Daphlongamines I and J. New Yuzurine-type Alkaloids from *Daphniphyllum longeracemosum*

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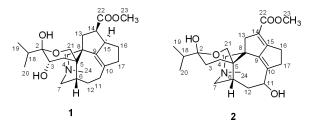
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Two new Yuzurine-type *Daphniphyllum* alkaloids, daphlongamines I and J (1, 2), were isolated from the fruits of *Daphniphyllum longeracemosum*. Their structures were established by spectroscopic methods, especially 2D NMR techniques.

Key words: Daphniphyllum Alkaloids, Yuzurine, Daphlongamine, Daphniphyllum longeracemosum

Introduction

Daphniphyllum alkaloids are a family of structurally specific and complex metabolites elaborated by trees of genus Daphniphyllum [1-3]. The unusual ring systems have attracted great attention for total synthesis [4] as well as biosynthetic studies [5]. In our continuing search for biogenetically and structurally interesting alkaloids [6, 7], two new Daphniphyllum alkaloids, daphlongamines I and J (1, 2) were isolated from fruits of Daphniphyllum longeracemosum. Herein, we report the isolation and structure determination of 1 and 2.



Results and Discussion

Daphlongamine I (1), a colorless amorphous powder, was indicated to have the molecular formula C₂₄H₃₇NO₅ by HR-ESI-MS (m/z = 420.2743, [M+H]⁺, calcd. 420.2749), suggesting seven degrees of unsaturation. The IR absorption bands at 1735 and 3432 cm⁻¹ implied the existence of ester carbonyl and hydroxyl groups, respectively. The ¹³C NMR and DEPT spectra displayed 24 carbon atoms (Table 1), including one carbonyl ($\delta_{\rm C} = 177.4$), one olefinic bond $(\delta_{\rm C} = 135.0, 147.7)$, three sp^3 quaternary carbons, five methines, nine methylenes, and four methyl groups (one MeO, one MeN). Among them, two methylenes $(\delta_{\rm C} = 57.5, 62.9)$ were ascribed to those connected to the N atom, one quaternary carbon ($\delta_{\rm C}$ = 99.4), one methine ($\delta_{\rm C} = 65.8$) and one methylene carbon atom $(\delta_{\rm C} = 63.3)$ were deduced to be those bearing the oxygen atom. Detailed analyses of the 1D NMR data suggested that 1 should be a yuzurine-type Daphniphyllum alkaloid [1] containing a hexacyclic ring system. The position of the double bond and the hydroxyl group were further established by the analyses of 2D NMR spectra including ¹H-¹H COSY, HSQC, and HMBC data. The ¹H-¹H COSY spectrum indicated the presence of four partial fragments: a (C-18 to C-19 and C-20), b (C-3 to C-4), c (C-6 to C-7 and C-12, and C-11 to C-12), and d (C-13 to C-17) drawn with bold bonds in Fig. 1A. In the HMBC spectrum, correlations of H-18 with the oxygenated carbon atoms C-2 ($\delta_{\rm C}$ = 99.4) and C-3 ($\delta_{\rm C}$ = 65.8) implied that C-2 and C-3 were substituted by hydroxyl groups of hemiketal and hydroxyl functionalities, respectively. The existence of the double bond between C-9 and C-10 was verified by the HMBC correlations of H-15 to C-9 and C-10, as well as H₂-17 to C-9 and C-10. Thus, the gross structure of daphlongamine I was established as 1 shown in Fig. 1A.

The relative stereochemistry of **1** was elucidated by ROESY data as shown in a computer-generated 3D drawing (Fig. 1B). The ROESY correlations observed between the proton pairs of H-21b/H-6 and

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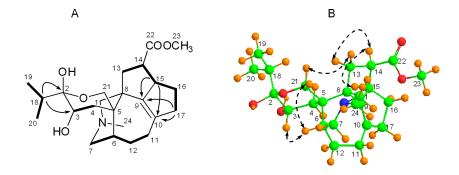


Table 1. 1 H (400 MHz) and 13 C (100 MHz) data of Daphlongamines I (1) and J (2).

