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FOUR SESQUITERPENES FROM THE INSECTICIDAL PLANT CELASTRUS ANGULATUS

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Key Word Index—Celastrus angulatus; Celastraceae; sesquiterpenes; dihydro- β -agarofuran derivatives; insect antifeedant.

Abstract—The structures of four new highly esterified sesquiterpenes isolated from the insecticidal plant Celastrus angulatus were elucidated on the basis of spectral analysis, including 2D-NMR spectroscopy, especially long range ${}^{1}\text{H}{-}{}^{13}\text{C}$ correlation.

INTRODUCTION

Celastrus angulatus Maxim. has been used as a natural insecticide for a long time in China [1]. We recently reported on the isolation of 15 new sesquiterpenes based on the dihydro- β -agarofuran skeleton from this plant, and antifeedant effects against some insects were shown [2-7]. In recent years, compounds of this type have also attracted interest as a consequence of their cytotoxic and antitumour promoter activities [8-11]. Further investigation of *C. angulatus* has led to the isolation of four new sesquiterpenes (1-4). In the structural elucidation of these sesquiterpenes, it is difficult to determine the linkage sites of the respective ester groups because more than three kinds of acids are involved in ester formation. This problem can be solved conveniently by using long range ¹H-¹³C correlation (COLOC) techniques.

RESULTS AND DISCUSSION

The chloroform fraction of the acetone extract of the root-bark of *C. angulatus* was subjected to repeated chromatography (CC and HPLC) to afford compound 1. Its IR spectrum revealed the characteristic absorptions of ester carbonyl at 1730 cm^{-1} and hydroxyl at 3500 cm^{-1} . The mass spectrum exhibited peaks due to the loss of methyl, acetic acid and isobutyric acid units: m/z 679 $[M - Me]^+$, 634 $[HOAc]^+$ and 606 $[(M - C_3H_7COOH)]^+$. It contained also fragmentation ions attributable to the presence of furoylate (m/z 95), iso-

butyrate (m/z 71) and acetate (m/z 43) groups [12]. The ¹H and ¹³C NMR spectra (Table 1) indicated the presence of six ester groups (ester carbonyls at δ 160.9, 169.4, 169.6, 169.7, 175.6 and 176.8): three acetate esters ($\delta_{\rm H}$ 1.85, 1.63, 1.56; $\delta_{\rm C}$ 21.4, 21.1, 20.5), one furoylate ester ($\delta_{\rm H}$ 7.94 s, 6.62 bs, 6.78 bs; $\delta_{\rm C}$ 149.0, 118.0, 109.8, 144.0) and two isobutyrate esters ($\delta_{\rm H}$ 1.29 d, 1.28 d, 1.24 d, 1.20 d, 2.67 m, 2.78 m; $\delta_{\rm C}$ 19.0, 19.0, 34.1, 18.9, 18.7, 34.0). In addition, the ¹³C NMR spectrum showed signals attributed to three methyls (δ 29.6, 25.5, 24.5), two methylenes (δ 65.5, 42.1), six oxygenated methines (δ 68.1–75.6) and four quaternary carbons (δ 91.5, 83.5, 70.0, 54.1). The ¹HNMR spectrum contained signals assignable to protons on the carbon atoms carrying the five secondary ester groups, i.e. $\delta 6.42$ (1H, s, H-6), 5.99 (1H, s, H-9), 5.82 (2H, m, H-1, 2), 5.60 (1H, d, J = 2.8 Hz, H-8) and a primary ester group, i.e. δ 5.21, 5.02 (2H, AB_a, J = 12.7 Hz, H-11a, b) on the basis of the multiplicity and the ${}^{1}H{}^{-1}H$ COSY spectrum. These data suggested that 1 is a dihydro-\beta-agarofuran sesquiterpene substituted with six ester groups.

