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A new dinorclerone diterpenoid glycoside from *Tinospora sinensis*

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A new dinorclerone diterpenoid glycoside, named 1-deacetyltinosposide A (1), was isolated from the stem of *Tinospora sinensis* together with 10 known compounds. Their structures were elucidated on the basis of extensive spectroscopic techniques (MS, IR, 1D and 2D NMR experiments).

Keywords: Tinospora sinensis; Menispermaceae; 1-deacetyltinosposide A

1. Introduction

In the Menispermaceae, the genus *Tinospora* comprises about 20 species, which are mainly distributed in the tropical parts of the eastern hemisphere (Hou, 1998). Various research groups have been working on the chemical constituents of this genus, especially *Tinospora cordifolia*, which is an important medicinal plant, and it has been widely cultivated and used in Ayurvedic preparations for treatment of various ailments throughout the Indian subcontinent (Gangan, Pradhan, Sipahimalani, Bhave, & Patil, 1997). More than 100 clerodane-type diterpenoids and their glycosides have been isolated, as well as alkaloids, sesquiterpenes, lignans and ecdysones (Gangan, Pradhan, & Sipahimalani, 1997; Guo et al., 1999; Hungerford, Sands, & Kitching, 1998; Maurya, Dhar, & Handa, 1997; Ragasa, Cruz, Gula, & Rideout, 2000) from the genus *Tinospora*.

In China, the stems of *Tinospora sinensis* Merr. have been used as folk medicine to treat strains of the lumber muscles, rheumatism and bruises (Jiangsu New Medicinal College, 1977). Its anti-inflammatory, immunomodulatory and antidiabetic activities have been demonstrated by pharmacological experiments (Li, Lin, Myers, & Leach, 2003; Manjrekar, Jolly, & Narayanan, 2000; Yonemitsu, Fukuda, & Kimura, 1993). In previous chemical studies, the isolation of two dinorditerpene glucosides, tinosinesides A (2) and B, was reported (Yonemistu, Fukuda, Kimura, Isobe, & Komori, 1995), as well as three phenylpropanoid glycosides (Li et al., 2004; Yonemitsu et al., 1993), tinosinen (8), 4-allyl-2-methoxyphenyl 6-O- β -D-apiofuranosyl (1 \rightarrow 6)- β -D-glucopyranoside, icariside D1, seven lignan glucosides (Li et al., 2004), tinosposides A (2) and B, tanegoside (6), (-)-pinoresinol O- β -D-glucopyranoside (5), (-)-pinoresinol monomethyl ether O- β -D-glucopyranoside, (-)-syringaresinol O- β -D-glucopyranoside, and (-)-isolariciresinol 3 α -O- β -D-glucopyranoside, all from T. sinensis. During our chemical investigation of this plant, we obtained a

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new dinorclerone diterpenoid glycoside, named 1-deacetyltinosposide A (1), together with 10 known compounds: tinosineside A (2), tinocordifolioside (3), palmatine (4), (-)-pinoresinol 4-O- β -D-glucopyranoside (5), 8'-epitanegool (6), syringin (7), (E)-1-(3-hydroxy1-propenyl)-3,5-dimethoxyphenyl-4-O- β -D-apiofuranosyl-(1 \rightarrow 3)- β -D-glucopyranoside (8), stigmasta-5, 11 (12)-dien-3 β -ol (9), β -sitosterol (10), and daucosterol (11). This article describes the isolation and structural elucidation of a new dinorclerone diterpenoid glycoside, 1-deacetyltinosposide A (1), besides the 10 known compounds.

