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# New 9, 19-cycloartane triterpenoid from the root of *Cimicifuga foetida*

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#### [ABSTRACT]

AIM: To study the 9, 19-cycloartane triterpenes from the roots of Cimicifuga foetida.

**METHOD:** Chromatographic separations by silica gel, C<sub>18</sub> reversed phase silica gel, and high-performance liquid chromatography (HPLC) were used. All of the structures were elucidated on the basis of spectroscopic analysis and chemical methods.

**RESULTS:** Five 9, 19-cycloartane triterpenes,  $(3\beta, 12\beta, 15\alpha, 24R)$ -12, 2'-diacetoxy-24, 25-epoxy-15-hydroxy-16, 23-dione-3-O- $\alpha$ -L-arabinopyranoside (1), actein (2), 23-epi-26-deoxyactein (3), asiaticoside B (4), and  $12\beta$ -hydroxycimigenol (5) were isolated from the roots of *Cimicifuga foetida*.

**CONCLUSION:** Compound 1 is a new triterpene with two acetoxy groups at C-2' and C-12.

[KEY WORDS] Cimifuga foetida; Ranuncluaceae; 9, 19-Cycloartane triterpene; Acetoxy groups

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# Introduction

Cimicifuga species have a long history of being used as medicinal herbs <sup>[1]</sup>. In Europe and the United States, black cohosh (*C. racemosa*) is a well-known dietary supplement for women's health in alleviating menstrual pain and for menopausal disorders <sup>[2-3]</sup>. In China, the roots of *C. foetida* L., *C. dahurica* (Turcz.) Maxim., and *C. heracleifolia* Korn. are a source of a popular herbal medicine, "shengma", which has been used as an antipyretic and analgesic agent since ancient times <sup>[4-6]</sup>. Up to now, three main classes of compounds have been isolated from *Cimicifuga* spp: 9, 19-cycloartane triterpene glycosides, chromones, and cinnamic acid derivatives, of which the triterpene glycosides are considered

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to be the bioactive components <sup>[7]</sup>. Our research group has been studying the chemical constituents of *C. foetida*, *C. yumnanensis* P.K. Hsiao, *C. dahurica*, and *C. heracleifolia* <sup>[8-11]</sup> and reported a series of new cycloartane triterpenes, as well as their cytotoxic and anticomplement activities <sup>[12-15]</sup>. Further chemical investigations on the roots of *C. foetida* collected from Yulong County led to isolation of one new 9,19-cycloartane triterpene glycoside (1), together with four known compounds, actein (2) <sup>[16]</sup>, 23-*epi*- 26-deoxyactein (3) <sup>[17]</sup>, asiaticoside B (4) <sup>[18]</sup> and  $12\beta$ -hydroxycimigenol (5) <sup>[19]</sup>.

## **Results and Discussion**

Compound 1 was obtained as a white powder. The IR spectrum showed absorptions of hydroxyl (3 442 cm<sup>-1</sup>) and carbonyl groups (1 738 cm<sup>-1</sup>), respectively. Its molecular formula ( $C_{39}H_{58}O_{12}$ ) with eleven degrees of unsaturation was deduced from the analyses of <sup>13</sup>C-DEPT and HR-EI-MS data (m/z 718.393 7 [M]<sup>+</sup>; calcd. 718.392 8).

The  $^{1}$ H NMR spectrum of **1** (Table 1) showed the presence of the characteristic cyclopropane methylene signals at  $\delta_{\rm H}$  0.31 and 0.60 (each 1H, d, J=4.2 Hz), one secondary methyl signal at  $\delta_{\rm H}$  1.33 (d, J=6.6 Hz), and six tertiary methyl groups at  $\delta_{\rm H}$  0.95–1.58. Additionally, signals for an



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anomeric proton at  $\delta_{\rm H}$  4.76 (1H, d, J = 7.8 Hz) and two acetyl methyl groups at  $\delta_{\rm H}$  2.14 and 2.25 were observed. The  $^{13}{\rm C}$  DEPT spectrum of 1 exhibited 39 signals, of which 30 were attributed to the aglycon, five to a pentose residue, and four to two acetyl groups. The  $^{13}{\rm C}$ -DEPT spectrum of the aglycon of 1 also showed two ketone carbonyls at  $\delta_{\rm C}$  219.59 and

205.63. The aforementioned data suggested that **1** was a 9,19-cycloartane triterpene glycoside with two acetoxy and two carbonyl groups. The NMR spectroscopic data of **1** (Table 1) closely resembled that of bugbanoside D, except for the presence of an additional acetoxy group and the absence of two olefinic carbons <sup>[20]</sup>.