The positions of the ester groups were established unambiguously by using COLOC experiments. The carbonyl ¹³C signal at δ 160.9 showed long range correlations with the ¹H signals at δ 5.99 (H-9) and 6.62 (H-4'). Thus, the furoyl group must be located on C-9. The long range couplings of the carbonyl signals at δ 169.4, 169.6 and 169.7 with the proton signals at δ 6.42 (H-6) and 5.82 (H-1, 2) and the acetyl methyl signals at δ 1.85, 1.63 and 1.56 indicated the presence of acetyl groups on C-6, C-1 and C-2. The last two carbonyl ¹³C signals at δ 1.75.6 and 176.8 were correlated with the ¹H signals at δ 5.60 (H-8), 2.67 (H-22), 1.29 (H-20), 1.24 (H-21) and 5.21, 5.02 (H-11a, b), 2.78 (H-18), 1.28 (H-16) and 1.20 (H-17), respectively. Thus, the two isobutyryl groups had to be situated on

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C-8 and C-11. Therefore, a planar structure for 1 was established.

The relative stereochemistry of 1 was confirmed via the NOE difference spectra. When the H-11b signal was irradiated, an increase (16.3%) of the H-6 signal intensity and an increase (6.28%) of the H-12 signal intensity occurred. On irradiation of the H-1 and H-2 signal, the intensity of the H-3_{ax} signal was increased, while on irradiation of the H-11a signal, the intensity of the H-9 signal was increased (6.9%). In the ¹H NMR spectrum of 1, H-9 appeared as singlet. This is due to the angle 8α , 9 β adjacent to 90° as reported in the literature [13]. These results indicated that the orientations of the protons attached to the oxygen-bearing carbon atoms were H-1ax, H-2eq, H-6ax, H-8eq and H-9eq. Considering the above data for 1 the structure was elucidated to be 1β , 2β,6α-triacetoxy-4α-hydroxy-8β,11-diisobutyryloxy-9αfuroyloxydihydro-β-agarofuran.

Compound 2 was found to be spectroscopically similar to 1. Its spectral data (Table 2) suggested the presence of three acetate esters, two furoylate esters and one isobutyrate ester and the 1 β , 2 β , 6 α , 8 β , 9 α ,11-hexaesterified dihydro- β -agarofuran parent. Close comparison of the ¹H and ¹³C NMR spectra of 1 and 2 clearly revealed the furoylate at C-8 of 2 replacing an isobutyrate of 1. In the same way as in 1 (COLOC experiment), the positions of the three acetate esters were shown to be situated at C-1, C-2 and C-6, the two furoylate esters at C-8 and C-9, and the one isobutyrate ester at C-11, respectively. Thus, the structure of **2** is 1β , 2β , 6α -triacetoxy- 4α -hydroxy- 8β , 9α difuroyloxy-11-isobutyryloxydihydro- β -agarofuran.

Compound 3 was found to be spectroscopically very similar to 2. Close examination of the ¹H NMR spectrum of 3 clearly revealed the absence of an isobutyroyl function at C-11, as did the ¹³C NMR data when compared with those for 2. It was replaced by a 3-methyl-isobutyroyl group. All the remaining spectroscopic features of 3 were consistent with 2. So, the structure of 3 was established to be $1\beta,2\beta,6\alpha$ -triacetoxy-4 α -hydroxy-8 β , 9α -difuroyloxy-11-(3-methyl) isobutyroyloxydihydro- β -agarofuran.

Compound 4 was assigned as $1\beta_2\beta_11$ -triacetoxy-4 $\alpha_1, 6\alpha_2$ -dihydroxy- $8\alpha_2$ -isobutyryloxy- $9\beta_2$ -benzoyloxydihydro- β_2 -agarofuran based on the followig data. The IR spectrum showed hydroxyl and ester group bands. The mass spectrum exhibited peaks due to the losses of methyl and acetic acid units: m/z 619 [M – Me]⁺ and 574 [HOAc]⁺ and fragmentation ions attributable to the presence of benzoate (m/z 105), isobutyrate (m/z 71) and acetate (m/z 43) groups: one benzoate ester [$\delta_H 7.83 d, 7.53 t', 7.39 t'; \delta_c 128.7 (2 \times CH), 129.5 (1 \times C,$ $2 \times CH), 133.4 (CH) and 165.8 (ester carbonyl)], one$

Table 1. ¹H and ¹³C NMR spectral data for compound 1 (in C₆D₆)