2. Results and discussion

Compound 1, a colourless amorphous powder, showed a [M–H]⁻ ion peak at m/z 513.1956 in the negative ion HRFAB⁻MS, indicating the molecular formula $C_{24}H_{34}O_{12}$. The IR spectrum showed characteristic absorptions for hydroxyl groups (3415 cm⁻¹), a δ -lactone (1722 cm⁻¹), and a furan ring (1505, 1022, 875 cm⁻¹). The UV spectrum [202 nm (log ε = 3.91)] was indicative of the presence of a furan moiety. The presence of a furan ring was also confirmed by a positive Ehrlich colour test. Its negative FAB⁻MS spectrum gave a fragment ion at m/z 351 [M–C₆H₁₀O₅ – H]⁻, suggesting the presence of a glucopyranosyl unit in the molecule of 1, which was supported by the presence of six carbon signals of the glucopyranosyl at δ = 101.3 (d), 74.9 (d), 78.7 (d), 71.9 (d), 79.1 (d) and 63.1 (t) in its ¹³C NMR spectrum. Furthermore, the coupling constant of the anomeric proton of a sugar moiety (J = 7.8 Hz) suggested the glycosidic linkage to have a β -glucopyranose.

The aglycone moiety of 1 has 18 carbons besides the sugar moiety. The 1H NMR spectrum of this compound was very similar to that of clerodane furano-diterpenes, and the assignments are given in Table 1. The signals at δ 7.47 (d, J=1.6 Hz, 1H), 7.42 (br s, 1H), and 6.49 (d, J=1.6 Hz, 1H) were assigned two α -protons and β -proton of a β -substituted furan moiety. A signal for the angular methyl group was observed as a singlet at δ =1.51. By detailed comparison of the 1H , ^{13}C NMR data, and combined

C-4', 5'

| Position | $\delta_{ m H}$ | $\delta_{ m C}$ | HMBC |
|----------|--------------------------------|-----------------|------------------------|
| 1 | 3.99 (dd, J = 2.9, 10.8 Hz) | 74.1 (d) | C-10 |
| 2 3 | 4.33 (m) | 71.7 (d) | C-1, 3, 4,10 |
| 3 | 1.61 (m) | 32.8 (t) | C-1, 2, 4, 5 |
| | 2.64 (d, J = 15.2 Hz) | | C-4, 5 |
| 4 | 4.08 (m) | 76.2 (d) | C-2, 6, 10, 1' |
| 5 | 1.68 (m) | 39.2 (d) | C-1, 4, 7, 10 |
| 6 | 1.61 (m) | 26.8 (t) | C-7, 8, 10 |
| | 2.35 (m) | | C-5, 7, 8 |
| 7 | 1.88 (m) | 30.8 (t) | C-5, 6, 8, 17 |
| | 2.77 (d, J = 12.0 Hz) | | C-5, 6 |
| 8 | | 76.3 (s) | |
| 9 | | 40.8 (s) | |
| 10 | 2.84 (t, J = 10.8 Hz) | 36.4 (d) | C-1, 5, 20 |
| 11 | 3.04 (t, J = 12.3 Hz) | 37.6 (d) | C-8, 9, 10, 12, 13, 20 |
| | 3.38 (dd, J = 3.5, 13.4 Hz) | | C-8, 9, 20 |
| 12 | 6.11 (dd, $J = 3.5$, 12.3 Hz) | 72.2 (d) | C-11, 13, 14, 16, 17 |
| 13 | | 127.3 (s) | |
| 14 | 6.49 (d, J = 1.6 Hz) | 109.8 (d) | C-12, 13, 15, 16 |
| 15 | 7.47 (d, J = 1.6 Hz) | 143.8 (d) | C-13, 14, 16 |
| 16 | 7.42 (br s) | 140.1 (d) | C-12, 13, 14, 15 |
| 17 | | 173.6 (s) | |
| 20 | 1.51 (s) | 14.4 (q) | C-8, 9, 10, 11 |
| 1' | 4.94 (d, J = 7.8 Hz) | 101.3 (d) | C-4, 3', 5' |
| 2' | 3.87 (m) | 74.9 (d) | C-1', 3', 4' |
| 3′ | 4.21 (m) | 78.7 (d) | C-1', 2', 4' |
| 4′ | 4.04 (m) | 71.9 (d) | C-3', 5', 6' |
| 5' | 3.91 (m) | 79.1 (d) | C-1', 4', 6' |
| 6' | 4.54 (dd, J = 2.1, 11.8 Hz) | 63.1 (t) | C-4', 5' |

Table 1. ¹H- and ¹³C NMR and spectra data, HMBC correlations for 1 (in C₅D₅N).

