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One new dihydrobenzofuran lignan from Vitex trifolia

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A new benzofuran-type lignan, vitrifol A (1), was isolated from the fruits of *Vitex trifolia*, along with three known compounds 2-4. Their structures were elucidated mainly on the basis of 1D, 2D NMR and MS spectral data. Compound 1 was first isolated as a dimer of dihydrobenzofuran-type lignan.

Keywords: Vitex trifolia; Verbenaceae; vitrifol A; dimer of dihydrobenzofuran

1. Introduction

Vitex trifolia L. (Verbenaceae) has long been used as a medicinal plant and is widely distributed in Asia, Australia, and East Africa. The ripened fruits of this plant, Viticis Fructus, have traditionally been used as a folk medicine in China for the treatment of headache, cold, migraine, eyepain, and rheumatism, and its leaves have also been used for the treatment of wounds.¹ In early reports, flavonoids,^{2,3} sterols,² and diterpenes⁴⁻⁷ were isolated from this plant. In this paper, we present the isolation and structural elucidation of one new compound, vitrifol A (1), along with three known compounds, dihydrodehydrodiconifenyl alcohol (2),⁸ stigmast-4-en-6 β -ol-3-one (3),⁹ and eucommiol $(4)^{10}$ from Vitex trifolia. To our knowledge, all compounds were isolated from this plant for the first time.

2. Results and discussion

Compound 1 was obtained as pale yellow oil and has the molecular formula $C_{30}H_{34}O_9$ as

deduced by its negative HR-ESI-MS at m/z537.2139 $[M - H]^-$, requiring 14 degrees of unsaturation. The UV spectrum showed absorption maximum at 205 (4.84) nm, indicating the presence of an aromatic ring. The IR spectrum showed absorption bands at 3424, 1604, 1498 and 1465 cm^{-1} , revealing the presence of hydroxyl groups and aromatic rings. The ¹H NMR spectrum displayed signals at $\delta_{\rm H}$ 6.86 (1H, d, J = 1.8 Hz), 6.81 (1H, m) and 6.79 (1H, d, J = 8.1 Hz) for an AMX spin system of a 1,3,4-trisubstituted aromatic A-ring and four singlets (4H) at $\delta_{\rm H}$ 6.86, 6.84 and 6.62 for two tetrasubstituted aromatic ring with four benzylic methines. The remaining ¹H NMR signals indicated three methoxyl substituents, a *n*-propanol and two -O-CH-CH-CH₂Ospin systems. The ¹³C NMR spectrum consisted of 30 resonances (Table 1), including 11 quaternary carbons, of which six were oxygenated aromatic; seven aromatic and four aliphatic methines, of which two were oxymethines; five methylenes, of which three were oxygenated; and three methoxyls ($\delta_{\rm C}$

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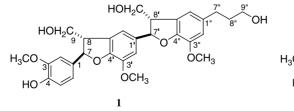
^{*}Corresponding author. Email: chenjj@mail.kib.ac.cn

Table 1. 1 H (400 MHz) and 13 C NMR (100 MHz) spectral data of compounds 1 and 2 (CDCl₃, δ in ppm).

