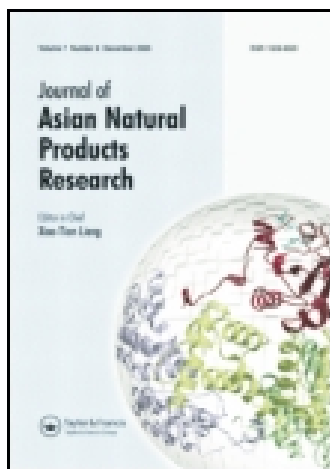


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### Lignans from the bark of *Zanthoxylum simulans*

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## Lignans from the bark of *Zanthoxylum simulans*

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Investigation on the EtOAc extract of the bark of *Zanthoxylum simulans* led to the isolation of four new lignans including zanthoxylumin A (**1**), zanthoxylumin B (**2**), (–)-magnolol (**3**), and (–)-pinoselinol-di-3,3-dimethylallyl ether (**4**). Their structures were established by comprehensive analysis of the spectral data, especially 1D and 2D NMR spectra.

**Keywords:** *Zanthoxylum simulans*; Rutaceae; lignans

### 1. Introduction

The bark of *Zanthoxylum simulans* (Rutaceae) has been widely used as a folk medicine in China mainly to treat rheumatoid arthritis and swelling [1]. Alkaloids, coumarins, and lignans are major constituents in this genus [2,3]. However, up to now, the specific components against rheumatoid arthritis in this plant species were not well clarified [4–7]. As a part of our study on medicinal plants, we investigated the bark of this plant, from which four new lignans including zanthoxylumin A (**1**), zanthoxylumin B (**2**), (–)-magnolol (**3**), and (–)-pinoselinol-di-3,3-dimethylallyl ether (**4**) were isolated (Figure 1). Herein, we describe the isolation and structural elucidation of these compounds.

### 2. Results and discussion

Compound **1** was obtained as a colorless oil, and had a molecular formula C<sub>18</sub>H<sub>22</sub>O<sub>5</sub> from the HR-EI-MS at *m/z* 318.1459 [M]<sup>+</sup>. Its IR spectrum revealed the existence of a five-membered lactone ring (1770 cm<sup>-1</sup>)

and benzene ring (1514 and 1464 cm<sup>-1</sup>). <sup>1</sup>H NMR data (Table 1) implied an ABX system at δ<sub>H</sub> 6.82 (dd, *J* = 8.2, 1.5 Hz), 6.85 (d, *J* = 8.2 Hz), and 6.88 (d, *J* = 1.5 Hz), indicative of the presence of a 1,3,4-trisubstituted benzene ring. In addition, <sup>13</sup>C NMR (Table 1) and DEPT spectra displayed 18 signals including one carbonyl carbon (δ<sub>C</sub> 178.2), one benzylic oxygenated methine carbon (δ<sub>C</sub> 86.0), two methine carbons (δ<sub>C</sub> 48.1 and 46.0, respectively), two oxygenated methylene carbons (δ<sub>C</sub> 70.0 and 69.8, respectively), three aromatic unsaturated quaternary carbons, and three aromatic unsaturated methine carbons, suggesting that **1** might be a lignan derivative with a five-membered lactone ring. Detailed analysis of the 1D and 2D NMR spectral data revealed that **1** was very similar to forsythenin [8], except for a methoxy moiety at the benzene ring being replaced by an oxygenated isopentenyl moiety. This oxygenated isopentenyl moiety can be confirmed by the <sup>1</sup>H NMR spectral data at δ<sub>H</sub> 4.58 (2H, d, *J* = 6.6 Hz), 5.50 (1H, t,