Table 1 <sup>1</sup>H NMR and <sup>13</sup>C-DEPT data of compound 1 (600 MHz and 150 MHz, in pyridine-d<sub>5</sub>, J in Hz)

Position	$\delta_{\mathrm{C}}$	$\delta_{\mathrm{H}}$	Position	$\delta_{\mathrm{C}}$	$\delta_{\mathrm{H}}$	Position	$\delta_{\mathrm{C}}$	$\delta_{\mathrm{H}}$
1	32.45 (t)	1.53 (m)	12	77.85 (d)	5.54 (dd, 3.6, 10.2)	27	18.55 (q)	1.29 (s)
		1.10 (m)	13	45.14 (s)		28	12.29 (q)	1.15 (s)
2	30.22 (t)	2.28 (m)	14	47.16 (s)		29	25.81 (q)	1.09 (s)
		1.86 (m)	15	82.97 (d)	4.63 (s)	30	15.64 (q)	0.95 (s)
3	88.65 (d)	3.35 (dd, 4.2, 11.4)	16	219.59 (s)		C3-Sugar		
4	41.46 (s)		17	59.20 (d)	2.74 (d, 2.4)	Ara-1'	105.01 (d)	4.76 (d, 7.8)
5	47.31 (d)	1.29 (m)	18	14.09 (q)	1.58 (s)	2'	74.85 (d)	5.96 (t, 9.0)
6	20.97 (t)	1.54 (m)	19	31.43 (t)	0.60 (d, 4.2)	3'	72.97 (d)	4.21 (dd, 3.6, 9.0)
		0.73 (m)			0.31 (d, 4.2)	4'	70.30 (d)	4.31 (m)
7	26.74 (t)	2.13 (m)	20	27.27 (d)	2.89 (m)	5'	67.79 (t)	4.30 (m)
		1.17 (m)	21	23.56 (q)	1.33 (d, 6.6)			3.78 (d, 11.4)
8	47.34 (d)	1.93 (dd, 3.6, 10.2)	22	46.75 (t)	2.96 (m)	12- <u>C</u> OCH <sub>3</sub>	171.20 (s)	
9	20.73 (s)		23	205.63 (s)		12-CO <u>C</u> H <sub>3</sub>	21.80 (q)	2.25 (s)
10	27.65 (s)		24	66.00 (d)	3.65 (s)	2'- <u>C</u> OCH <sub>3</sub>	170.54 (s)	
11	36.63 (t)	2.80 (dd, 10.2, 15.6)	25	61.43 (s)		2'-CO <u>C</u> H <sub>3</sub>	21.80 (q)	2.14 (s)
		1.23 (dd, 3.6, 15.6)	26	24.86 (q)	1.31 (s)			

In the  $^{1}\text{H}$ - $^{1}\text{H}$  COSY spectrum (Fig. 1), a correlation of H-7 with H-8 was observed, which indicated that the double bond between C-7 and C-8 was reduced in **1**. In addition, the  $^{1}\text{H}$ - $^{1}\text{H}$  COSY correlations of a downfield resonance at  $\delta_{\rm H}$  5.96 (t, J=9.0 Hz) with H-3' ( $\delta_{\rm H}$  4.21) and H-1' ( $\delta_{\rm H}$  4.76), together with the HMBC correlation from a carbonyl group ( $\delta_{\rm C}$  170.54) to the proton resonance ( $\delta_{\rm H}$  5.96, t, J=9.0 Hz) located the acetoxy group at C-2'.

Fig. 1 Key HMBC and  $^1\mathrm{H}\text{-}^1\mathrm{H}$  COSY correlations of compound 1

The relative configurations of H-3, H-5, H-8, H-12, and H-17 were established as in the  $\alpha$ -orientation on the basis of the ROESY correlations of H-3/H-1'; H-3/H-5; H-8/Me-28; H-17/Me-28; and H-12/H-17 (Fig. 2). H-15 was elucidated as having a  $\beta$ -orientation on the basis of the ROESY correlation with Me-18. The absolute configuration of C-24 was assigned as R, by comparing the chemical shifts of H-24 and C-24 of 1 with those of known 9,19-cyclolanostane triterpene glycosides [20]. Therefore, the chemical structure of 1 was determined to be  $(3\beta, 12\beta, 15\alpha, 24R)$ -12, 2'-diacetoxy-24, 25-epoxy-15-hydroxy-16, 23-dione-3-O- $\alpha$ -L-arabinopyranoside.

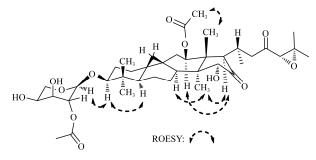


Fig. 2 Key ROESY correlations of compound 1

Compound **1** white powder (MeOH); UV (MeOH)  $\lambda_{max}$ : 203 and 240 nm; IR (KBr)  $\nu_{max}$ : 3 442 (OH), 2 937, 1 738 (C=O), 1 635 (C=C), 1 456, 1 379, 1 240, 1 146, 1 055, 923, 605 cm<sup>-1</sup>; positive HR-EI-MS m/z 718.3937 [M]<sup>+</sup>, (C<sub>39</sub>H<sub>58</sub>O<sub>12</sub>, Calcd. 718.3928), <sup>1</sup>H- and <sup>13</sup>C-DEPT data, see Table 1.