		¹ H–H COSY				³ H- ¹³ C COSY	Significant COLOC
н	δ	J(Hz)	correlation with	С	δ (DEPT)	correlation with	correlation with
1	5.82 m			1	68.1 (CH)*	H-1	
2	5.82 m		H-3a, b	2	70.8 (CH)*	H-2	
3a	2.03 m		H-3b/H-2	3	42.1 (CH ₂)	H-3a, b	
3Ь	1.80 m		H-3a/H-2				
4				4	70.0 (C)		
5				5	91.5 (C)		
6	6.42 s		H-7 (w)	6	75.6 (CH)	H-6	
7	2.41 d	2.8	H-6 (w)/H-8	7	53.1 (CH)	H-7	
8	5.60 d	2.8	H-7/H-9 (w)	8	76.1 (CH)	H-8	
9	5.99 s		H-8 (w)	9	71.5 (CH)	H-9	
10				10	54.1 (C)		
11a	5.21 d	12.7	H-11b	11	65.6 (CH ₂)	H-11a,b	
11b	5.02 d	12.7	H-11a				
12	1.43 s			12	29.6 (CH ₃)	H-12	
13				13	83.5 (C)		
14	1.52 s			14	25.5 (CH ₃)	H-14	
15	1.27 s			15	24.5 (CH ₃)	H-15	
Fu							
2′	7.94 s		H-4'/H-5'	2′	149.0 (CH)	H-2′	
3′				3'	118.0 (C)		
4′	6.62 bs		H-2'/H-5'	4′	109.8 (CH)	H-4'	
5′	6.78 bs		H-2'/H-4'	5'	144.0 (CH)	H-5'	
6′				6'	160.9 (C)		H-9/H-4′
Bu							
16	1.28 d	7	H-18	16	19.0 (CH ₃)	H-16	
17	1.20 d	7	H-18	17	19.0 (CH ₃)	H-17	
18	2.78 m		H-16/H-17	18	34.1 (CH)	H-18	
19				19	176.8 (C)		H-11a, b/H-16, 17, 18
20	1.29 d	7	H-22	20	18.9 (CH ₃)	H-20	
21	1.24 d	7	H-22	21	18.7 (CH ₃)	H-21	
22	2.67 m		H-20/H-21	22	34.0 (CH)	H-22	
23			·	23	175.6 (C)		H-8/H-20, 21, 22
Ac							
24	1.85 s			24	21.4 (CH ₃)	H-24	
25				25	169.4 (C)		H-6/H-24
26	1.63 s			26	21.1 (CH ₃)	H-26	
27				27	169.6 (C)		H-1 (or H-2)/H-26
28	1.56 s			28	20.5 (CH ₃)	H-28	
29				29	169.7 (C)		H-2 (or H-1)/H-28

*Assignments in the same vertical column may be reversed.

w: weak.

isobutyrate ester $[\delta_H 0.87 d, 0.99 d, 2.32 m; \delta_c 18.5$ (CH₃), 18.6 (CH₃), 34.1 (CH)] and three acetate esters $[\delta_H 2.32 s, 2.06 s, 2.03 s; \delta_c 21.5$ (CH₃), 21.0 (CH₃), 20.5 (CH₃)]. In addition, the ¹³C NMR spectrum showed signals attributed to the remaining skeletal moiety: three methyls ($\delta 24.3$, 26.3 and 30.1), two methylenes ($\delta 41.3$ and 61.5), six methines ($\delta 53.8$, 67.5, 73.5, 75.0, 75.5 and 76.7) and four quaternary carbons ($\delta 50.8$, 72.2, 84.6 and 91.6). Thus, 4 is also a polyester dihydro- β -agarofuran sesquiterpene. On the basis of the multiplicity and the ¹H-¹H COSY spectrum, the signals at $\delta 6.04$, 5.60, 5.44, 5.36, 5.20, 4.76, 4.71 and 2.57 in the ¹H NMR spectrum of 4 were assigned to H-9, H-8, H-1, H-2, H-6, H-11a, b and H-7, respectively. The shifts of H-3, whose signals overlapped in the ¹H NMR spectrum, were ascertained by correlation with H-2. The relative stereochemistry of compound 4 was determined as H-1_{eq}, H-2_{eq}, H-6_{ax}, H-8_{ax} and H-9_{ax} according to the coupling constants of each proton $(J_{1,2} = 3.4 \text{ Hz}, J_{7,8} = 3.1 \text{ Hz}, J_{8,9} = 9.8 \text{ Hz})$ and by comparison with the data for angulatueoids A-D isolated from the same plant and for which X-ray data were available [4]. In order to determine the location of the ester groups, the COLOC spectrum of compound 4 was measured. Cross peaks were observed between the carbonyl carbon at $\delta 165.8$ and $\delta 6.04$ (H-9) and 7.83 (H-2', 6'), and thus the benzoyl group must be located on