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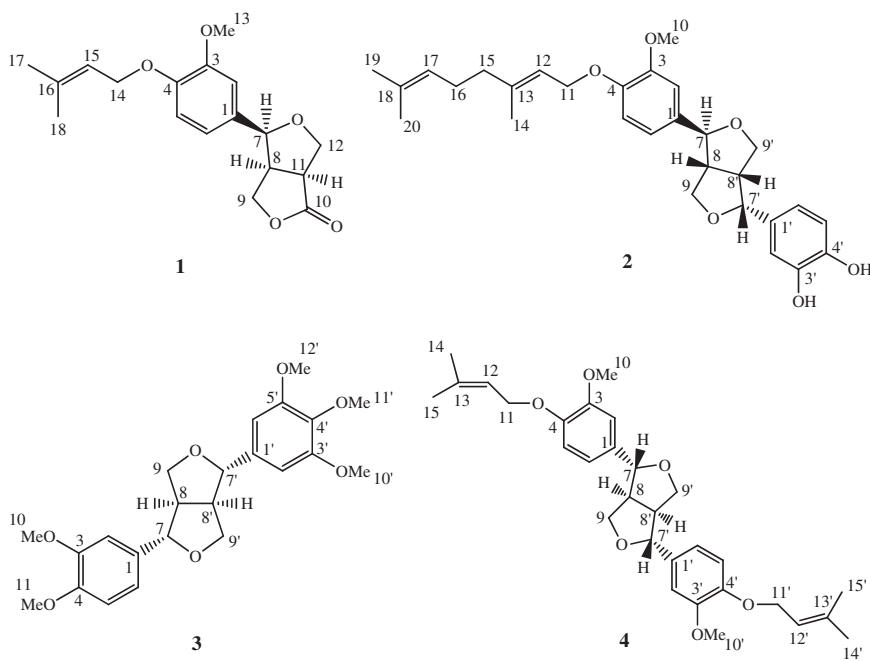


Figure 1. Chemical structures of compounds **1**–**4**.

$J = 6.6$  Hz), 1.77 (3H, s), 1.73 (3H, s). The key HMBC correlation between  $H_2$ -14 and C-4 ( $\delta_C$  148.4) indicated that the isopentyl moiety was connected to C-4 of the 1,3,4-trisubstituted benzene ring via an ether bond (Figure 2). The ROESY spectrum showed the correlations between H-8 and H-7/H-11/H-9a, H-11 and H-7/H-8/H-9a/H-12a, indicating these protons to be on the same side. Thus, the relative configuration of compound **1** was assigned as shown in Figure 2 and named as zanthoxylum A.

Compound **2** was isolated as a white solid, and had the molecular formula  $C_{29}H_{36}O_6$  as determined by HR-EI-MS at  $m/z$  480.2530  $[M]^+$ . IR and NMR spectra established that **2** possessed tetrahydrofuran lignan skeleton [9,10]. The  $^1H$  NMR data (Table 1) exhibited the presence of two ABX systems at  $\delta_H$  6.91 (1H, d,  $J = 8.0$  Hz), 6.89 (1H, dd,  $J = 8.0, 1.2$  Hz), 6.98 (1H, d,  $J = 1.2$  Hz) and  $\delta_H$  6.72 (1H, d,  $J = 8.0$  Hz), 6.65 (1H, dd,  $J = 8.0,$

1.4 Hz), 6.80 (1H, d,  $J = 1.4$  Hz), suggesting two 1,3,4-trisubstituted benzene rings. Meanwhile, signals for one geranyloxy group were observed in the NMR spectra of **2** (Table 1), which was further supported by COSY correlations of  $H_2$ -11/ $H_2$ -12,  $H_2$ -15/ $H_2$ -16/ $H_2$ -17 and by HMBC correlations of  $H_2$ -11 to C-12 and C-13, and  $H_2$ -15 to C-12, C-13, C-14, C-16, and C-17, and H-17 to C-19 and C-20. The key HMBC correlations between  $H_2$ -11 and C-4 indicated that the geranyloxy group was directly connected to C-4 of the 1,3,4-trisubstituted benzene ring (Figure 2). The general features of its NMR spectroscopic data (Table 1) were markedly similar to those of planispine B [11]. Detailed comparison of the NMR and MS data of these two compounds indicated that a methoxy group ( $\delta_C$  55.8,  $\delta_H$  3.84) in planispine B was replaced by a hydroxyl group in **2**. Previous studies have suggested that the essential elements of the furofuran lignan skeleton have regular stereo-

Table 1.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectral data of compounds 1–4.

