This article was downloaded by: [Kunming Institute of Botany]

On: 18 May 2015, At: 19:38 Publisher: Taylor & Francis

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered

office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK





Journal of Asian Natural Products Research

Publication details, including instructions for authors and subscription information:

http://www.tandfonline.com/loi/ganp20

Two new phenol derivatives from Stereum hirsutum FP-91666

Yuan-Chang Duan^{ab}, Xin-Xin Meng^c, Yan-Long Yang^{ab}, Yin-He Yang^a & Pei-Ji Zhao^a

^a State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650201, China

^b University of Chinese Academy of Sciences, Beijing 100049, China

^c Faculty of Life Science and Technology, Kunming University of Science and Technology, Kunming 650500, China Published online: 08 Oct 2014.

To cite this article: Yuan-Chang Duan, Xin-Xin Meng, Yan-Long Yang, Yin-He Yang & Pei-Ji Zhao (2015) Two new phenol derivatives from Stereum hirsutum FP-91666, Journal of Asian Natural Products Research, 17:4, 324-328, DOI: 10.1080/10286020.2014.959439

To link to this article: http://dx.doi.org/10.1080/10286020.2014.959439

PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms &

Conditions of access and use can be found at http://www.tandfonline.com/page/terms-and-conditions



Two new phenol derivatives from Stereum hirsutum FP-91666

Yuan-Chang Duan^{ab}, Xin-Xin Meng^c, Yan-Long Yang^{ab}, Yin-He Yang^a and Pei-Ji Zhao^a*

^aState Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650201, China;
^bUniversity of Chinese Academy of Sciences, Beijing 100049, China;
^cFaculty of Life Science and Technology, Kunming University of Science and Technology, Kunming 650500, China

(Received 28 May 2014; final version received 25 August 2014)

Two new phenol derivatives, 2-(3-methyl-2-buten-1-yl)-4-methoxyethyl-phenol (1) and 5-hydroxy-4-(hydroxymethyl)-2-(3-methylbut-2-en-1-yl)cyclohex-4-en-1-one (2), together with eight known compounds consisting of phenol derivatives (3 and 4), niacinamide (5), and five ergosta type compounds (6–10), were isolated from solid fermentation products of *Stereum hirsutum* FP-91666. Two new structures were elucidated by extensive spectroscopic methods, including 1D NMR and 2D NMR, and HR-EI-MS experiments.

Keywords: Stereaceae; *Stereum hirsutum* FP-91666; 2-(3-methyl-2-buten-1-yl)-4-methoxyethyl-phenol; 5-hydroxy-4-(hydroxymethyl)-2-(3-methylbut-2-en-1-yl)cyclohex-4-en-1-one

1. Introduction

The fungus of Stereum belongs to the Stereaceae family (basidiomycetes), and can produce diversiform secondary metabolites [1-3], especially produce structurally diverse sesquiterpenes [4]. In process of studying on the biosynthesis of vibralactone [5], which was produced by Stereum vibrans [6], we had searched analogous genes of biosynthetic vibralactone and found that Stereum hirsutum (S. ostrea) can produce vibaralactone type compounds [7,8]. According to genome data analysis, S. hirsutum would produce more secondary metabolites [9], so we investigated the chemical constituent of S. hirsutum FP-91666. And two new phenol derivatives, 2-(3-methyl-2-buten-1-yl)-4-methoxyethyl-phenol (1) and 5hydroxy-4-(hydroxymethyl)-2-(3-methylbut-2-en-1-yl)cyclohex-4-en-1-one (2), together with eight known compounds were isolated from solid fermentation

products of *S. hirsutum* FP-91666. This report describes the isolation and structural elucidation of these compounds.