Table 2. ¹H and ¹³C NMR spectral data for compound 2 (in CDCl₃)

н 	δ	J(Hz)	correlation with	С	δ (DEPT)	correlation with	Significant COLOC correlation with
1	5.52 m			1	70.4 (CH)*	H-1	
2	5.52 m		H-3a, b	2	68.1 (CH)*	H-2	
3a	1.94 m		H-2/H-3b	3	42.1 (CH ₂)	H-3	
3b	2.20 m		H-2/H-3a				
4				4	70.0 (C)		
5				5	91.4 (C)		
6	6.34 s		H-7	6	75.7 (CH)	H-6	
7	2.45 d	2.8	H-6/H-8	7	53.6 (CH)	H-7	
8	5.43 d	2.8	H-7	8	76.2 (CH)	H-8	
9	5.73 s			9	70.1 (CH)	H-9	
10				10	54.4 (C)		
11a	4.96 d	13	H-11b	11	66.0 (CH ₂)	H-11	
11b	4.76 d	13	H-11a				
12	1.57 s			12	29.7 (CH ₃)	H-12	
13				13	83.4 (C)		
14	1.68 s			14	25.5 (CH ₃)	H-14	
15	1.46 s			15	24.8 (CH ₃)	H-15	
Fu							
2′	8.19 s		H-4'/H-5'	2′	148.9 (CH)	H-2'	
3′				3′	118.6 (C)		
4′	6.83 s		H-2'/H-5'	4′	109.8 (CH)	H-4′	
5′	7.44 s		H-2'/H-4'	5'	143.9 (CH)	H-5'	
6'				6′	160.5 (C)		H-9
8′	7.99 s		H-10'/H-11	8′	148.7 (CH)	H-8′	
9′				9′	118.4 (C)		
10′	6.72 s		H-8'/H-11'	10′	109.8 (CH)	H-10′	
11′	7.41 s		H-8'/H-10'	11′	143.9 (CH)	H-11'	
12′				12′	161.4 (C)		H-8
Bu							
16	1.12 d	7	H-18	16	18.4 (CH ₃)	H-16	
17	1.05 d	7	H-18	17	18.4 (CH ₃)	H-17	
18	2.56 m		H-17/H-17	18	34.0 (CH)	H-18	
19				19	176.7 (C)		H-11/H-16, 17, 18
Ac							
20	2.17 s				21.4 (CH ₂)	H-20	
21	2.12 s				21.1 (CH ₃)	H-21	
22	1.66 s				20.5 (CH ₃)	H-22	
					169.7 (C)		H-1/H-2/H-6 and
					169.7 (C)		H-20, 21, 22
					169.7 (C)		

*Assignments in the same vertical column may be reversed.

C-9. The long range coupling of the carbonyl signals at δ 169.4, 169.5 and 170.2 with the proton signals at δ 5.36 (H-2), 5.44 (H-1), 4.76 and 4.71 (H-11a,b) and the acetyl methyl signals at δ 2.06, 1.53 and 2.32 indicated the presence of acetyl groups on C-2, C-1 and C-11. The last carbonyl carbon at δ 175.9 was correlated with the proton signals at δ 5.60 (H-8), 0.87 (H-16), 0.99 (H-17) and 2.32 (H-18). Thus, this isobutyryl group had to be situated on C-8. It left only one oxygenated sp³-carbon atom (C-6), which must bear a hydroxyl group. Therefore, the structure was established as 4.

Compounds bearing these ester groups and having $H-1_{ax}$, $H-2_{eq}$, $H-6_{ax}$, $H-8_{eq}$ and $H-9_{eq}$ stereochemistry

have not been reported earlier. Some related sesquiterpenes [14-17] have been reported from the Celastraceae. However, compounds 1-4 are different from the formerly isolated compounds both in substitution pattern and stereochemistry.