No.	1		2		3		4	
	$\delta_{\text{H}}$ ( $J$ in Hz)	$\delta_{\text{C}}$	$\delta_{\text{H}}$ ( $J$ in Hz)	$\delta_{\text{C}}$	$\delta_{\text{H}}$ ( $J$ in Hz)	$\delta_{\text{C}}$	$\delta_{\text{H}}$ ( $J$ in Hz)	$\delta_{\text{C}}$
1		131.0		135.7		133.5		133.6
2	6.88 d (1.5)	109.1	6.98 d (1.2)	111.3	6.89 d (1.8)	109.2	6.90 br s	109.4
3		149.8		150.3		149.3		149.7
4		148.4		149.3		148.8		148.0
5	6.85 d (8.2)	112.7	6.91 d (8.0)	115.3	6.82 d (8.2)	111.1	6.85 br s	112.8
6	6.82 dd (8.2, 1.5)	118.3	6.89 dd (8.0, 1.2)	120.0	6.86 dd (8.2, 1.8)	118.4	6.85 br s	118.3
7	4.62 d (6.9)	86.0	4.42 d (7.0)	89.4	4.72 d (4.5)	85.9	4.75 d (4.2)	86.0
8	3.10–3.16 m	48.1	2.89–2.92 m	55.9	3.06–3.12 m	54.6	3.10–3.14 m	54.3
9a, b	4.50 dd (9.8, 6.8), 4.33 dd (9.8, 2.0)	69.8	4.08 d (9.3), 3.81–3.83 m	72.0	3.88–3.90 m, 4.24–4.28 m	72.1	3.88 dd (8.7, 3.6), 4.26 dd (8.7, 6.8)	71.9
10		178.2	3.84 s	56.6	3.88 s	56.1	3.88 s	56.1
11	3.45 ddd (9.1, 9.1, 3.8)	46.0	4.58 d (6.4)	67.1	3.86 s	56.1	4.57 d (6.7)	65.9
12	4.37 dd (9.1, 9.1), 4.19 dd (9.1, 3.8)	70.0	5.44–5.46 m	121.4			5.51 td (6.7)	120.0
13		55.9		142.2				137.9
14	3.88 s	65.7	1.72 s	16.8				26.1
15	4.58 d (6.6)	119.7	2.05–2.07 m	40.8			1.76 s	18.4
16	5.50 t (6.6)	137.9	2.10–2.14 m	27.5			1.72 s	
17	1.77 s	25.8	5.08–5.10 m	125.1				
18	1.73 s	18.2		132.7				
19			1.65 s	26.0				
20			1.60 s	17.9				
1'				131.1				133.6
2'			6.80 d (1.4)	114.2	6.55 s	102.8	6.90 br s	109.4
3'				147.1		153.6		149.7
4'				146.2		137.5		148.0
5'			6.72 d (8.0)	116.2		153.6	6.85 br s	112.8
6'			6.65 dd (8.0, 1.4)	117.9	6.55 s	102.8	6.85 br s	118.3
7'			4.79 d (5.3)	83.7	4.74 d (4.5)	86.2	4.75 d (4.2)	86.0
8'			3.32–3.35 m	51.4	3.06–3.12 m	54.2	3.10–3.14 m	54.3

