A Rarely Reported Trinorsesquiterpene-Type Structure in an Isolate from *Pulicaria insignis*

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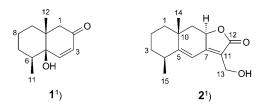
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A trinorsesquiterpene, **1**, and its possible precursor sesquiterpene **2** were obtained from the Tibetan folk medicine *Pulicaria insignis* (Ming \cdot chen \cdot serpo). Their structures were elucidated on the basis of spectral methods including 2D-NMR. The trinorsesquiterpene skeleton of **1** is the second example of this type of structure found in a plant, after a first compound of this type was isolated from bird's nest fungi (*Cyanthus bulleri*).

Introduction. – *Pulicaria insignis* has been traditionally used to reduce the symptoms of flu and common cold, including treatments of fever and pain-relief, although there are no human studies that have been done to support this [1]. Up to now, no chemical-component studies of this species have been reported. In an endeavor to find bioactive chemical compounds, we investigated the constituents of *Pulicaria insignis* and obtained trinorsesquitepene 1^1), which is the second example of this rarely reported structure type from plants [2], besides a possible precursor of **1**, eudesmane sesquiterpene 2^1) [3]. This discovery indicates that both plant and fungi can degrade eudesmane or germacrane sesquiterpenes, respectively, to such a trinorsesquiterpene skeleton in a similar oxidative biosynthetic way, which constitutes an important supplementary branch of the sesquiterpene biosynthesis and metabolism system. Both compounds **1** and **2** showed weak inhibitory activity against influenza virus H1N1 neuraminidase in an *in vitro* assay [4a]. At a concentration of 200 µg/ml, compounds **1**



1) Trivial atom numbering; for systematic names, see *Exper. Part.*

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and **2** showed $19.5 \pm 1.4\%$ and $18 \pm 0.7\%$ inhibition, respectively. Unfortunately, both **1** and **2** were very strongly toxic against the MDCK cells in the MTT (=2-(4,5-dimethylthiazol-2-yl)-3,5-dimethyl-2*H*-tetrazolium bromide) assay [4b]. Further modification of the two compounds will be necessary to reduce the toxicity while increasing the antiviral activity.

Results and Discussion. – From the HR-ESI-MS $(m/z \ 217.1203 \ ([M+Na]^+,$ $C_{12}H_{18}NaO_{7}^{+}$) and the DEPT data (2 Me, 4 CH₂, 3 CH, and 3 C), the molecular formula of **1** was deduced to be $C_{12}H_{18}O_2$. The ¹H-NMR spectrum (400 MHz, CDCl₃) of **1** (*Table*) showed two Me groups, one attached to a quaternary C-atom (δ 1.13 (s)) and one attached to a tertiary C-atom (δ 1.08 (d, J = 7.0 Hz)). H-C(3) (δ 5.97 (d, J =10.0 Hz)) and H–C(4) (δ 6.76 (d, J = 10.0 Hz)) in the ¹H-NMR spectrum together with C(2) (δ 200.7 (s)), C(3) (δ 130.1 (d)), and C(4) (δ 151.4 (d)) in the ¹³C-NMR spectrum (*Table*) were typical of an α . β -unsaturated ketone structure in a six-membered ring. The ¹H, ¹H-COSY revealed the correlations Me(11) (δ 1.08 (d, J = 7.0 Hz))/H-C(6) (δ 1.98 (dd, J = 7.0, 9.0 Hz)), $H - C(6)/CH_2(7)$ ($\delta 2.03 - 2.10$ and 1.43 (2 m)), $CH_2(7)/$ CH₂(8) (δ 1.71-1.77 and 1.47 (2 m)), and CH₂(8)/CH₂(9) (δ 1.89 and 1.18 (2 dd)). This indicated the connections C(6)-C(7)-C(8)-C(9) in another ring. In addition, in the HMBC spectrum the long-range correlations of $CH_2(1)$, H-C(4), H-C(6), $CH_2(9)$, and Me(12) with a quaternary C-atom at δ 40.3 (s, C(10)), and of H-C(4), H-C(6), and H–C(3) with an O-bearing C-atom at δ 73.5 (s, C(5) allowed to assign C(10) and C(5) (Fig.). Thus 1 was elucidated to be 4a,5,6,7,8,8a-hexahydro-4a-hydroxy-5,8adimethylnaphthalen-2(1H)-one. In the NOESY experiments, the NOE correlations

Table 1. ¹*H*- and ¹³*C*-*NMR Data* (400 and 100 MHz, resp.; CDCl₃) of Compounds **1** and **2**¹). δ in ppm, *J* in Hz.