EXPERIMENTAL

Celastrus augulatus Maxim. was collected in Kunming. A voucher specimen is deposited in the Herbarium of Kunming Institute of Botany, Academia Sinica.

General. 1D NMR: 400 MHz (¹H) and 125 MHz (¹³C); 2D NMR: 500 MHz (¹H) and 125 MHz (¹³C). MS: Finnigan MAT 90.

Table 3. ¹H and ¹³C NMR spectral data for compound 3 (in CDCl₃)

н	δ	J(Hz)	С	δ (DEPT)
1	5.52 m		1	70.7 (CH)*
2	5.52 m		2	68.1 (CH)*
3a	2.20 m		3	42.2 (CH ₂)
3b	1.94 m			
4			4	70.0 (C)
5			5	91.4 (C)
6	6.31 s		6	75.7 (CH)*
7	2.45 d	2.9	7	53.6 (CH)
8	5.43 d	2.9	8	76.1 (CH)*
9	5.71 s		9	70.3 (CH)*
10			10	54.4 (C)
11a	4.93 d	12.6	11	65.2 (CH ₂)
11b	4.77 d	12.6		
12	1.57 s		12	29.7 (CH ₃)
13			13	83.4 (C)
14	1.65 s		14	25.6 (CH ₃)
15	1.47 s		15	24.6 (CH ₃)
Fu				
2'	8.20 s		2'	148.9 (CH)
3'			3'	118.9 (C)
4′	6.83 d	1.7	4'	110.0 (CH)
5'	7.44 d	1.7	5'	143.9 (CH)
6'			6'	160.6 (C)
8′	8.00 s		8′	148.8(CH)
9′			9'	118.2 (C)
10'	6.72 d	1.6	10'	109.8 (CH)
11'	7.41 d	1.6	11'	143.9 (CH)
12'			12'	160.6 (C)
MeBu				
16	1.10 d	7.0	16	16.6 (CH ₃)
17	0.75 t	7.4	17	11.4 (CH ₃)
18	1.38 m		18	26.5 (CH ₃)
19	2.45 m		19	41.2 (CH)
20			20	176.5 (C)
Ac				
	2.11 s			21.4 (CH ₃)
	2.03 s			21.1 (CH ₃)
	1.64 s			20.6 (CH ₃)

Isolation of sesquiterpenes. The air-dried and powdered root-bark (2 kg) of C. angulatus, collected in Kunming, was extracted with Me₂CO at room temp. The Me₂CO exact (354 g) was extracted (\times 4) with CHCl₃. The CHCl₃ extract was partitioned against 2 1 petrol-MeOH-H₂O (30:20:1). The aq. layer was concd in vacuo to give a yellow residue (70 g). The residue was dissolved in CHCl₃, passed through a column of neutral alumina, then concd to give 50 g of residue. 10 g of this residue was chromatographed on a silica gel column eluting with petrol- Et_2O (1:1), Et_2O and EtOAc. A subfr. of the petrol- $Et_2O(1:1)$ eluant which was negative against Dragendorff's reagent, was subjected to HPLC (RP-18, MeOH-H₂O, 3:1, 1.0 ml min⁻¹, RI detector) and further purified by HPLC (diol, nhexane-EtOAc, 3:1, 1.8 ml min⁻¹, RI detector) to afford compounds 1 (12 mg), 2 (9 mg), 3 (4 mg) and 4 (7 mg).

1β, 2β, 6α-*Triacetoxy*-4α-hydroxy-*hydroxy*-8β,11*diisobutyryoxy*-9α-*furoyloxydihydro*-β-*agarofuran* (1). Amorphous; $[\alpha]_{D}$: -17° (c 0.45, CHCl₃); IR v cm⁻¹ (KBr): 3500 (br), 1730, 1380, 1230; EIMS (rel. int.): *m/z* 679 (1.8), 634 (5.4), 606 (10.2), 574 (6.2), 488 (7.9), 192 (49), 95 (100), 71 (34.6), 43 (74.6); ¹H and ¹³C NMR: Table 1.

