

SHORT COMMUNICATION

Inhibitory Activities of Pancreatic Lipase and Phosphodiesterase from Korean Medicinal Plant Extracts

Yun Mi Lee,^{1†} Young Sook Kim,^{1†} Youngseop Lee,¹ Junghyun Kim,¹ Hang Sun,²
Joo Hwan Kim³ and Jin Sook Kim^{1*}

¹Diabetic Complications Research Center, Division of Traditional Korean Medicine (TKM) Integrated Research, Korea Institute of Oriental Medicine (KIOM), Daejeon 305-811, Republic of Korea

²Laboratory of Biodiversity and Biogeography, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming, Yunnan 650204, Peoples Republic of China

³Department of Life Science, Kyungwon University, Seongnam, Kyonggi-do 461–701, Republic of Korea

To find new pancreatic lipase (triacylglycerol acylhydrolase, EC 3.1.1.3) inhibitors from natural products, 61 medicinal plants from Korea were screened for their antilipase activity for prevention of obesity. Dried and powdered plants were extracted three times with EtOH and extracts were obtained by removal of the solvent *in vacuo*. Lipase activity was determined by measuring the hydrolysis of *p*-nitrophenyl butyrate to *p*-nitrophenol. Also, the inhibitory effect was measured on phosphodiesterase (PDE), another therapeutic target for obesity. Of the extracts tested, *Sorbus commixta* (stem, leaf) and *Viscum album* (whole plant) exhibited antilipase activity (with IC₅₀ values of 29.6 µg/mL and 33.3 µg/mL, respectively) and only anti-PDE activity (IC₅₀ values of 20.08 µg/mL and 35.15 µg/mL, respectively). Copyright © 2011 John Wiley & Sons, Ltd.

Keywords: pancreatic lipase; phosphodiesterase; Korean medicinal plants; *Sorbus commixta*; *Viscum album*.

INTRODUCTION

Obesity is characterized by excessive accumulation of fat and is a serious problem that is reaching epidemic proportions worldwide (James *et al.*, 2004). It is closely associated with lifestyle-related diseases, such as hypertension, hyperlipidemia, arteriosclerosis and diabetes mellitus (Campbell and Mathys, 2001). Triglycerides are the high-calorie energy sources, and suppression of triglyceride absorption is one possible way of preventing obesity and obesity-related diseases (Carek and Dickerson, 1999). Pancreatic lipase is a key enzyme for triglyceride absorption in the small intestine. It is secreted from the pancreas and hydrolyses triglycerides into glycerol and free fatty acids (Lowe, 1994). Orlistat inhibits the action of gastrointestinal lipase and thus reduces absorption of dietary fat. However, it has serious side effects, such as steatorrhea, stomach pain, irregular menstrual periods and headaches (Bray, 2009). Phosphodiesterase inhibitors (PDEIs) are a class of drugs that are widely used because of their various pharmacological properties; thus they may have potential as therapeutic targets for cardiovascular disease and obesity (Rahimi *et al.*, 2010; Sato *et al.*, 2006; Kim *et al.*, 2010). In this paper,

extracts obtained from 61 traditional Korean medicinal plants were screened in pursuit of new sources of pancreatic lipase inhibitors from relatively safe and effective natural products. *In vitro*, PDE activity was also measured.

MATERIALS AND METHODS

Plant materials. Sixty-one traditional Korean herbal medicines (Table 1) were collected from all areas of South Korea, from September 2004 to July 2007, and identified by Professor J. H. Kim, Division of Life Science, Kyungwon University, Seongnam, Korea. Voucher specimens have been deposited at the Herbarium of Diabetic Complications Research Center, Korea Institute of Oriental Medicine.

Preparation of plant extracts. Dried and powdered plant (0.2 g) was extracted three times with 4 mL of EtOH and extracts were obtained by removal of the solvent *in vacuo*. Concentrated samples were stored at –20 °C for further studies. Extracts were dissolved in DMSO at a concentration that in the total volume (3%) did not affect enzyme activity.

Chemicals. Lipase (Type II; from porcine pancreas), orlistat and *p*-nitrophenyl butyrate were purchased from Sigma Chemical Co. (St Louis, MO, USA). All reagents were of biochemical grade.

* Correspondence to: Dr Jin Sook Kim, Diabetic Complications Research Center, Division of Traditional Korean Medicine (TKM) Integrated Research, Korea Institute of Oriental Medicine (KIOM) 483 Expore, Yuseong-gu, Daejeon, 305–811, Republic of Korea.
E-mail: jskim@kiom.re.kr

†These authors contributed equally to this work.