No.	1		2	
	$\delta_{ m H}$	$\delta_{\rm C}$	$\delta_{ m H}$	δ_{C}
1		132.9 s		132.8 s
2	6.86 (1H, d, 1.8 Hz)	108.9 d	6.83 (1H, d, 1.5 Hz)	108.9 d
3		146.8 s		146.8 s
4		145.6 s		145.3 s
5	6.79 (1H, d, 8.1 Hz)	114.4 d	6.70 (1H, d, 8.1 Hz)	114.4 d
6	6.81 (1H, dd, 8.1, 1.8 Hz)	119.2 d	6.75 (1H, dd, 8.1, 1.5 Hz)	118.8 d
7	5.48 (1H, d, 7.5 Hz)	87.9 d	5.40 (1H, d, 6.9 Hz)	87.6 d
8	3.51-3.48 (1H, m)	53.6 d	3.43 (1H, m)	53.4 d
9	3.85 (2H, m)	63.4 t	3.52 (2H, m)	63.5 t
1'		134.7 s		135.1 s
2'	6.84 (1H, br s)	110.5 d	6.55 (1H, s)	112.0 d
3'		144.0 s		143.5 s
4′		148.2 s		145.8 s
5'		128.4 s		127.9 s
6′	6.86 (1H, br s)	114.4 d	6.57 (1H, s)	116.1 d
7′	5.49 (1H, d, 7.8 Hz)	88.3 d	2.51 (2H, t, 7.1 Hz)	31.5 t
8′	3.57-3.52 (1H, m)	53.8 d	1.73 (2H, m)	34.0 t
9′	3.76 (2H, m)	63.9 t	3.78 (2H, m)	61.3 t
1″		135.4 s		
2"	6.62 (1H, br s)	112.3 d		
3″		144.2 s		
4″		146.2 s		
5″		127.9 s		
6″	6.62 (1H, br s)	116.1 d		
7″	2.60 (2H, t, 7.5 Hz)	31.9 t		
8″	1.82 (2H, m)	34.4 t		
9″	3.61 (2H, t, 6.4 Hz)	61.8 t		
3-OMe	3.82 (s)	55.9 q	3.73 (s)	55.5 q
3'-OMe	3.80 (s)	55.9 q	3.67 (s)	55.5 q
3"-OMe	3.80 (s)	55.9 q	~ /	1

56.0). By comparing the IR, UV and NMR spectral data for **1** with the literature,¹¹ it showed that compound **1** was a dimer of dihydrobenzofuran neolignan. In the HMBC spectrum (Figure 2), the observed cross-peaks of H-7' ($\delta_{\rm H}$ 5.49, IH, d, J = 7.8 Hz) with C-1',C-2',C-6',C-8',C-9',C-4",H-7 ($\delta_{\rm H}$ 5.48,

1H,d,J = 7.5 Hz) with C-1,C-2,C-6,C-8,C-9 and C-4',H-2' ($\delta_{\rm H}$ 6.84,1H, br s), H-6' ($\delta_{\rm H}$ 6.86,1H, br s) with C-7', as well as the crosspeaks of H-2 ($\delta_{\rm H}$ 6.86, IH, d, J = 1.8 Hz) and H-6($\delta_{\rm H}$ 6.81, IH, m) with C-7, suggested that one dihydrobenzofuran moiety was connected to 1,3,4-trisubstituted aromatic ring (A) and the



HOH₂C, 8 5' 6' 7' 9' OH H₃CO 7 4' 3' OH HO 4_{5} 6 OCH₃ 2

Figure 1. Structures of compounds 1-2.

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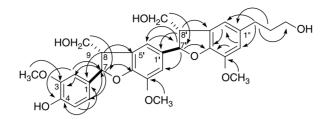


Figure 2. Key HMBC correlations of compound 1.

other dihydrobenzofuran structure (B and C). In the ROESY spectrum (Figure 3), the correlations observed between H-7 and H-9,H-7' and H-9', and the correlations between H-2 and H-8, H-6' and H-8' indicated threo arrangements for phenyl and hydroxymethylene groups at the C-7 and C-8, C-7' and C-8'. Comparison of the chemical shifts of C-7, C-8,C-9,C-7',C-8' and C-9' of **1** with those of **2** established that compounds **1** and **2** had the same configuration, which were further confirmed by the ROSEY experiment (Figure 3) and the coupling constants of H-7 and H-7.