No.	$\delta_{\rm H}$ (mult., <i>J</i> in Hz) ^{<i>a</i>}	$\delta_{\rm C}{}^{\rm a}$	$\delta_{\rm H}$ (mult., <i>J</i> in Hz) ^b	$\delta_{\rm C}{}^{ m b}$
1a	2.76 (1H, m)	57.5 t	2.42 (2H, m)	58.6 t
1b	2.68 (1H, m)			
2	-	99.4 s	-	97.6 s
3	3.85 (1H, dd, 11.2, 5.6)	65.8 d	1.57 (2H, m)	24.5 t
4a	1.45 (1H, t, 12.0)	33.7 t	1.93 (1H, m)	21.5 t
4b	2.23 (1H, m)		1.73 (1H, m)	
5	-	41.9 s	-	34.9 s
6	2.22 (1H, m)	36.4 d	2.43 (1H, m)	33.0 d
7a	2.39 (1H, d, 12.0)	62.9 t	2.96 (1H, m)	54.6 t
7b	2.27 (1H, d, 12.0)		2.74 (1H, m)	
8	-	48.0 s	-	46.0 s
9	-	147.7 s	-	148.9 s
10	-	135.0 s	-	152.7 s
11a	2.46 (1H, m)	28.5 t	4.17 (1H, m)	65.9 d
	2.05 (1H, m)			
12a	2.24 (1H, m)	28.6 t	2.06 (1H, m)	34.4 t
12b	1.63 (1H, m)		2.31 (1H, m)	
13a	2.72 (1H, dd, 14.8, 2.8)	41.1t	2.84 (1H, m)	39.8 t
13b	1.65 (1H, m)		3.20 (1H, m)	
14	2.90 (1H, dt, 9.6, 2.7)	44.0 d	-	118.6 s
15	3.45 (1H, m)	56.4 d	-	168.3 s
16a	1.28 (1H, m)	29.4 t	2.70 (1H, m)	25.7 t
16b	1.83 (1H, m)			
17a	2.55 (1H, m)	43.8 t	2.35 (1H, d, 15.0)	43.6 t
	2.32 (1H, m)		3.25 (1H, d, 15.0)	
18	1.97 (1H, dd, 13.9, 6.2)	36.9 d	1.69 (1H, m)	38.8 d
19	0.99 (3H, d, 6.9)		0.92 (1H, d, 7.0)	16.8 q
20	0.97 (3H, d, 6.9)	18.4 q	0.92 (1H, d, 7.0)	16.2 q
21a	4.26 (1H, d, 12.5)	63.3 t	3.76 (1H, d, 9.6)	62.5 t
	3.78 (1H, dd, 12.5, 3.0)		3.36 (1H, dd,	
			9.6, 2.4)	
22	-	177.4 s	-	166.5 s
23	3.61 (3H, s)	51.9 q	3.71 (3H, s)	51.1 q
24	2.21 (3H, s)	1	2.38 (3H, s)	45.1 q
^a Massured in CDaOD: ^b massured in CDCla				

^a Measured in CD₃OD; ^b measured in CDCl₃.

H-6/H-3 suggested that H-6 and C-3 were in a β -orientation. Furthermore, H-14 and H-15 were determined to be in an α -configuration by the correlations between H-13a/H-14, as well as H-14/H-15.

Daphlongamine J (2) was obtained as a colorless solid. Its molecular formula was determined as Fig. 1. (A) ${}^{1}H{}^{-1}H$ COSY (bold) and HMBC (arrow, $H{\rightarrow}C$) correlations of **1**. (B) NOESY correlations of **1**.

C₂₄H₃₅NO₄ by HR-ESI-MS (m/z = 418.2743, calcd. 418.2749), with 8 degrees of unsaturation. Extensive comparison of 1D NMR data of **2** (Table 1) with those of **1** suggested that both compounds were similar, except that there was one more olefinic bond in **2**, which was confirmed by the 2D NMR data (¹H-¹H COSY, HSQC, and HMBC). In the HMBC spectrum, the observed cross-peaks of H₂-12 with C-11 at $\delta_C = 62.5$, and H-11 with C-9 and C-10 implied that C-11 was substituted by a hydroxyl group, instead of C-3 in **1**. In addition, the conjugated double bonds between C-8/C-9 and C-14/C-15 were suggested by the HMBC correlations of H₂-13 to C-14 and C-15, and H₂-16 to C-9, C-10 and C-15.

