo, M.C. Wani, M.E. Wall, H. Navarro, R. Mukherjee, N.R. Farnsworth, A.D. Kinghorn, Phytochemistry 55 (2000) 35.