Table 1. Lipase inhibitory activity of traditional Korean medicinal plants

Scientific name	Family name	Part used	Conc. ($\mu\text{g/mL}$)	Inhibition (%) ^a	IC ₅₀ ($\mu\text{g/mL}$)
<i>Abies koreana</i> E.H.Wilson	Pinaceae	Twig, leaf	65	47.87 \pm 3.10	85.41
			80	49.43 \pm 1.72	
			100	51.71 \pm 1.81	
<i>Acer tataricum</i> subsp. <i>ginnala</i> (Maxim.) Wesm.	Aceraceae	Fruit	20	38.80 \pm 1.32	44.44
			35	45.05 \pm 2.69	
			50	52.95 \pm 2.20	
<i>Actinidia polygama</i> (Siebold & Zucc.) Maxim.	Actinidiaceae	Leaf, stem	100	6.09 \pm 1.48	> 100
<i>Agrimonia pilosa</i> Ledeb.	Rosaceae	Whole plant	100	28.10 \pm 1.0	> 100
<i>Anemarrhena asphodeloides</i> Bunge	Haemodoraceae	Root, stem	100	11.78 \pm 4.80	> 100
<i>Angelica decursiva</i> (Miq.) Franch. & Sav.	Umbelliferae	Stem, leaf	100	1.91 \pm 3.43	> 100
<i>Artemisia capillaries</i> Thunb.	Compositae	Whole plant	100	34.29 \pm 2.97	> 100
<i>Aster ageratoides</i> Turcz.	Compositae	Whole plant	100	12.03 \pm 4.27	> 100
<i>Astilbe rubra</i> Hook.f. & Thomson	Saxifragaceae	Root	65	42.31 \pm 1.46	88.54
			80	45.25 \pm 2.08	
			100	54.83 \pm 0.17	
<i>Camellia japonica</i> subsp. <i>rusticana</i>	Theaceae	Stem, leaf	20	32.31 \pm 1.30	48.83
			35	41.28 \pm 3.03	
			50	50.82 \pm 6.28	
<i>Castanea crenata</i> var. <i>dulcis</i>	Fagaceae	Staminate flower	20	31.52 \pm 0.71	49.82
			35	40.55 \pm 1.02	
			50	50.26 \pm 1.22	
<i>Castanea crenata</i> var. <i>dulcis</i>	Fagaceae	Bark, twig	65	46.82 \pm 0.19	89.39
			80	49.59 \pm 1.99	
			100	50.94 \pm 1.54	
<i>Castanea crenata</i> var. <i>dulcis</i>	Fagaceae	Leaf	65	43.76 \pm 1.57	96.54
			80	47.35 \pm 0.35	
			100	50.53 \pm 1.92	
<i>Catalpa bignonioides</i> Walter	Bignoniaceae	Twig, leaf	100	29.52 \pm 1.09	> 100
<i>Celastrus orbiculatus</i> Thunb.	Celastraceae	Stem, leaf, fruit	100	18.53 \pm 2.01	> 100
<i>Chrysanthemum indicum</i> L.	Compositae	Stem, root	100	27.54 \pm 1.12	> 100
<i>Chrysanthemum indicum</i> L.	Compositae	Whole plant	100	27.32 \pm 3.65	> 100
<i>Cimicifuga heracleifolia</i> Kom.	Ranunculaceae	Whole plant	100	18.18 \pm 6.26	> 100
<i>Clematis terniflora</i> DC.	Ranunculaceae	Stem	100	3.01 \pm 5.45	> 100
<i>Cnidium japonicum</i> Miq.	Umbelliferae	Whole plant	100	8.06 \pm 3.72	> 100
<i>Crataegus pinnatifida</i> Bunge	Rosaceae	Fruit	100	1.42 \pm 4.55	> 100
<i>Crataegus pinnatifida</i> Bunge	Rosaceae	Stem, twig	100	42.50 \pm 1.19	> 100
<i>Cudrania tricuspidata</i> (Carr.) Bur.	Moraceae	Root	100	39.08 \pm 3.26	> 100
<i>Dictamnus dasycarpus</i> Turcz.	Rutaceae	Aerial part	100	2.36 \pm 3.53	> 100
<i>Dictamnus dasycarpus</i> Turcz.	Rutaceae	Root	100	1.54 \pm 3.50	> 100
<i>Eleutherococcus senticosus</i> Rupr. & Maxim.	Araliaceae	Stem	100	8.03 \pm 1.27	> 100
<i>Epimedium koreanum</i> Nakai	Berberidaceae	Leaf, stem	100	8.71 \pm 8.62	> 100
<i>Euonymus alatus</i> (Thunb.) Siebold	Celastraceae	Stem	100	44.67 \pm 3.53	> 100
<i>Eurya japonica</i> Thunb.	Theaceae	Twig, leaf	100	37.35 \pm 6.51	> 100
<i>Euodia daniellii</i> var. <i>hupehensis</i> (Dode) C.C.Huang	Rutaceae	Twig, leaf	100	32.76 \pm 5.87	> 100
<i>Forsythia Koreana</i> (rehd.) Nakai	Oleaceae	Stem, leaf	100	5.36 \pm 7.61	> 100
<i>Geum aleppicum</i> Jacq.	Rosaceae	Whole plant	100	6.40 \pm 1.88	> 100
<i>Glycyrrhiza pallidiflora</i> Maxim.	Leguminosae	Leaf, stem	100	39.94 \pm 5.30	> 100
<i>Hemiptelea davidii</i> (Hance) Planch.	Ulmaceae	Twig, stem, leaf	100	16.27 \pm 4.51	> 100
<i>Houttuynia cordata</i> Thunb.	Saururaceae	Whole plant	100	3.08 \pm 3.25	> 100
<i>Ilex cornuta</i> Lindl. & Paxton	Aquifoliaceae	Stem, leaf	100	6.08 \pm 2.49	> 100
<i>Lagerstroemia indica</i> L.	Lythraceae	Stem, leaf	100	22.04 \pm 6.74	> 100
<i>Litsea japonica</i> (Thunb.) Jussieu	Lauraceae	Twig, leaf	100	9.0 \pm 5.04	> 100
<i>Lycopus ramosissimus</i> var. <i>japonicus</i>	Labiatae	Whole plant	100	17.48 \pm 2.31	> 100
<i>Neillia uyekii</i> Nakai for. Uyekii	Rosaceae	Twig, stem, leaf	100	36.55 \pm 1.62	> 100
<i>Paeonia suffruticosa</i> Andr.	Ranunculaceae	Stem	100	18.60 \pm 3.60	> 100
<i>Patrinia scabiosaeifolia</i> Fisch. ex Link	Valerianaceae	Whole plant	100	17.34 \pm 8.12	> 100
<i>Petasites japonicas</i> (Siebold & Zucc.) Maxim.	Compositae	Stem, leaf	100	3.86 \pm 3.26	> 100
<i>Prunella vulgaris</i> L.	Labiatae	Leaf, stem	100	16.51 \pm 10.42	> 100
<i>Pueraria lobata</i> (Willd.) Ohwi	Leguminosae	Stem, leaf	100	9.05 \pm 3.15	> 100
<i>Rhododendron schlippenbachii</i> Maxim.	Ericaceae	Twig, stem, leaf	100	26.09 \pm 2.06	> 100