	1		2	
	$\delta(H)$	$\delta(C)$	$\delta(H)$	$\delta(C)$
CH ₂ (1)	2.93 (d, J = 16.5),	51.1 (t)	1.62 - 1.64 (m),	39.6 (<i>t</i>)
	1.93 (d, J = 16.5)		1.68 (dd, J = 13.0, 4.0)	
$C(2) \text{ or } CH_2(2)$	_	200.7(s)	1.46 - 1.57 (m), 1.88 - 2.01 (m)	17.9 (t)
$H-C(3)$ or $CH_2(3)$	5.97 (d, J = 10.0)	130.1(d)	1.46 - 1.57 (m), 1.71 - 1.73 (m)	34.0 (t)
H-C(4)	6.76 (d, J = 10.0)	151.4 (d)	2.75 (ddq, J=7.5, 6.5, 2.0)	40.5(d)
C(5)	_	73.5 (s)	_	163.5 (s)
H-C(6)	1.98 (dd, J = 7.0, 9.0)	38.4(d)	6.36 (<i>s</i>)	112.6(d)
$CH_2(7)$ or $C(7)$	2.03–2.10 (<i>m</i>), 1.43 (br.)	26.8(t)	_	160.0(s)
$CH_{2}(8)$ or $H-C(8)$	1.71–1.77 (<i>m</i>), 1.47 (br.)	15.9(t)	4.78 (dd, J = 13.0, 6.0)	76.4(d)
CH ₂ (9)	1.89 (dd, J = 4.0, 13.0),	33.0 (t)	2.14 (dd, J = 13.0, 6.0),	43.1 (<i>t</i>)
	1.18 (dd, J = 0.5, 13.0)		1.54 (dd, J = 13.0, 13.0)	
C(10)	_	40.3(s)	_	38.5(s)
Me(11) or C(11)	1.08 (d, J = 7.0)	16.7(q)	_	118.3 (s)
Me(12) or C(12)	1.13 (s)	22.4(q)	_	174.8 (s)
CH ₂ (13)	_	-	4.40 (s)	55.3 (t)
Me(14)	_	-	1.32(s)	29.4(q)
Me(15)	_	-	1.29(d, J = 7.5)	20.6(q)
ОН	-	-	2.68 (br.)	-

 $H-C(6)/H_a-C(8)$, $H_a-C(8)/H_a-C(1)$, $H_{\beta}-C(7)/H_{\beta}-C(9)$, and $H_{\beta}-C(7)/Me(12)$ revealed that the two Me groups and the OH group of **1** were on the same side, thus establishing its relative configuration (*Fig.*).

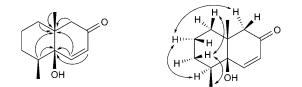
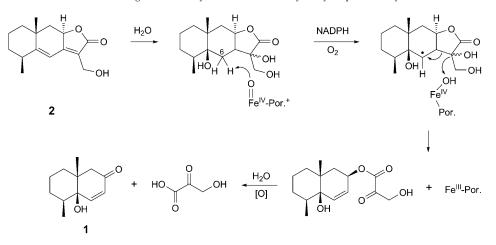


Figure. *HMBC* $(H \rightarrow C)$ and *ROESY* $(H \leftrightarrow H)$ correlation for compound **1**

Another sesquiterpene was identified as **2** by attributing the obtained NMR data to the reported structure (*Table*).

Structure 2 was considered to be the precursor of 1 based on the biogenetic route established by previous reports [5-14]; this biogenetic route is a fully enzyme-catalyzed route (*Scheme*).

Scheme. Biogenetic Route from 2 to 1 via Catalyses by a Special Enzyme



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Experimental Part

Isolation. The dried *Pulicaria insignis* was powdered and extracted with 95% MeOH. The residue was washed with petroleum ether to remove most fatty oil. The residue was then extracted with AcOEt, and the AcOEt extract separated by repeated column chromatography (*ODS*, 10-80% H₂O/MeOH): **1** (5 mg) and **2** (18.8 mg).