¹H⁻¹H COSY, and HMBC of **1**, compound **1** had the same skeleton as tinosineside A (**2**) (Li et al., 2004), except for the presence of a hydroxyl group and the disappearance of a acetoxyl group at C-1 in **1**. The relative stereochemistry of **1** was fixed on the basis of a ROESY experiment. The important correlations of H-1 with H-2, H-5, Me-20, H-5 with H-4, Me-20, and H-10 with H-12 were observed, and indicated that H-1, H-2, H-4, H-5 and Me-20 were on the same side of the molecule whereas H-10 and H-12 were on the opposite side (Figure 1). On the basis of these data, the structure of **1** was elucidated as 1-deacetyltinosposide A.

1-Deacetyltinosposide A (1) and tinosposide A (2) were tested for *in vitro* cytotoxicity against HepG2 and Raji cells. However, they were non-cytotoxic.

3. Experimental

3.1. General experimental procedures

4.29 (m)

UV spectra were obtained on a UV-210A spectrometer. IR spectra were taken in KBr on a Bio-Rad FTS-135 infrared spectrophotometer. Optical rotations were performed on a JASCO DIP-370 digital polarimeter. Negative FAB mass spectra were measured on a VG Auto spec-3000 spectrometer and high-resolution ESI mass spectra were recorded on an

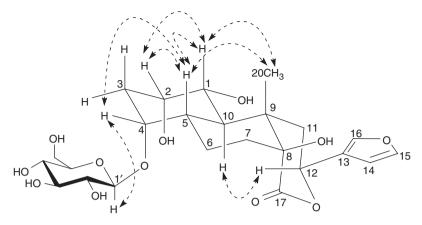


Figure 1. Key ROESY correlations of 1-deacetyltinosposide A (1).

API Qstar Pulsar instrument. 1D and 2D NMR experiments were performed on Bruker AM-400 and DRX-500 instruments, with TMS as the internal standard. Column chromatography was performed on silica gel (200–300 mesh, Qingdao Marine Chemical Inc., Qingdao, P.R. China) or on silica gel H (10–40 μm, Qingdao Marine Chemical Inc.). MPLC were performed on a BÜCHI Pump Module C-605, a BÜCHI Pump Manager C-615, and a BÜCHI Fraction Collector C-660.

3.2. Plant material

The stems of *T. sinensis* were collected from Xishuangbanna, Yunnan province, P.R. China, in August 2005. The plants were identified by Prof. Guo-Da Tao, Xishuangbanna Tropical Botanical Garden, Chinese Academy of Science. A voucher specimen (No. 200503) is deposited in the State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany.

3.3. Extraction and isolation

The dried and powdered stems of T. sinensis (5.0 kg) were extracted with 70% Me₂CO and filtered at RT. The filtrate was concentrated and partitioned successively between EtOAc and water, then n-BuOH and water. The EtOAc extract (20 g) was applied to column chromatography (petroleum ether: AcOEt 9:1, 8:2, 5:5 v/v) to afford 9 (14 mg), 10 (2.0 g), and 11 (1.3 g). The n-BuOH extract (40 g) was applied to column chromatography (CHCl₃: MeOH 20:1, 9:1, 8.5:1.5, 8:2 v/v) to afford 1 (94 mg), 2 (127 mg), 3 (30 mg), 4 (7 mg), 5 (16 mg), 6 (18 mg), 7 (35 mg), and 8 (30 mg).

3.3.1. 1-deacetyltinosposide A (1)

 $C_{24}H_{34}O_{12}$, colourless amorphous powder; $[\alpha]_D^{25} = -11.7$ (c = 0.3, pyridine); UV (MeOH) λ_{max} (log ε): 202 nm (3.91); IR (KBr) ν_{max} (cm⁻¹): 3415, 2925, 2874, 1722, 1505, 1022, 875; ¹H NMR and ¹³C NMR, see Table 1; Negative FAB-MS m/z: 513 [M-H]⁻, 351 [M-glucose – H]⁻; HRESI-MS m/z: 513.1956 [M-H]⁻ ($C_{24}H_{33}O_{12}$ Calcd 513.1972).

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