

This article was downloaded by: [Kunming Institute of Botany]

On: 18 May 2015, At: 19:51

Publisher: Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



## Journal of Asian Natural Products Research

Publication details, including instructions for authors and subscription information:

<http://www.tandfonline.com/loi/ganp20>

### Two new compounds from *Ganoderma lucidum*

Xin-Fang Wang<sup>a</sup>, Yong-Ming Yan<sup>bc</sup>, Xin-Long Wang<sup>b</sup>, Xiu-Jing Ma<sup>d</sup>,  
Xue-Yan Fu<sup>a</sup> & Yong-Xian Cheng<sup>b</sup>

<sup>a</sup> College of Pharmacy, Ningxia Medical University, Yinchuan 750004, China

<sup>b</sup> State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650201, China

<sup>c</sup> Graduate School of Chinese Academy of Sciences, Beijing 100039, China

<sup>d</sup> Center for Drug Evaluation, State Food and Drug Administration, Beijing 100038, China

Published online: 08 Oct 2014.



[Click for updates](#)

To cite this article: Xin-Fang Wang, Yong-Ming Yan, Xin-Long Wang, Xiu-Jing Ma, Xue-Yan Fu & Yong-Xian Cheng (2015) Two new compounds from *Ganoderma lucidum*, *Journal of Asian Natural Products Research*, 17:4, 329-332, DOI: [10.1080/10286020.2014.960858](https://doi.org/10.1080/10286020.2014.960858)

To link to this article: <http://dx.doi.org/10.1080/10286020.2014.960858>

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms & Conditions of access and use can be found at <http://www.tandfonline.com/page/terms-and-conditions>

## Two new compounds from *Ganoderma lucidum*

Xin-Fang Wang<sup>a1</sup>, Yong-Ming Yan<sup>b,c1</sup>, Xin-Long Wang<sup>b</sup>, Xiu-Jing Ma<sup>d,\*</sup>, Xue-Yan Fu<sup>a,\*</sup>  
and Yong-Xian Cheng<sup>b,\*</sup>

<sup>a</sup>College of Pharmacy, Ningxia Medical University, Yinchuan 750004, China; <sup>b</sup>State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650201, China; <sup>c</sup>Graduate School of Chinese Academy of Sciences, Beijing 100039, China; <sup>d</sup>Center for Drug Evaluation, State Food and Drug Administration, Beijing 100038, China

(Received 25 August 2014; final version received 29 August 2014)

Two pairs of new enantiomers, lucidulactones A and B (**1** and **2**), and two known compounds were isolated from *Ganoderma lucidum*. Their structures were determined by means of spectroscopic methods. The chiral HPLC was used to separate the (–)– and (+)–antipodes of the new compounds.

**Keywords:** *Ganoderma lucidum*; lactones; enantiomers

### 1. Introduction

*Ganoderma lucidum* (Lingzhi), belonging to the genus *Ganoderma* which includes about 80 species, is a well-known traditional medicine used for centuries in the Orient for the treatment of various diseases including cancer, asthma, and sleep disorders [1]. In recent years, hundreds of compounds have been identified from this genus, and many of them are triterpenoids and polysaccharides [2–4]. Recently, we have identified a meroterpenoid from the fruiting bodies of *G. lucidum* which exhibited significant inhibitory effect on the phosphorylation of Smad3 [5]. Encouraged by this discovery, we undertook an in-depth study on this species which resulted in the isolation of four compounds including two new lactones (Figure 1). Herein, we describe their isolation and structure characterization.

### 2. Results and discussion

Compound **1** had the molecular formula C<sub>12</sub>H<sub>14</sub>O<sub>6</sub> derived from its HR-EI-MS at

*m/z* 254.0782 [M]<sup>+</sup>, <sup>13</sup>C NMR and DEPT spectra, indicating six degrees of unsaturation. The <sup>13</sup>C NMR and DEPT spectra (Table 1) showed 12 carbons attributed to two methoxyl, one oxygenated methylene, four methine including two olefinic and one oxygenated, four olefinic quaternary carbons, and one carbonyl carbon. In addition to a benzene ring and carbonyl, there should be one ring in the structure. The COSY spectrum showed cross peaks of H-3/H-4/H-5. The architecture of **1** was assembled by HMBC correlations, which were H-2', H-6'/C-4, OCH<sub>3</sub>/C-3', 5', H-5'/C-2, C-3, C-4, C-1', H-3/C-2, C-4, C-5, and C-1' (Figure 2). C-2 and C-5 are linked via an ether bond evidenced from HMBC correlations of H-5/C-2 (δ<sub>C</sub> 176.7). The relative configuration was determined by ROESY correlations of H-3/H-2', H-6' (Figure 3), and the large coupling constant of H-3 (*J* = 11.1 Hz), characteristic of a trans relationship of H-3 with H-4. The structure of **1** was therefore assigned as shown, named lucidulactone A. The opti-