2. Results and discussion

Compound 1 was obtained as colorless amorphism. The HR-EI-MS data indicated a molecular formula of C₁₄H₂₀O₂ based on the $[M]^+$ ion signal at m/z 220.1456. The ¹³C NMR data (Table 1) revealed four quaternary carbons at $\delta_{\rm C}$ 153.8, 135.6, 134.9, and 126.7, five methines at $\delta_{\rm C}$ 127.9, 125.5, 121.8, 115.6, and 79.2, and one methylene ($\delta_{\rm C}$ 30.1) and four methyls, and from ¹H NMR spectrum, three aromatic protons ($\delta_{\rm H}$ 7.06 (brs), 7.07 (dd, 8.0, 2.1), and 6.81 (d, 8.0)) indicated the presence of three-substituted benzene ring; and two methyl singlets ($\delta_{\rm H}$ 1.80 (s) and 1.81 (s)) and one sp² proton ($\delta_{\rm H}$ 5.33–5.36 (m)) suggested compound 1 was prenylated phenol derivatives (Figure 1) [10].

^{*}Corresponding author. Email: zhaopeiji@mail.kib.ac.cn

Position	1 ^a		2 ^b	
	¹ H	¹³ C	¹ H	¹³ C
1	_	153.8, s	_	212.7, s
2	_	126.7, s	2.38-2.44 (m)	46.5, d
3	7.06 (brs)	127.9, d	2.78-2.82 (m); 2.36 (d, 16)	34.5, t
4		135.6, s	-	138.4, s
5	7.07 (dd, 8.0, 2.1)	125.5, d	_	177.8, s
6	6.81 (d, 8.0)	115.6, d	4.24 (s)	54.0, t
7	3.39 (d, 7.2)	30.1, t	2.38-2.44 (m); 2.10-2.16 (m)	30.3, t
8	5.33-5.36 (m)	121.8, d	5.06-5.09 (m)	121.8, d
9	_ ` ´	134.9, s	_ ` ′	134.9, s
10	1.80 (s)	25.8, q	1.68 (s)	25.9, q
11	1.81 (s)	17.9, q	1.63 (s)	17.9, q
12	4.24 (q, 6.4)	79.2, d	4.56–4.60 (m)	61.3, t
13	1.44 (d, 6.4)	23.8, q		
12-OCH ₃	3.22 (s)	56.2, q		
- OH	5.12 (s)			

Table 1. ¹H and ¹³C NMR spectral data of compounds 1 and 2.

The HMBC experiment (Table 1) showed correlations between the proton at $\delta_{\rm H}$ 4.24 (H-12) and carbons at $\delta_{\rm C}$ 125.5 (C-5), 127.9 (C-3), 23.8 (C-13), 56.2 (12-OCH₃), and 135.6 (C-4), between the proton at $\delta_{\rm H}$ 1.44 (H-13) and the carbons at $\delta_{\rm C}$ 79.2 (C-12) and 135.6 (C-4), which indicated that one methoxyethyl group connected with prenylated phenol at position C-4 ($\delta_{\rm C}$ 135.6) (Figure 2). C-12 was a chiral center, but its configuration cannot be determinated by current data. Together with other

data (Table 1), compound **1** was elucidated to be 2-(3-methyl-2-buten-1-yl)-4-methoxyethyl-phenol (Figure 1).

Compound **2** was obtained as colorless amorphism. The HR-EI-MS data indicated a molecular formula of $C_{12}H_{18}O_3$ based on the [M]⁺ ion signal at m/z 210.1250. The ¹³C NMR data (Table 1) revealed four quaternary carbons at δ_C 212.7, 138.4, 177.8, and 134.9, two methines at δ_C 46.5 and 121.8, four methylenes (δ_C 54.0, 34.5, 30.3, and 61.3), and two methyls, which

Figure 1. The compound structures from S. hirsutum FP-91666.

^a The NMR data are obtained on Avance III 600 in CDCl₃.

^b The NMR data are obtained on Bruker AV-800 in CD₃OD.

