## Chemical Constituents of New Diterpenes From Taxus yunnanensis

LIU Xi-Kui (刘锡葵), WU Da-Gang (吴大刚) and WANG Zong-Yu (王宗玉)

(Kunming Institute of Botany, Academia Sinica, Kunming 650204, PRC)

Received July 25, 1992

Keywords: Taxaceae, Taxus yunnanensis, diterpene, taxayunnansin A.B, taxol.

Recently, the intensive investigations of chemical constituents of Taxaceae, especially the antitumor diterpenes related to taxane, have already been stimulated by the significant discovery of antitumor activity of taxol<sup>[1]</sup>. Such studies will- eventually result in the development of a new type of chemotherapeutic antitumor agents.

Taxus yunnanensis Cheng et L. K. Tu, is distributed in China, especially in Yunnan Province as an especial species. Two new diterpenoid esters taxayunnansin A(1) and B(2) were isolated from EtOH extract existing between water and chloroform from the branch of *T. yunnanensis*. The chromatography of the chloroform fraction on silica gel afforded the two compounds by elution with CHCl<sub>3</sub>-MeOH. Taxayunnansins A mp. 338—340°C and taxayunnansin B mp. 318—319°C were crystallized in diluted EtOH as white tabular crystals. Their structures are related to the isotaxane skeleton, which was previously derived by the rearrangement of taxol with the electrophilic reagent<sup>[2]</sup>. Structurally, this carbon skeleton arises from the cleavage of  $C_{11}$ — $C_{12}$  bond in taxane, so that carbon  $C_1$  is attached to  $C_{11}$  to deduce an isopropyl  $C_{15}$ ,  $C_{16}$  and  $C_{17}$ . The group is oxidated as a hydroxyl isopropyl in taxayunnansins A and B, its signals appear at 1.07 (3H, s, CH<sub>3</sub>), 1.691(3H, s, CH<sub>3</sub>) ppm and 74.82(C), 12.48(CH<sub>3</sub>), 25.19(CH<sub>3</sub>) ppm in the <sup>1</sup>HNMR and <sup>13</sup>CNMR spectra respectively.

Fig. 1. Molecular structure. 1;  $R_1 = OAc$ ,  $R_2 = OH$ . 2;  $R_1 = OH$ ,  $R_2 = OAc$ .

The structures of compounds taxayunnansins A and B were elucidated by UV, IR, MS, FAB-MS, <sup>1</sup>HNMR and <sup>13</sup>CNMR spectra, including <sup>1</sup>H-<sup>1</sup>HCOSY, <sup>1</sup>H-<sup>13</sup>CCOSY, DEPT and COLOG spectroscopic experiments. Their spectroscopic data are

summarized in the Tables 1 and 2. X-ray diffraction analysis was undertaken to confirm the structure proposed for taxayunnansin A and to reveal the relative configuration of the diterpenes in taxayunnansins A and B (Fig. 2).

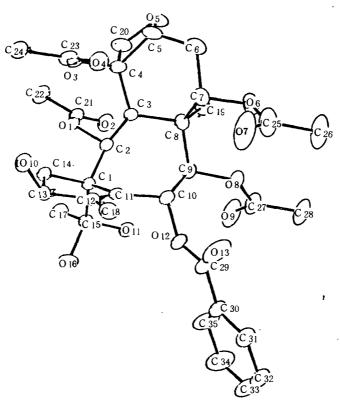


Fig. 2. X-ray diffraction of taxayunnansin A.

Table 1 UV, IR, MS and Mp. Data of Taxayunnansins A and B

Compound	Taxayunnansin A	Taxayunnansin B C <sub>35</sub> H <sub>44</sub> O <sub>13</sub>	
Formula	C <sub>35</sub> H <sub>44</sub> O <sub>13</sub>		
Mp. (°C)	338-340	318-319	
UV (nm)	206, 229, 271	205, 229, 271	
IR (KBr, cm <sup>-1</sup> )	3525, 2979, 2930, 2880,	3525, 2979, 2960, 2930,	
	1735, 1721, 1650, 1591,	2880, 1735, 1721, 1655,	
	1575, 1441, 1430, 1362,	1595, 1579, 1445, 1431,	
	1330, 1260, 1230, 1180,	1370, 1330, 1260, 1240,	
	1130, 1085, 1065, 1021,	1180, 1130, 1085, 1065,	
	710.	1021, 710.	
MS(m/z)	$695[M + Na]^+$ , $673[M + 1]^+$ ,	594[M-18-60], 494,	
	635, 611, 573, 551, 491,	490, 447, 372, 312,	
	431, 373, 313, 293, 253,	252, 122, 105, 77,	
	223, 195, 106, 91, 77,	60, 43 (MS. 20 eV).	
	59. (FAB-MS. glycerol)		