3. Experimental

3.1 General experimental procedures

Optical rotations were measured with a Horiba SEPA-300 polarimeter. UV spectra were obtained using a Shimadzu UV-210A spectrophotometer. IR spectra were measured on a Bio-Rad FTS-135 spectrometer (Bio-Rad, Richmond, CA) with KBr pellets. MS spectra were performed on a VG Autospec-3000 spectrometer under 70 eV. 1D and 2D NMR spectra were recorded on Bruker AM-400 and DRX-500 spectrometer. Chemical shifts (δ) were expressed in ppm with reference to the solvent signals. Column chromatography was performed with silica gel (200-300 mesh, Qing-dao Marine Chemical Factory, Qingdao, China), silica gel H (60 µm, Qing-dao Marine Chemical Factory). Lichroprep RP-18 (43-63 µm, Merck, Darmstadt, Germany), MCI gel (75-150 µm, Mitsubishi Chemical Corporation, Japan) and Sephadex LH-20 (Pharmacia Fine Chemical Co. Ltd., Uppsala, Sweden) were also used for column chromatography. Fractions were monitored by TLC and spots were visualized by heating silica gel plates sprayed with 5% H_2SO_4 in EtOH.

3.2 Plant material

The fruits of *Vitex trifolia* were collected from Xishuangbanna, Yunnan Province, China, in May 2001, and identified by senior engineer Mr Jing-Yun Cui, Xishuangbanna Tropic Botanical Garden, the Chinese Academy of Sciences, where a voucher specimen (2001-2005) is deposited.

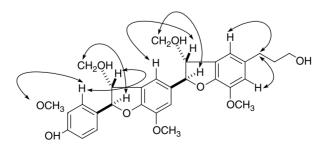


Figure 3. Key ROESY correlations of compound 1.

3.3 Extraction and isolation

The n-BuOH extract (240 g) was subjected to silica column chromatography gel $(10 \times 150 \text{ cm}, 200-300 \text{ mesh}, 1500 \text{ g})$ eluting with CHCl₃/MeOH/H₂O (from 9:1:0.1 to 7:3:0.3) to provide fractions A-J. Fraction A (10 g) was chromatographed on silica gel $(4.5 \times 60 \text{ cm}, 200 - 300 \text{ mesh}, 200 \text{ g})$ column with an elution of CHCl₃/Me₂CO (from 9:1 to 7:3) to obtain fractions A_1 - A_3 . Fraction A_3 (3g) was chromatographed on silica gel column $(3 \times 60 \text{ cm}, 200-300 \text{ mesh}, 90 \text{ g})$ eluting with CHCl₃/Me₂CO (8:2), followed by recrystallizing in CHCl₃ to yield compound 3 (120 mg). Fraction B (15 g) was submitted to a silica gel column $(4.5 \times 60 \text{ cm},$ 200-300 mesh, 300 g) using gradient elution of CHCl₃/Me₂CO (from 8:2 to 1:1) to obtain fractions B_1 - B_5 . Fraction B_3 (4 g) was performed on silica gel column $(4.5 \times 60 \text{ cm},$ silica gel H, 120 g) eluting with CHCl₃/EtOAc (7:3) to produce the mixture of compounds 1 and 2. The mixture was repeatedly purified over silica gel (silica gel H) eluting with petroleum ether/EtOAc (2.5:7.5) to afford compounds 1 (10 mg) and **2** (20 mg). Fraction J (30 g) was decolorized on MCI gel CHP-20 $(1.5 \times 30 \text{ cm})$ with an elution of MeOH/H₂O (90:10) to yield a pale yellow gum (15 g), which was subjected to silica gel, RP-18 (1.5×35 cm), and Sephadex LH-20 $(1.5 \times 60 \text{ cm})$ column subsequently to give compound 4 (727 mg).

3.3.1 Vitrifol A (1)

Pale yellow oil. $[\alpha]_D^{28} = +5.95$ (c = 0.20, MeOH); UV (MeOH) λ_{max} (log ε): 205 (4.84), 282 (3.80), 357 (2.51) nm; IR (KBr) ν_{max} : 3424, 2924, 2853, 1604, 1517, 1498, 1465, 1377, 1303, 1286, 1139, 1123, 1037, 982, 946, 852, 813, 744 cm⁻¹; ¹H and ¹³C NMR spectral data (see Table 1); negative ESI-MS: m/z 537 [M – H]⁻; negative HR-ESI-MS m/z: 537.2139 [M – H]⁻ (calcd for $C_{30}H_{33}O_9$, 537.2124).

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