Figure 2. Selected HMBC correlations of new compounds 1 and 2.

indicated that compound 2 was also prenylated phenol derivatives except the phenol was oxidized [10]. The detail structure was confirmed by 2D NMR. The HMBC experiment (Table 1) showed the proton at $\delta_{\rm H}$ 5.06–5.09 (H-8) correlated with carbons at $\delta_{\rm C}$ 17.9 (C-11), 25.9 (C-10), 30.3 (C-7), and the protons of methylene at $\delta_{\rm H}$ 2.38–2.44 and 2.10–2.16 (H-7) with carbons at $\delta_{\rm C}$ 121.8 (C-8) and 134.9 (C-9), and two methyl group protons at $\delta_{\rm H}$ 1.68 (H-10) and 1.63 (H-11) with carbons at $\delta_{\rm C}$ 121.8 (C-8) and 134.9 (C-9), which indicated the existence of an isopentenyl group. H-12 at $\delta_{\rm H}$ 4.60 correlated with C-4, C-5, C-3, and H-2 correlated with C-1 and C-3, H-6 correlated with the C-1, C-4, and C-5, and H-3 correlated with the C-1, C-2, C-4 and C-5, which indicated compound 2 contained 5-hydroxy-4-(hydroxymethyl)cyclohex-4en-1-one, which would come from oxidized phenol. These two units connected by C-C bond based on the correlations between protons of methylene at $\delta_{\rm H}$ 2.38– 2.44 and 2.10-2.16 (H-7) and C-2 (Figure 2). The configuration of C-2 is uncertain because H-2 was overlapped with H-7, and there is no NOE correlation between H-3 and H-7. Based on above data, compound 2 was determined to be 5-hydroxy-4-(hydroxymethyl)-2-(3-methylbut-2-en-1yl)cyclohex-4-en-1-one (Figure 1).

In addition to the above-mentioned new phenol derivatives, eight known compounds including 3-(hydroxymethyl)-4-methoxy-2-(3-methylbut-3-en-1-yn-1-yl)phenol (3) [2], 4-methoxy-benzoic acid

methyl ester (4) [11], niacinamide (5) [11], ergosterol peroxid (6) [11], (22E,24R)-ergosta-5,7,22-trien-3 β -ol (7) [12], 3β ,5 α ,6 β -triol-ergosta-7,22-diene (8) [13], 3β ,5 α -diol-6 β -methoxy-ergosta-7,22-diene (9) [14], and 3β ,5 α ,9 α -trihy-droxy-ergosta-7,22-dien-6-one (10) [15] were also isolated and identified by comparing their NMR and MS data with those reported in the literatures.

3. Experimental

3.1 General experimental procedures

Optical rotations were measured on a Jasco DIP-370 digital polarimeter (JASCO, Tokyo, Japan). UV spectra were measured on a Shimadzu UV-2401PC spectrophotometer (Shimadzu, Tokyo, Japan), λ_{max} (log ϵ) in nm. IR spectra were scanned with Bruker Tensor-27 infrared spectrometer with a KBr disk (Bruker, Karlsruhe, Germany). NMR experiments were carried out on Bruker AM-400, Bruker DRX-500 NMR, Avance III 600, and Bruker AV-800 spectrometers (Bruker) with TMS as internal standard. ESI-MS and HR-EI-MS were recorded on a Finnigan LCQ Advantage mass spectrometer (Thermo, San Jose, CA, USA) and a VG Auto-Spec-3000 mass spectrometer (VG, Manchester, England). Column chromatography was carried out on silica gel (G, 200-300 mesh and H, Qingdao Marine Chemical Factory, Qingdao, China) and Sephadex LH-20 (Pharmacia Fine Chemicals, Uppsala, Sweden). Precoated silica gel GF254 plates (Qingdao Marine Chemical Factory) were used for thin-layer chromatography.

3.2 Fungal material

S. hirsutum FP-91666, stored in glycerol at -80° C in State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, was used to inoculate on YMG (yeast extract 4.0 g/l, malt extract 10.0 g/l,

glucose 4.0 g/l, agar 15.0 g/l, pH 7.3 before sterilization) media in plate at 26°C for 14 days to afford seed cultures. The *S. hirsutum* (151) was cultured on YMG dish at temperature of 26°C for 14 days.