(Continues)

Table 1. (Continued)

Scientific name	Family name	Part used	Conc. (µg/mL)	Inhibition (%) ^a	IC ₅₀ (µg/mL)
<i>Rhus javanica</i> L.	Anacardiaceae	Twig, stem, leaf	100	6.75 ± 3.25	> 100
<i>Saururus chinensis</i> Baill.	Saururaceae	Leaf, stem	65	49.18 ± 1.85	81.05
			80	49.89 ± 2.24	
			100	51.18 ± 1.02	
<i>Schisandra chinensis</i> Baillon	Magnoliaceae	Stem	100	11.94 ± 1.02	> 100
<i>Sedum kamtschaticum</i> Fisch. & Mey.	Crassulaceae	Root	65	43.97 ± 2.01	87.14
			80	48.70 ± 2.40	
			100	52.99 ± 0.92	
<i>Sophora flavescens</i> Aiton	Leguminosae	Root	100	48.63 ± 3.59	> 100
<i>Sorbus commixta</i> Hedl.	Rosaceae	Stem, leaf	20	46.45 ± 1.75	29.62
			35	53.1 ± 2.41	
			50	55.23 ± 0.84	
<i>Stephanandra incisa</i> (Thunb.) Zabel	Rosaceae	Twig, stem, leaf	100	27.79 ± 2.0	> 100
<i>Styrax obassia</i> Siebold & Zucc.	Styracaceae	Twig, stem	100	11.46 ± 1.63	> 100
<i>Teucrium viscidum</i> Blume	Labiatae	Whole plant	100	15.92 ± 6.25	> 100
<i>Viola takesimana</i> Nakai	Violaceae	Whole plant	100	12.59 ± 10.56	> 100
<i>Viscum album</i> L.	Loranthaceae	Whole plant	10	31.06 ± 1.79	33.32
			20	41.71 ± 1.57	
			35	50.55 ± 1.32	
<i>Vitex rotundifolia</i> L.f.	Verbenaceae	Stem	100	10.78 ± 1.11	> 100
<i>Vitex rotundifolia</i> L.f.	Verbenaceae	Leaf	100	7.52 ± 3.57	> 100
<i>Weigela subsessilis</i> L.H. Bailey	Caprifoliaceae	Twig, stem, leaf	100	0.74 ± 5.37	> 100
<i>Xanthium strumarium</i> L.	Compositae	Whole plant	100	8.92 ± 7.93	> 100
Orlistat (positive control)			0.025	18.03 ± 2.98	0.076
			0.05	35.64 ± 1.25	(0.154 µM)
			0.1	63.68 ± 1.41	