The known compounds actein (2), 23-epi-26-deoxyactein (3), asiaticoside B (4), and  $12\beta$ -hydroxycimigenol (5) were identified by comparing their physical and spectroscopic data with reported data.

#### **Experimental**

## General methods

Optical rotations were measured in MeOH with a Horiba SEAP-300 polarimeter.  $^{1}$ H- and  $^{13}$ C-DEPT spectra were recorded in pyridine- $d_{5}$  on Bruker DRX-500 and Avance III-600 MHz spectrometer (Bruker, Zűrich, Switzerland). ESI-MS and HR-EI-MS data were obtained using a Waters Autospec Premier-P776 spectrometer. Infrared spectrum was recorded on a Shimadzu IR-450 instrument with KBr pellets. Thin-layer chromatography was performed on precoated TLC plates (200–250  $\mu$ m thickness, silica gel 60 F<sub>254</sub>, Qingdao

Marine Chemical, Inc.) and spots were visualized by heating after spraying with 10% aq.  $\rm H_2SO_4$  soln. Semi-preparative HPLC was performed on an Agilent 1100 liquid chromatograph with a Zorbax SB-C-18 column (5  $\mu m$ , 4.6  $mm \times 150$  mm). Silica gel (200–300 mesh, Qingdao Marine Chemical, Inc.) and Lichroprep Rp-18 (40–63  $\mu m$ , Merck) were used for column chromatography (cc).

#### Plant material

The roots of *C. foetida* (82 kg) were collected in 2010 from Yulong County, Yunnan Province and identified by Prof. PEI Sheng-Ji, Kunming Institute of Botany, Chinese Academy of Sciences. A voucher specimen (KUN No. 20100906) has been deposited at the State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, China.

#### Extraction and isolation

The air-dried and powdered roots of C. foetida (82 kg) were refluedx with 95% MeOH (3  $\times$  100 L $\times$  5 h). The residue was extracted successively with EtOAc and n-BuOH. The EtOAc (5.6 kg) extract was subjected to silica gel cc, eluted with CHCl<sub>3</sub>-MeOH (CHCl<sub>3</sub>, 100 : 1, 50 : 1, 20 : 1, 5 : 1) to give five fractions (Fr. I-Fr. V). Fr. IV (350 g) was divided into five sub-fractions (Fr. IV.1-5) after performing silica gel cc, eluted with CHCl<sub>3</sub>-acetone (10:1). Compounds 2 (50 g) and 3 (20 g) were recrystallized from Fr. IV.5. Fr. IV.3 (10 g) and Fr. IV.4 (20 g) were repeatedly chromatographed over Rp-18 (50%, 60%, 70%, 80%, and 90% MeOH-H<sub>2</sub>O). Fr. IV.4.4 (80% MeOH-H<sub>2</sub>O, 860 mg) of Fr. IV.4 was subjected to cc on silica gel (1.5 g) eluting with CHCl<sub>3</sub>-acetone (5 : 1) to give fractions IV.4.4.1–7. Compounds 1 (1.2 mg) and 5 (20 mg) were purified from Fr. IV.4.4.6 (80 mg) by HPLC, eluted with CH<sub>3</sub>CN-H<sub>2</sub>O (45:55) (13.1 and 18.4 min). Fr. IV.3.4 (80% MeOH-H<sub>2</sub>O, 1.3 g) of Fr. IV.3 was further purified by repeated CC of silica gel (CHCl<sub>3</sub>-acetone, 25:1) and HPLC to obtain compound 4 (1.3 mg, CH<sub>3</sub>CN-H<sub>2</sub>O, 70: 30, 15.5 min).

## Hydrolysis and identification of the sugar moieties in compound 1

Compound 1 (1.2 mg) was dissolved in MeOH (5 mL), then 4%  $K_2CO_3$  (5 mL) was added, and the solution was stirred at rt overnight. The solution was neutralized with 10% HOAc and extracted with EtOAc (3 × 15 mL). The EtOAc extract, after removal of solvent, was dissolved in MeOH (5 mL) and refluxed with 0.5 mol·L<sup>-1</sup> HCl (3 mL) for 4 h. The aqueous layer was then neutralized with  $Ag_2CO_3$ , and the formed precipitate was filtered to give a monosaccharide, which had an  $R_f$  (EtOAc–CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O, 3 : 2 : 2 : 1) and specific rotation  $[\alpha]_D^{20} + 82.78$  (c 0.05, MeOH) corresponding to the data for of L-arabinose.

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