9'a, b	3.30–3.32 m, 3.79–3.81 m	70.9	3.88–3.90 m, 4.24–4.28 m	71.9	3.88 dd (8.7, 3.6), 4.26 dd (8.7, 6.8)	71.9
10'			3.85 s	56.3	3.88 s	56.1
11'			3.81 s	61.1	4.57 d (6.7)	65.9
12'			3.85 s	56.3	5.51 td (6.7)	120.0
13'						137.9
14'					1.76 s	26.1
15'					1.72 s	18.4

chemistry, namely, the chemical shifts of C-7 and C-7' in their  $^{13}\text{C}$  NMR spectra have diagnostic values in the different types of furofurano lignans [3,7,11]. More specifically, the chemical shifts of C-7 and C-7' were 89.4 and 83.7 ppm, respectively, indicating that the relative configurations were determined as  $8S^*$ ,  $7R^*$ ,  $8'S^*$ , and  $7'S^*$ . The ROESY spectrum further showed the correlations between H-7 and H-2/H-9a/H-9'a, H-8 and H-8'/H-9'b, H-7' and H-9b/H-8'. Therefore, the relative configuration of compound **2** was elucidated as shown in Figure 2 and named as zanthoxylum B.

Compound **3** was obtained as a colorless oil. The HR-EI-MS ( $m/z$  416.1838  $[\text{M}]^+$ , calcd for 416.1835) suggested its molecular formula to be  $\text{C}_{23}\text{H}_{28}\text{O}_7$ .  $^1\text{H}$  and  $^{13}\text{C}$  NMR data (Table 1) showed the characteristic pattern of a lignan with tetrahydrofuran moieties. NMR data of **3** were nearly the same as those of the known lignan (+)-magnolol [12]. The only difference is in the optical rotation value, (+)-magnolol was positive  $[\alpha]_{\text{D}}^{20} + 53.4$  ( $c = 3.1$ ,  $\text{CHCl}_3$ ) [12], but **3** was negative  $[\alpha]_{\text{D}}^{23.5} - 15.3$  ( $c = 0.1$ ,  $\text{CHCl}_3$ ). Thus, they are enantiomers, and compound **3** was determined as (–)-magnolol.

Compound **4** was isolated as a white solid. Its molecular formula was established as  $\text{C}_{30}\text{H}_{38}\text{O}_6$  ( $m/z$  494.2673  $[\text{M}]^+$ , calcd for 494.2668) on the basis of its HR-EI-MS analyses, but in the  $^{13}\text{C}$  NMR spectrum, only 15 carbon signals were observed, which indicated the existence of a symmetrical structure in **4**.  $^1\text{H}$  and  $^{13}\text{C}$  NMR data (Table 1) also implied **4** was a lignan with tetrahydrofuran moieties as compound **3**. All NMR spectral data of **4** were extremely close to those of the known lignan (+)-pinoresinol-di-3,3-dimethylallyl ether [13]. The only difference is in the optical rotation value, (+)-pinoresinol-di-3,3-dimethylallyl ether was positive  $[\alpha]_{\text{D}}^{25} + 41.6$  ( $c = 0.13$ ,  $\text{CHCl}_3$ ) [13], but **4** was negative  $[\alpha]_{\text{D}}^{26.8} - 32.1$