The ROESY spectrum suggested that 2 had the same relative stereochemistry as 1. However, the relative configuration of the hydroxyl functionality at C-11 could not be determined yet.

Experimental Section

General experimental procedures

IR spectra were measured with a Bio-Rad FTS-135 spectrometer from KBr pellets. Optical rotations were obtained on a Perkin-Elmer model 241 polarimeter. ESI and highresolution mass spectra were recorded on a Finnigan MAT 90 instrument and a VG Auto Spec-3000 spectrometer. 1D and 2D NMR spectra were measured on Bruker DRX-500 or AM-400 spectrometers, using TMS as internal standard, and chemical shifts were recorded as δ values. Column chromatography was performed on silica gel H (10–40 μ m; Qingdao Marine Chemical Factory) and Sephadex LH-20 (40–70 μ m, Amersham Pharmacia Biotech AB, Uppsala, Sweden).

Plant material

The Fruits of *Daphniphyllum longeracemosum* were collected in Hekou of Yunnan Province, People's Republic of China, in October 2005. The sample was identified by Prof.

Xun Gong, Kunming Institute of Botany, Chinese Academy of Sciences, and a voucher specimen (KIB 05110021) was deposited at the State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences.

Extraction and isolation

The fruits (60 kg) of D. longeracemosum were extracted with 95% EtOH, and the crude extract was adjusted to pH = 2 with 2% HCl. The acidic mixture was defatted with petroleum ether (PE), which was followed by extraction with CHCl₃. The pH of the aqueous layer was roughly adjusted to pH = 10 with 3 % NaOH, and then exhaustively extracted with CHCl3 to give the crude alkaloid. The crude extracts were subjected to a silica gel column (CHCl3/MeOH, $1:0 \rightarrow 0:1$) to obtain six major fractions (F1-F6). Fraction 3 was further chromatographed over a silica gel (300-400 mesh) column (PE/Et₂NH, 50:1 \rightarrow 10:1) followed by a Sephadex LH-20 column eluted with CH₃OH to afford daphlongamine J (2, 7.0 mg). Fraction 4 was subjected to a RP-18 silica gel column (MeOH-H₂O) to give 4 parts (P1-P4). Part 3 was subjected to silica gel (PE/EtOAc/Et₂NH, $50:1:0.2 \rightarrow 10:1:0.2$) followed by Sephadex LH-20 col-

- For a review of *Daphniphyllum* alkaloids, see: J. Kobayashi, H. M. orita in *The Alkaloids*, Vol. 60, (Ed.: G. A. Cordell), Academic Press, New York, **2003**, pp. 165 – 205, and refs. cited therein.
- [2] a) Q. Zhang, Y.T. Di, C.S. Li, X. F. ang, C.J. Tan, Z. Z. hang, Y. Z. hang, H. P. He, S.L. Li, X. J. Hao, Org. Lett. 2009, 11, 2357–2359; b) C.S. Li, Y. T. Di, H. P. He, S. G. ao, Y. H. Wang, Y. L. u, J.L. Zhong, X. J. Hao, Org. Lett. 2007, 9, 2509–2512; c) Y. T. Di, H. P. He, Y.S. Wang, L. B. Li, Y. L. u, J. B. Gong, X. F. ang, N. C. Kong, S.L. Li, H.J. Zhu, X.J. Hao, Org. Lett. 2007, 9, 1355–1358; d) Y. Zhang, H. P. He, Y. T. Di, S. Z. Mu, Y. H. Wang, J. S. Wang, C. S. Li, N. C. Kong, S. G. Gao, X. J. Hao, Tetrahedron Lett. 2007, 48, 9104–9107; e) C. S. Li, H.P. He, Y.T. Di, Y. H. Wang, S. Z. Mu, S. L. Li, S. G. ao, Z. L. Gao, X. J. Hao, Tetrahedron Lett. 2007, 48, 2737–2740.
- [3] a) S. Saito, T. K. ubota, E. F. ukushi, J. K. awabata, H. P. Zhang, J. K. obayashi, *Org. Lett.* 2007, *9*, 1207– 1209; b) Z. Y. Li, P. C. heng, H. G. Xu, Y. M. Yang, S. Y. Peng, Z. Z. Zhao, Y. W. Guo, *Org. Lett.* 2007, *9*, 477–480; c) C. Q. Fan, S. Y. in, J. J. Xue, J. M. Yue, *Tetrahedron* 2007, *63*, 115–119; d) W. Zhang, Y. W. Guo, K. K. rohn, *Chem. Eur. J.* 2006, *12*, 5122–5127.
- [4] a) S. E. Denmark, R. Y. Baiazitov, J. Org. Chem. 2006,

umn chromatography eluted with CH_3OH to isolate daphlongamine I (1, 21.0 mg).