\*Corresponding authors. Email: [yxcheng@mail.kib.ac.cn](mailto:yxcheng@mail.kib.ac.cn); [fuxueyan1215@163.com](mailto:fuxueyan1215@163.com); [maxiujing@sina.com](mailto:maxiujing@sina.com)

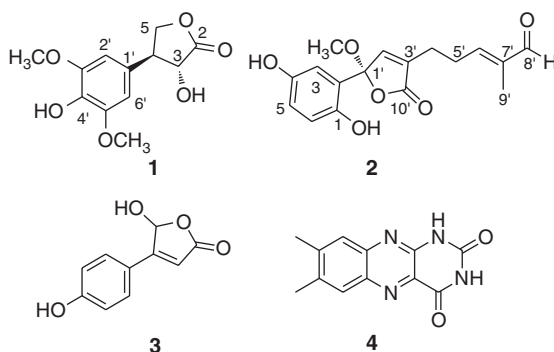


Figure 1. The structures of compounds 1–4.

cal rotation of **1** indicated a racemic nature, which was subsequently separated by HPLC on a chiral phase to afford two enantiomers, (+)-**1** and (–)-**1**.

The molecular formula of compound **2** was established as  $C_{17}H_{18}O_6$  from its HR-EI-MS at  $m/z$  318.1109  $[M]^+$ ,  $^{13}C$  NMR, and DEPT spectra, indicative of nine degrees of unsaturation. The  $^1H$  and  $^{13}C$  NMR spectral data of **2** (Table 1) were similar to those of fornicin A [4]. However, compared to fornicin A, in addition to the presence of a methoxyl

group in **2**, an oxygenated methine and a methyl in fornicin A were, respectively, replaced by an oxygenated quaternary carbon ( $\delta_C$  107.7) and an aldehyde ( $\delta_C$  195.2) in **2**. HMBC (Figure 2) correlation of  $OCH_3/C-1'$  suggested that the methoxyl group was located at C-1'. The  $^1H-^1H$  COSY correlations of H-4'/H-5'/H-6' and the HMBC correlations of H-8'/C-6', C-7', C-9' indicated the presence of a terminal aldehyde. The ROESY correlation of H-6'/H-8' (Figure 3) suggested that the geometry of double bond is trans. The optical

Table 1.  $^1H$  and  $^{13}C$  NMR spectral data for compounds **1** and **2** (600 MHz for  $^1H$  and 150 MHz for  $^{13}C$ , **1** in  $DMSO-d_6$  and **2** in  $acetone-d_6$ ).

No.	<b>1</b>		No.	<b>2</b>	
	$\delta_H, J$ (Hz)	$\delta_C$		$\delta_H, J$ (Hz)	$\delta_C$
2		176.7 s	1		151.1 s
3	4.57 dd (11.1, 7.0)	72.4 d	2		123.4 s
4	3.37–3.40 m	49.3 d	3	6.95 d (2.8)	114.3 d
5a	4.45–4.47 m	69.0 t	4		148.7 s
5b	4.14 dd (11.1, 8.8)		5	6.73 dd (8.6, 2.8)	118.0 d
1'		127.2 s	6	6.76 d (8.6)	118.3 d
2', 6'	6.69 s	105.3 d	1'		107.7 s
3', 5'		148.0 s	2'	7.48 s	147.4 d
4'		134.8 s	3'		135.2 s
3-OH	6.07 d (7.0)		4'	2.53–2.56 m	24.4 t
4'-OH	8.32 s		5'	2.70–2.73 m	27.2 t
3',5'- $OCH_3$	3.74 s	56.0 q	6'	6.61 t (7.2)	153.0 d
			7'		140.8 s
			8'	9.38 s	195.2 d
			9'	1.67 s	9.2 q
			10'		171.4 s
			$OCH_3$	3.26 s	51.8 q

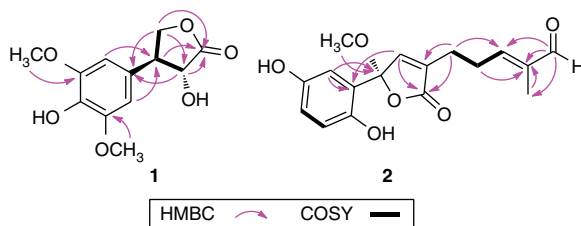


Figure 2. The COSY and HMBC correlations of **1** and **2**.