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rel-(4aR,5S,8aR)-4a,5,6,7,8,8a-Hexahydro-4a-hydroxy-5,8a-dimethylnaphthalen-2(1H)-one (1): White amorphous powder. $[\alpha]_{\rm D} = +40.5$ (c = 0.0014 g/ml, CHCl₃). IR (KBr): 3413 (OH), 1664 (C=O), 1618 (C=C). ¹H- and ¹³C-NMR: *Table*. ESI-MS: 217 ($[M+Na]^+$), 411 ($[2 M+Na]^+$). HR-ESI-MS: 217.1203 ($C_{12}H_{18}NaO_2^+$; calc. 217.1204).

rel-(5R,8aS,9aS)-6,7,8,8a,9,9a-Hexahydro-3-(hydroxymethyl)-5,8a-dimethylnaphtho[2,3-b]furan-2(5H)-one (2): Light yellow gum. $[a]_{\rm D} = +249.2 \ (c = 0.00915 \ {\rm g/ml}, \ {\rm CHCl}_3)$. ¹H- and ¹³C-NMR: *Table*. HR-ESI-MS (pos.): 271.1314 ($[M + {\rm Na}]^+, \ {\rm C}_{15}{\rm H}_{20}{\rm NaO}_3^+$; calc. 271.1310).

REFERENCES

- Editorial Board of Flora of China, 'Flora of China', Science Press, Beijing, 1997, Vol. 75, pp. 286– 293.
- [2] V. U. Ahmad, K. Fizza, A. Stultana, Phytochemistry 1989, 28, 3081.
- [3] Q.-X. Wu, Y.-P. Shi, Z.-J. Jia, Nat. Prod. Rep. 2006, 23, 699; Y. Chao, Q. X. Zhu, W. Yong, Z. J. Jia, Chin. Chem. Lett. 2001, 12, 597; W. A. Ayer, L. M. Browne, S. Fung, Can. J. Chem. 1976, 54, 3276.
- [4] a) C. Y. Su, S. Y. Wang, J. J. Shie, K. S. Jeng, N. J. Temperton, J. M. Fang, C. H. Wong, Y. S. Cheng, *Antiviral Res.* **2008**, *79*, 199; b) A. H. Ory, T. C. Owen, J. A. Barltrop, J. G. Cory, *Cancer Commun.* **1991**, *3*, 207.
- [5] K. P. Adam, R. Thiel, J. Zapp, H. Becker, Arch. Biochem. Biophys. 1998, 354, 181.
- [6] U. Warmers, W. A. Köning, Phytochemistry 2000, 53, 645.
- [7] V. Stanjek, M. Miksch, P. Lueer, U. Matern, W. Boland, Angew. Chem., Int. Ed. 1999, 38, 400.
- [8] M. G. El-Ghazouly, N. A. El-Sebakhy, A. A. S. El-Din, Z. C. Bohlmann, *Phytochemistry* 1989, 28, 1949.
- [9] P. K. Chakravarty, S. Tyagarajan, T. L. Shih, S. Salva, C. Snedden, M. J. Wyvratt, M. H. Fischer, P. T. Meinke, Org. Lett. 2002, 4, 1291.
- [10] H. Wehlan, M. Dauber, M.-T. Mujica Fernaud, J. Schuppan, R. Mahrwald, B. Ziemer, M.-E. Juarez Garcia, U. Koert, Angew. Chem., Int. Ed. 2004, 43, 4597.
- [11] B. Wu, J. M. Karle, E. B. Watkins, M. A. Avery, Tetrahedron Lett. 2002, 43, 4095.
- [12] Y.-K. Wu, H.-J. Liu, J.-L. Zhu, Synlett 2008, 4, 621; J. L. Shamshina, T. S. Snowden, Org. Lett. 2006, 8, 5881; B. A. Barner, 'e-EROS: Encyclopedia of Reagents for Organic Synthesis', Online Database, J. Wiley &Co., 2001.
- [13] M. T. Rogers, J. L. Burdett, Can. J. Chem. 1965, 43, 1516; J. L. Burdett, M. T. Rogers, J. Am. Chem. Soc. 1964, 86, 2105; G. M. McCann, C. M. McDonnell, L. Magris, R. A. M. O'Ferrall, J. Chem. Soc., Perkin Trans. 2002, 2, 784; N. Capponi, I. G. Gut, B. Hellrung, G. Persy, J. Wirz, Can. J. Chem. 1999, 77, 605; J. R. Keeffe, A. J. Kresge, N. P. Schepp, J. Am. Chem. Soc. 1988, 110, 1993; N. Capponi, I. Gut, J. Wirz, Angew. Chem., Int. Ed. 1986, 25, 344.
- [14] J. R. Keeffe, A. J. Kresge, in 'The Chemistry of Enols', Ed. Z. Rappoport, John Wiley & Co., New York, 1990, Chapt. 7, p. 399.

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