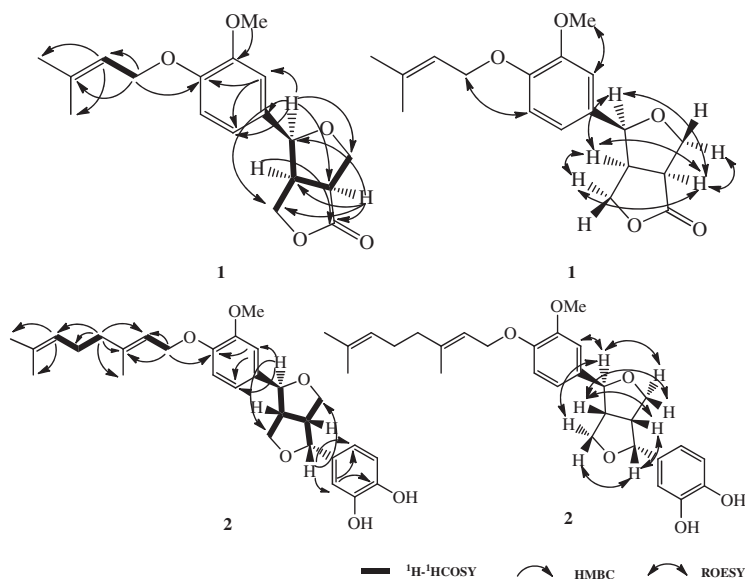


Figure 2. Key HMBC,  $^1\text{H}$ - $^1\text{H}$  COSY, and ROESY correlations of compounds **1** and **2**.

( $c = 0.3$ ,  $\text{CHCl}_3$ ). Therefore, they are also enantiomers, and compound **4** was identified as (–)-pinoresinol-di-3,3-dimethylallyl ether.

### 3. Experimental

#### 3.1 General experimental procedures

Optical rotations were recorded on a Horiba SEPA-300 polarimeter (Horiba, Tokyo, Japan). UV spectra were measured on a Shimadzu UV-2401PC spectrophotometer (Shimadzu Corporation, Kyoto, Japan). IR spectra were obtained on a Tensor 27 (Bruker Optics GmbH, Ettlingen, Germany) with KBr pellets. NMR spectra were recorded on a Bruker AV-400 or a DRX 500 or a Bruker Avance III-600 MHz spectrometer (Bruker BioSpin GmbH, Rheinstetten, Germany). EI-MS was obtained on a Finnigan-4510 spectrometer. ESI-MS and HR-EI-MS were determined with an API QSTAR Pulsar 1 spectrometer (MDS Sciex, Concord, ON, Canada). Silica gel (200–300 mesh, Qingdao Marine Chemical, Inc., Qingdao, China), RP-18 gel (40–63  $\mu\text{m}$ , Daiso Co., Osaka, Japan), and Sephadex LH-20 (Amersham

Biosciences, Uppsala, Sweden) were used for column chromatography.

#### 3.2 Plant material

The bark of *Z. simulans* was purchased from Hunan Corporation of Materia Medica, Hunan Province, China, in February 2011, and authenticated by the corresponding author (X.J. Zhou). A voucher specimen (No. ZHXJ-0012) has been deposited at our laboratory in Hunan University of Chinese Medicine.

#### 3.3 Extraction and isolation

The dried powdered bark of *Z. simulans* (30 kg) was extracted with ethanol ( $2 \times 180\text{l}$ ) to give an extract (3370 g), which was suspended in water and partitioned by petroleum ether and EtOAc (each  $4 \times 8\text{l}$ ), respectively. The EtOAc extract (470 g) was fractionated by a silica gel column eluted with  $\text{CHCl}_3$  with increasing amounts of methanol to afford seven fractions (Frs 1–7). Fr. 2 (89 g) was divided into five parts (Frs 2-1–2-5) by an MCI gel CHP 20P column eluting with

gradient aqueous MeOH. Fr. 2-5 (12 g) was first subjected to silica gel column (CHCl<sub>3</sub>/Me<sub>2</sub>CO, 50:1), followed by Sephadex LH-20 (CHCl<sub>3</sub>/MeOH, 6:4), and further purified by semi-preparative HPLC (SHIMADZU LC-10A HPLC system, Ultimate XB-C-18, 5 μm, 10 × 250 mm) eluting with 50% aqueous MeOH (flow rate, 2 ml/min) to produce compound **1** (24 mg; *t<sub>R</sub>* 17.02 min) and compound **3** (16 mg; *t<sub>R</sub>* 23.37 min). Fr. 2-1 (7.5 g) was chromatographed by silica gel column (CHCl<sub>3</sub>/Me<sub>2</sub>CO, 60:1), Sephadex LH-20 (CHCl<sub>3</sub>/MeOH, 6:4) and finally preparative thin layer chromatography (CHCl<sub>3</sub>/Me<sub>2</sub>CO, 20:1) to give compound **2** (9 mg; *R<sub>f</sub>* 0.45) and compound **4** (45 mg; *R<sub>f</sub>* 0.69).