1β, 2β, 6α-Triacetoxy-4α-hydroxy-8β,9α-difuroyloxy-11-isobutyryloxydihydro-β-agarofuran (2). Amorphous; $[\alpha]_{D}$: - 17° (c 0.38, CHCl₃); IR v cm⁻¹ (KBr): 3500 (br), 1730, 1380, 1230; EIMS m/z: 564, 546, 532, 461, 392, 305, 244, 192, 95; ¹H and ¹³C NMR: Table 2.

1β, 2β, 6α-*Triacetoxy*-4α-hydroxy-8β,9α-difuroyloxy-11-(3-methyl)isobutyryloxydihydro-β-agarofuran (3). Amorphous; $[α]_D$: 15° (c 0.18, CHCl₃); CIMS m/z (rel. int.): 733 $[M + 1]^+$ (15.7), 732 (13.3), 672 (96.7), 620 (100), 612 (83.1), 578 (43.4), 560 (29.9), 500 (27.1), 192 (67.4), 95 (85), 57 (30.3); ¹H and ¹³C NMR: Table 3.

1β, 2β, 11-Triacetoxy-4α,6α-dihydroxy-8α-isobutyryloxy-9β-benzoyloxydihydro-β-agarofuran (4). Amorphous; $[\alpha]_{\rm D}$: -9° (c 0.23, CHCl₃); IR v cm⁻¹ (KBr): 3500 (br), 1730, 1380, 1280, 1230, 710; EIMS m/z: 619, 574, 546, 244, 202, 164, 105, 71, 43; ¹H and ¹³C NMR: Table 4.

*Assignments in the same vertical column may be reversed.

H	δ	J(Hz)	¹ H–H COSY correlation with	С	δ (DEPT)	¹ H- ¹³ C COSY correlation with	Significant COLOC correlation with
1	5.63 d	3.4	H-2	1	67.5 (CH)	H-1	
2	5.44 q	3.4	H-1/H-3	2	75.0 (CH)	H-2	
3	2.01 m		H-2	3	41.3 (CH ₂)	H-3	
4				4	72.2 (C)		
5				5	91.6 (C)		
6	5.20 s			6	76.7 (CH)	H-6	
7	2.57 d	3.1	H-8	7	53.8 (CH)	H-7	
8	5.60 dd	9.8, 3.1	H-7/H-9	8	73.5 (CH)	H-8	
9	6.04 d	9.8	H-8	9	75.5 (CH)	H-9	
10				10	53.8 (C)		
11a	4.76 d	12.7	H-11b	11	61.5 (CH ₂)	H-11	

Table 4. ¹H and ¹³C NMR spectral data for compound 4 (in CDCl₃)

Continued overleaf

Table 4 Continue

н	δ	J(Hz)	¹ H–H COSY correlation with	С	δ (DEPT)	¹ H- ¹³ C COSY correlation with	Significant COLOC correlation with
11b	4.71 d	12.7	H-11a				
12	1.70 s			12	30.1 (CH ₃)	H-12	
13				13	84.6 (C)		
14	1.76 s			14	26.3 (CH ₃)	H-14	
15	1.59 s			15	24.3 (CH ₃)	H-15	
Bz							
1′				1′	129.5(C)		
2', 6'	7.83 bd	7.8	H-3', 4', 5'	2', 6'	129.5 (CH)	H-2', 6'	
4'	7.53 't'	7.2	H-2', 3' 5' 6'	4′	133.4 (CH)	H-4′	
3', 5'	7.39 't'	7.8	H-2', 4', 6'	3', 5'	128.7 (CH)	H-3', 5'	
7'				7′	165.8 (C)		H-9/H-2′, 6′
iBu							
16	0.87 d	7	H-18	16	18.5 (CH ₃)	H-16	
17	0.90 d	7	H-18	17	18.6 (CH ₃)	H-17	
18	2.32 m		H-16/H-17	18	34.1 (CH)	H-18	
19				19	175.9 (C)		H-8/H-18/H-16, 17
Ac							
20	2.32 s			20	21.5 (CH ₃)	H-20	
21				21	170.2 (C)		H-11a, b/H-20
22	2.06 s			21	21.0 (CH ₃)	H-22	
23				23	169.4 (C)		H-1/H-22
24	1.53 s			24	20.5 (CH ₃)	H-24	
25				25	169.5 (C)		H-2/H-24

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