3.3 Extraction and isolation

Solid fermentation products of S. hirsutum (151) was cut into small pieces and extracted exhaustively with mixture solution (EtOAc/MeOH/HAc, 80:15:5, v/v/v) by three times to obtain the rude extract. The extracts were dissolved in water. and extracted with EtOAc, and then with *n*-butanol three times, respectively. The extract of EtOAc section residue (6.85 g) was subjected to a column of silica gel G (200–300 mesh) using petroleum ether (PE)-EtOAc (10:1 \rightarrow 6:4) and CHCl₃-MeOH (10:1 \rightarrow 0:100) gradient solvent system to give 10 fractions (Fr. 1–Fr. 10). The fraction Fr. 1 (1.10 g) was placed in a column of silica gel (200-300 mesh) and eluted with PE-EtOAc (100:1 \rightarrow 7:3) gradient solvent system to yield fractions of Fr. 1.1-Fr. 1.6. Fr. 1.2 (185 mg) was purified by silica gel (GF254) column using PE-acetone and Sephadex LH-20 (acetone) to produce compound 4 (56 mg). The fraction Fr. 3 (2.23 g) was subjected to a silica gel column (200–300 mesh) using PE-acetone (100:2 \rightarrow 7:3) gradient solvent system to give fractions of Fr. 3.1–Fr. 3.6. Fr. 3.2 (399 mg) was purified with Sephadex LH-20 eluted with acetone to give compound 7 (33 mg). Fr. 3.6 (125 mg) was placed on a column of silica gel H using PE-acetone (20:1) isocratic solvent system to get compound 6 (2 mg). The fraction Fr. 4 (0.257 g) was subjected to a column of silica gel G (200-300 mesh) using PE-acetone (100:1 \rightarrow 6:4) gradient solvent system to give fractions of Fr. 4.1 – Fr. 4.7. Subsequently Fr. 4.3 (130 mg) was purified by silica gel column (200-300 mesh) with PE-EtOAc (25:1) isocratic solvent system to obtain compound 1 (3 mg). The Fr. 6 (0.32 g) was subjected to a column of silica gel (200-300 mesh) using PE-acetone (10:1 \rightarrow 6:4) gradient solvent system to yield fractions of Fr. 6.1-Fr. 6.3. Fr. 6.3 (11 mg) was purified by Sephadex LH-20 (CHCl₃-MeOH; 1:1) and a column of silica gel (GF254) eluted by PE-acetone (20:1) gradient isocratic solvent system, to give compound 9 $(2.8 \,\mathrm{mg})$. The fraction Fr. 7 $(0.56 \,\mathrm{g})$ was placed to a column of silica gel (200-300 mesh) using PE-EtOAc (9:1 \rightarrow 6:4) gradient solvent system to provide fractions of Fr. 7.1-Fr. 7.4. Fr. 7.2 (19 mg) was first subjected to Sephadex LH-20 (CHCl₃-MeOH; 1:1), and then purified by semipreparative HPLC (LC3000 Semi-preparation Gradient HPLC System, Beijing, China) to yield compounds 2 (0.8 mg) and 3 (4 mg) (sample was performed in a RP- C_{18} column (250 mm × 10 mm) at ambient temperature with a detection wavelength at 254 nm and a mobile phase of methanol/ water (the water reduces from 50% to 0%) at a flow rate of 3 ml/min). Fr. 7.3 (22 mg) was purified by Sephadex LH-20 (CHCl₃-MeOH; 1:1), a column of silica gel (GF254) using PE-acetone (100:7) isocratic solvent system and Sephadex LH-20 (acetone) to provide compound 8 (2 mg). The fraction Fr. 8 (0.55 g) was placed on a column of silica gel (200-300 mesh) using PE-acetone (9:1 \rightarrow 6:4) gradient solvent system to pick up fractions of Fr. 8.1-Fr. 8.6. Then Fr. 8.3 (21 mg) was purified by a column of silica gel H using (CHCl₃-MeOH; 30:1) isocratic solvent system to produce compound 10 (9 mg). Fr. 8.5 (10 mg) was subjected to Sephadex LH-20 CHCl₃-MeOH (1:1) and a column of silica gel (GF254) with CHCl₃-MeOH (30:1) isocratic solvent system to produce compound 5 (5 mg).