### 3.3.1 Zanthoxylum A (**1**)

Colorless oil;  $[\alpha]_D^{21} - 42.8$  ( $c = 0.16$ , CHCl<sub>3</sub>); UV (CHCl<sub>3</sub>):  $\lambda_{\max}$  (log  $\epsilon$ ) 280.6 (2.67), 240.0 (2.85) nm; IR (KBr):  $\nu_{\max}$  1770, 1630, 1514, 1464, 1423, 1383, 1263, 1038 cm<sup>-1</sup>; for <sup>1</sup>H (400 MHz) and <sup>13</sup>C NMR (100 MHz) spectral data, see Table 1; EI-MS: *m/z* 318 [M]<sup>+</sup>; HR-EI-MS: *m/z* 318.1459 [M]<sup>+</sup> (calcd for C<sub>18</sub>H<sub>22</sub>O<sub>5</sub>, 318.1467).

### 3.3.2 Zanthoxylum B (**2**)

White solid;  $[\alpha]_D^{18.6} - 43.7$  ( $c = 0.11$ , MeOH); UV (MeOH):  $\lambda_{\max}$  (log  $\epsilon$ ) 280.5 (2.95), 227.5 (3.43) nm; IR (KBr):  $\nu_{\max}$  1630, 1610, 1514, 1463, 1419, 1384, 1267, 1138 cm<sup>-1</sup>; for <sup>1</sup>H (600 MHz) and <sup>13</sup>C NMR (150 MHz) spectral data, see Table 1; ESI-MS (positive): *m/z* 503 [M + Na]<sup>+</sup>; HR-EI-MS: *m/z* 480.2530 [M]<sup>+</sup> (calcd for C<sub>29</sub>H<sub>36</sub>O<sub>6</sub>, 480.2512).

### 3.3.3 (-)-Magnolin (**3**)

Colorless oil;  $[\alpha]_D^{23.5} - 15.3$  ( $c = 0.1$ , CHCl<sub>3</sub>); UV (C<sub>6</sub>H<sub>14</sub>):  $\lambda_{\max}$  (log  $\epsilon$ ) 276.0 (2.53), 203.6 (3.71) nm; IR (KBr):  $\nu_{\max}$  1514, 1463, 1419, 1383, 1126 cm<sup>-1</sup>; for

<sup>1</sup>H (600 MHz) and <sup>13</sup>C NMR (150 MHz) spectral data, see Table 1; EI-MS: *m/z* 416 [M]<sup>+</sup>; HR-EI-MS: *m/z* 416.1838 [M]<sup>+</sup> (calcd for C<sub>23</sub>H<sub>28</sub>O<sub>7</sub>, 416.1835).

### 3.3.4 (-)-Pinoresinol-di-3,3-dimethylallyl ether (**4**)

White solid,  $[\alpha]_D^{26.8} - 32.1$  ( $c = 0.3$ , CHCl<sub>3</sub>); UV (CHCl<sub>3</sub>):  $\lambda_{\max}$  (log  $\epsilon$ ) 281.6 (3.13), 241.2 (3.37) nm; IR (KBr):  $\nu_{\max}$  1585, 1516, 1466, 1417, 1264, 1236, 1139, 1069, 1028 cm<sup>-1</sup>; for <sup>1</sup>H (600 MHz) and <sup>13</sup>C NMR (150 MHz) spectral data, see Table 1; ESI-MS (positive): *m/z* 517 [M + Na]<sup>+</sup>; HR-EI-MS: *m/z* 494.2673 [M]<sup>+</sup> (calcd for C<sub>30</sub>H<sub>38</sub>O<sub>6</sub>, 494.2668).

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## Note

1. These authors contributed equally to this paper.

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