Table 2 <sup>1</sup>H and <sup>13</sup>CNMR. DEPT Data of Taxayunnansins A and B<sup>a)</sup>

Carbon	Taxayunnansin A			Taxayunnansin B		
	$\sigma_c$	$\sigma_{ m H}$	J(Hz)	$\sigma_{ m c}$	$\sigma_{\rm H}$	J(Hz)
1. <b>C</b>	67.6			67.7		
2. CH	69.1	6.06d	7.8	69.3	6.07d	7.8
3. CH	44.0	3.08d	7.7	44.1	3.10d	7.8
4. C	79.7			<i>7</i> 9.9		
5. CH	84.7	4.90d	7.6	85.0	4.91d	7.8
6. CH <sub>2</sub>	34.7	2.20dd	16.8, 7.0	34.8	2.20m	
		1.57dd	16.8, 7.2		1.58m	
7. CH	67.9	5.50t		68. I	4.46d	7.8
8. C	43.6			43.8		
9. CH	76.4	6.10d	10.8	76.8	6.11d	11.8
10. CH	77.3	6.49d	10.8	77.1	6.50d	3.11
11. <sup>'</sup> C	133.8			133.9		
12. C	151.5			151.6		
13. CH	67.6	4.45d	7.0	70.4	5.50t	8.2
14. CH <sub>2</sub> 39.5	39.5	2.49ddd	15.6, 7.6	39.6	2.48m	
			8.2			
		1.84ddd	15.6, 8.6		1.86m	
			6.0			
15. C	74.8			75.0		
16. · CH <sub>3</sub>	12.5	1.07s		12.6	1.07s	
17. CH <sub>3</sub>	25.2	1.69s		25.3	1.64s	
18. CH <sub>3</sub>	27.4	1.746		27.5	1.70s	
19. CH <sub>3</sub>	11.8	1.02s		11.7	1.16s	
20. CH <sub>2</sub>	75.4	4.45d	7.6	75.5	4.44d	7.6
_		4.36d	7.6		4.36d	7.6
21.3 21.3 21.2 169.7 169.7 170.3	20.5	1.97s		20.6	1.98s	
	21.3	1.94s		21.4	1.98s	
	21.5	2.02s		21.6	2.03s	
	21.2	2.11s		22.3	2.12s	
	169.7			169.8		
	1 <del>69</del> .7			169.8		
	170.3			170.4		
	170.9			171.1		
O-Bz C	164.0			164.1		
C	129.0			129.2		
CH	129.4	7.79d	7.8	129.5	7.80d	7.6
	128.7	7.37dd	7.6, 7.4	128.8	7.37t	7.7
	133.3	7.50dd	7.2, 7.5	133.4	7.50t	7.4
ОН		2.18d	7.2		2.07d	6.8
		2.73s			2.71s	

a) NMR was obtained using Bruker AM-400 spectrometer, the reagent was CDCl<sub>3</sub>, chemical shift in ppm relative to internal TMS.

In addition, a known compound taxol (3) was isolated from the plant simultaneously. Its spectroscopic data are corresponding to that of taxol in literature<sup>[1, 2]</sup>.

The diterpene skeleton of taxayunnansins A and B is related to taxane, but due to their distinction, it should be named isotaxane, which was previously obtained from the products of the rearrangement of taxol<sup>[3]</sup>. In fact, the skeleton isotaxane naturally occurs in the plant. These naturally occurring examples provide a definite stimulus for considering the biosynthetic link between various groups of taxane types.

The authors wish to thank Chen Neng\_yu and Wang Qi-guang of the Centre of Analysis and Measuring, Lanzhou University for running FAB-MS and X-ray diffraction analysis, and the staff of analytical group for measuring of UV, IR and NMR spectra.

## References

- 1 Wani, M. C., Tayor, H. L. & Wall, M. E., J. Amer. Chem. Soc., 1971, 93: 2325.
- 2 Chen Wei-ming, Zhang Pei-ling, Wu Bin et al., Acta Pharmaceutica 1991, 26: 747.
- 3 Kingston, D. G. I., Samapa, Ake. G. & Ivey C. A., J. Nat. Prod., 1990, 53: 1.