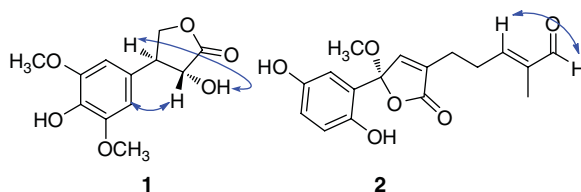


Figure 3. Key ROESY correlations of **1** and **2**.

rotation of **2** indicated a racemic nature, which was subsequently separated by HPLC on a chiral phase to afford two enantiomers, (+)-**2** and (–)-**2**, named lucidulactone B.

The known compounds were identified as hydroxybutenolide (**3**) [6] and lumi-chrome (**4**) [7], respectively, by comparison of their spectroscopic data with literature data.

### 3. Experimental

#### 3.1 General experimental procedures

Optical rotations were recorded on a Horiba SEPA-300 polarimeter (Horiba, Kyoto, Japan). UV spectra were recorded on a Shimadzu UV-2401PC spectrometer (Shimadzu, Kyoto, Japan). NMR spectra were recorded on a Bruker AV-600 spectrometer (Bruker, Karlsruhe, Germany), with TMS as an internal standard. EI-MS and HR-EI-MS were collected by AutoSpec Premier P776 spectrometer (Waters, Milford, MA, USA). ESI-MS were collected by API QSTAR Pulsar 1 spectrometer (Applied Biosystems/MDS Sciex, Foster City, CA, USA). Semi-preparative HPLC was carried out using an Agilent 1200 liquid chromatograph (Agilent, Santa Clara, CA, USA), the

column used was a 250 mm × 9.4 mm, i.d., 5 μm, Zorbax SB-C<sub>18</sub> and a 250 mm × 4.6 mm, i.d., 5 μm, Daicel Chiralpak IC. Column chromatography was performed on silica gel (200–300 mesh; Qingdao Marine Chemical, Inc., Qingdao, China), on C-18 silica gel (40–60 μm; Daiso Co., Osaka, Japan), MCI gel CHP 20P (75–150 μm, Mitsubishi Chemical Industries, Tokyo, Japan), and on Sephadex LH-20 (Amersham Pharmacia, Uppsala, Biosciences, Sweden).

#### 3.2 Plant material

*G. lucidum* was purchased from the Culture Base of Bei-Zhi-Tang Co., Ltd in Jilin Province, China, in July 2012. A voucher specimen (CHYX-0579) has been deposited at the State Key Laboratory of Photochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences.

#### 3.3 Extraction and isolation

The dried and powdered *G. lucidum* (80 kg) was extracted with 95% EtOH under reflux (2 × 360 l × 2 h) to give a crude extract, which was suspended in water followed by extracting with EtOAc

to afford a EtOAc-soluble fraction (1.1 kg). The EtOAc extract was divided into seven parts Fr. 1–Fr. 7 by a silica gel column eluted with a gradient of  $\text{CHCl}_3/\text{MeOH}$  (100:1 to 1:1). Fr. 3 (51 g) was fractionated by an MCI gel CHP 20P column eluting with gradient aqueous  $\text{MeOH}$  (20:80 to 100:0) to provide 11 portions Fr. 3.1–Fr. 3.11. Fr. 3.2 (8.4 g) was separated by Sephadex LH-20 ( $\text{MeOH}$ ) followed by an RP-18 column ( $\text{MeOH}/\text{H}_2\text{O}$ , 40:60 to 50:50), and semi-preparative HPLC ( $\text{MeOH}/\text{H}_2\text{O}$ , 25:75) to give compounds **1** ( $R_t = 16.5$  min, 5.5 mg) and **3** ( $R_t = 19.4$  min, 5 mg). Fr. 3.3 (31 mg) was purified by semi-preparative HPLC ( $\text{MeOH}/\text{H}_2\text{O}$ , 38:62) to give compounds **2** ( $R_t = 14.2$  min, 2 mg) and **4** ( $R_t = 20.8$  min, 10 mg). Compound **1** is a racemic mixture which was further purified by semi-preparative HPLC on a chiral phase (*n*-hexane/ethanol, 65:35, flow rate: 1 ml/min) to yield compounds (+)-**1** ( $R_t = 10.5$  min, 1.3 mg) and (–)-**1** ( $R_t = 13.6$  min, 1.5 mg). In the same manner as that of **1**, compound **2** was further purified via HPLC on a chiral phase (*n*-hexane/ethanol, 75:25, flow rate: 1 ml/min) to yield (+)-**2** ( $R_t = 13.8$  min, 0.5 mg) and (–)-**2** ( $R_t = 15.6$  min, 0.4 mg).