3.3.1 2-(3-Methyl-2-buten-1-yl)-4-methoxyethyl-phenol (1)

Colorless amorphism, $[\alpha]_D^{22} - 12.9$ (c = 0.70, CHCl₃); UV (MeOH) λ_{max} (log ϵ): 277 (3.17), 239 (3.04), 209

(2.85), 201 (2.85) nm; IR (KBr) $\nu_{\rm max}$: 3440, 2923, 1621, 1111, 1036 cm⁻¹; ¹H and ¹³C NMR spectral data see Table 1; EI-MS: m/z 220 [M]⁺; HR-EI-MS: m/z 220.1456 [M]⁺ (calcd for $C_{14}H_{20}O_2$, 220.1463).

3.3.2 3-Hydroxy-4-(hydroxymethyl)-6-(3-methylbut-2-en-1-yl)cyclohex-3-en-1-one (2)

Colorless amorphism, $[\alpha]_D^{22} - 6.6$ (c = 0.70, MeOH); UV (MeOH) λ_{max} (log ϵ): 202 (2.54) nm; 1 H and 13 C NMR spectral data see Table 1; EI-MS: m/z 210 [M] $^+$; HR-EI-MS: m/z 210.1250 [M] $^+$ (calcd for $C_{12}H_{18}O_3$, 210.1256).

Acknowledgments

The strain S. hirsutum FP-91666 was obtained as a gift from culture collection manager Ms Rita Rentmeester of Center for Forest Mycology Research, Northern Research Station, Madison, WI, USA. This work was supported by grants from the "973" Program of China (2013CB127505), the NSFC (31170061), and Applied Basic Research Foundation of Yunnan Province (2013FA018), the Young and Technical Leader Raising Foundation of Yunnan Province (2009CI071). We acknowledge the Department of Instrumental Analysis of Kunming Institute of Botany for measuring the optical rotations, UV, NMR, and mass spectra.

References

- [1] M.S.R. Nair and M. Anchel, *Phytochemistry* **16**, 390 (1977).
- [2] G.M. Dubin, A. Fkyerat, and R. Tabacchi, Phytochemistry 53, 571 (2000).
- [3] J.O. Omolo, H. Anke, and O. Sterner, *Phytochemistry* **60**, 431 (2002).
- [4] W.R. Abraham, Curr. Med. Chem. 8, 583 (2001).
- [5] P.J. Zhao, Y.L. Yang, L. Du, J.K. Liu, and Y. Zeng, *Angew. Chem. Int. Ed.* **52**, 2298 (2013).
- [6] D.Z. Liu, F. Wang, T.G. Liao, J.G. Tang, W. Steglich, H.J. Zhu, and J.K. Liu, *Org. Lett.* 8, 5749 (2006).
- [7] Kim J.P., Repub. Korean Kongkae Taeho Kongbo, KR 2010063970 A 20100614 (2010).
- [8] J.P. Kim, H.S. Kang, and S.H. Park, Repub. Korean Kongkae Taeho Kongbo, KR 2009089203 A 20090821 (2009).
- [9] G. Lackner, M. Misiek, J. Braesel, and D. Hoffmeister, *Fungal Genet. Biol.* 49, 996 (2012).
- [10] J. Yu, H.H. Zhang, Q. Yu, and L.J. Xuan, Helv. Chim. Acta 92, 1880 (2009).
- [11] K.G. Desai and K.R. Desai, *Molbank* **2006**, M477 (2006).
- [12] C. Xiang, J.K. Liu, and Z.W. Du, Anhui Med. Pharm. J. 12, 007 (2011).
- [13] J. Zhao, Y. Mou, T. Shan, Y. Li, L. Zhou, M. Wang, and J. Wang, *Molecules* 15, 7961 (2010).
- [14] I.S. Lee, W.Y. Jin, X. Zhang, T.M. Hung, K.S. Song, Y.H. Seong, and K. Bae, *Arch. Pharm. Res.* 29, 548 (2006).
- [15] H. Cai, X. Liu, Z. Chen, S. Liao, and Y. Zou, Food Chem. 141, 2873 (2013).