Identification

Daphlongamine I (1): Colorless powder. – ¹H NMR and ¹³C NMR data: see Table 1. – $[α]_D^{24} = -35.03$ (c = 0.59, CH₃OH). – IR (KBr): $v_{max} = 3432$, 2932, 2835, 1734 cm⁻¹. – MS ((+)-ESI): m/z = 420 [M+H]⁺. – HRMS ((+)-ESI): m/z = 420.2743 (calcd. 420.2749 for C₂₄H₃₈NO₅⁺, [M+H]⁺).

Daphlongamine J (2): Colorless solid. – ¹H NMR and ¹³C NMR data: see Table 1. – $[α]_D^{27}$ = 39.02 (*c* = 0.39, CH₃OH). – UV (MeOH): $λ_{max}(\log ε)$ = 295 (2.57). – IR (KBr): v_{max} = 3383, 2928, 2873, 1702, 1628 cm⁻¹. – MS ((+)-ESI): *m/z* = 418 [M+H]⁺. – HRMS ((+)-ESI): *m/z* = 418.2598 (calcd. 418.2593 for C₂₄H₃₆NO₅⁺, [M+H]⁺).

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71, 593-605; b) D. Sole, X. Urbaneja, J. Bonjoch, *Org. Lett.* **2005**, 7, 5461-5464.

- [5] a) H. Niwa, Y. H. irata, K. T. Suzuki, S. Y. amamura, *Tetrahedron Lett.* **1973**, *14*, 2129–2132; b) K. T. Suzuki, S. O. kuda, H. N. iwa, M. T. oda, Y. H. irata, S. Y. amamura, *Tetrahedron Lett.* **1973**, *14*, 799–802.
- [6] a) C. S. Li, Y. T. Di, S. Z. Mu, H. P. He, Q. Z. hang, X. F. ang, Y. Z. hang, S. L. Li, Y. L. u, Y. Q. Gong, X. J. Hao, *J. Nat. Prod.* **2008**, *71*, 1202 1206; b) Q. Zhang, Y. T. Di, H. Y. Liu, N. C. Kong, S. G. ao, C. S. Li, Y. Z. hang, S. L. Li, X. J. Hao, *Helv. Chim. Acta.* **2008**, *91*, 914 919; c) Y. T. Di, L. L. Liu, C. S. Li, Y. Z. hang, Q. Z. hang, S. Z. Mu, Q. Y. Sun, F. M. Yang, H. Y. Liu, X. J. Hao, *Helv. Chim. Acta* **2008**, *91*, 838 843; d) S. Z. Mu, J. S. Wang, X. S. Yang, H. P. He, C. S. Li, Y. T. Di, Y. W. ang, Y. Z. hang, X. F. ang, L. J. Huang, X. J. Hao, *J. Nat. Prod.* **2008**, *71*, 564 569.
- [7] a) S. Z. Mu, C. S. Li, H. P. He, Y. T. Di, Y. W. ang, Y. H. Wang, Z. Z. hang, Y. L. u, L. Z. hang, X. J. Hao, J. Nat. Prod. 2007, 70, 1628 1631; b) N. C. Kong, H. P. He, Y. H. Wang, S. Z. Mu, Y. T. Di, X. J. Hao, J. Nat. Prod. 2007, 70, 1348 1351; c) S. Z. Mu, Y. T. Di, H. P. He, Y. W. ang, Y. H. Wang, L. L. i, X. J. Hao, Chem. Biodivers. 2007, 4, 129 138; d) N. C. Kong, H. P. He, Y. H. Wang, S. Gao, Y. T. Di, X. J. Hao, Helv. Chim. Acta 2007, 90, 972 976.