### 3.3.1 Lucidulactone A (**1**)

Yellowish gum;  $\{[\alpha]_D^{22} + 67.2$  ( $c = 0.14$ ,  $\text{MeOH}$ ), (+)-lucidulactone **A**};  $\{[\alpha]_D^{23} - 82.0$  ( $c = 0.15$ ,  $\text{MeOH}$ ), (–)-lucidulactone **A**}. UV ( $\text{MeOH}$ )  $\lambda_{\text{max}}$  ( $\log \epsilon$ ) 272 (3.15), 206 (4.59) nm. For  $^1\text{H}$  (600 MHz) and  $^{13}\text{C}$  NMR (150 MHz) spectral data, see Table 1. ESI-MS:  $m/z$  253  $[\text{M}-\text{H}]^-$ . HR-EI-MS:  $m/z$  254.0782  $[\text{M}]^+$  (calcd for  $\text{C}_{12}\text{H}_{14}\text{O}_6$ , 254.0790).

### 3.3.2 Lucidulactone B (**2**)

Yellowish gum;  $\{[\alpha]_D^{22} + 17.7$  ( $c = 0.04$ ,  $\text{MeOH}$ ), (+)-lucidulactone **B**};  $\{[\alpha]_D^{23} - 19.1$  ( $c = 0.05$ ,  $\text{MeOH}$ ), (–)-lucidulactone **B**}. UV ( $\text{MeOH}$ )  $\lambda_{\text{max}}$  ( $\log \epsilon$ ) 310 (3.43), 226 (4.31), 201 (4.19) nm. For  $^1\text{H}$  (600 MHz) and  $^{13}\text{C}$  NMR (150 MHz) spectral data, see Table 1. EI-MS:  $m/z$  318  $[\text{M}]^+$ . HR-EI-MS:  $m/z$  318.1109  $[\text{M}]^+$  (calcd for  $\text{C}_{17}\text{H}_{18}\text{O}_6$ , 318.1103).

### Acknowledgements

This work was financially supported by the grants of NSFC-Joint Foundation of Yunnan Province (U1202222), National Natural Science Foundation of China (21472199), and State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany (P2013-ZZ03).

### Note

1. These authors contributed equally to this paper.

### References

- [1] J. Kenneth, *Reishi: Ancient Herb for Modern Times* (Sylvan Press, Issaquah, 1990).
- [2] M.S. Shiao, *Chem. Rec.* **3**, 172 (2003).
- [3] S. Fatmawati, R. Kondo, and K. Shimizu, *Bioorg. Med. Chem. Lett.* **23**, 5900 (2013).
- [4] X.M. Niu, S.H. Li, H.D. Sun, and C.T. Che, *J. Nat. Prod.* **69**, 1364 (2006).
- [5] Y.M. Yan, J. Ai, L.L. Zhou, A.C.K. Chung, R. Li, J. Nie, P. Fang, X.L. Wang, J. Luo, Q. Hu, F.F. Hou, and Y.X. Cheng, *Org. Lett.* **15**, 5488 (2013).
- [6] L.Y. Li, Z.W. Deng, H.Z. Fu, J. Li, P. Proksch, and W.H. Lin, *Pharmazie* **58**, 680 (2003).
- [7] Z.G. Ding, J.Y. Zhao, P.W. Yang, M.G. Li, R. Huang, X.L. Cui, and M.L. Wen, *Magn. Reson. Chem.* **47**, 366 (2009).