^aResults are the mean ± SD ($n = 4$).

Pancreatic lipase inhibitory activity. Determination of pancreatic lipase activity was modified from that of Kim *et al.* (2005). Briefly, an enzyme buffer was prepared by the addition of 6 µL a solution of porcine pancreatic lipase (Sigma) [2.5 mg/mL in 10 mM MOPS (morpholinopropanesulphonic acid) and 1 mM EDTA, pH 6.8] to 169 µL of Tris buffer (100 mM Tris-HCl and 5 mM CaCl₂, pH 7.0). Then, either 20 µL of the compound at the test concentration, or orlistat (Sigma), was mixed with 175 µL of the enzyme buffer and incubated for 15 min at 37 °C with 5 µL of the substrate solution [10 mM p-NPB (*p*-nitrophenyl butyrate) in dimethyl formamide]. The enzymatic reactions were allowed to proceed for 30 min at 37 °C. Lipase activity was determined by measuring the hydrolysis of p-NPB into p-nitrophenol. Increase in light absorption at 405 nm was measured using a plate reader (Bio-Tek, Synergy HT, USA). Inhibition of lipase activity was expressed as the percentage decrease in OD when porcine pancreatic lipase was incubated with the test compounds. Lipase inhibition (%) was calculated according the following formula:

$$\text{Inhibition (\%)} = 100 - \left(\frac{B - b}{A - a} \times 100 \right)$$

where A is the activity without inhibitor, a the negative control without inhibitor, B the activity with inhibitor and b is the negative control with inhibitor. The results were expressed as an average ($n = 4$).

Phosphodiesterase (PDE) activity assay. The PDE activity was assayed using the PDE-Glo™ phosphodiesterase activity kit (Promega Corp., WI, USA) according to the manufacturer's instructions. The results were expressed as an average ($n = 3$) (Kim *et al.*, 2010).

Statistical analysis. Unpaired Student's *t*-tests were performed and data are expressed as the mean ± SEM of multiple experiments (Prism Software, Graph Pad, San Diego, CA, USA). A difference in the mean values of $p < 0.05$ was considered as significant.

RESULTS AND DISCUSSION

Pancreatic lipase inhibition is one of the most widely studied mechanisms used to determine the potential efficacy of natural products as antiobesity drugs (Kim and Kang, 2005; Kim *et al.*, 2007; Rahimi *et al.*, 2010). The results of inhibitory activity of 61 Korean medicinal plants on pancreatic lipase have been summarized in Table 1. Lipase inhibition is expressed as the percentage (%) and IC₅₀ value (the concentration required to inhibit a lipase activity by 50%). Of the extracts tested, 11 showed an IC₅₀ of less than 100 µg/mL, and of the 11 extracts, five exhibited an IC₅₀ of less than 50 µg/mL. *Sorbus commixta* (stem, leaf) showed the strongest inhibitory activity on lipase with an IC₅₀ value of 29.62 µg/mL. Those of *Viscum album* (whole plant), *Acer ginnala* (fruit), *Camellia japonica* (stem, leaf) and

Castanea crenata (staminate flower) exhibited the inhibitory effect with an IC_{50} value of 30–50 $\mu\text{g/mL}$.

Previous study shows that 200 $\mu\text{g/mL}$ of *Eriochloa villosa*, *Orixa japonica* (81.3%) and *Setaria italica* (80.3%) extracts exhibit antilipase activity greater than 80% *in vitro* (Sharma *et al.*, 2005). In this study, *Sorbus commixta* (stem, leaf) extracts (IC_{50} = 29.62 $\mu\text{g/mL}$) showed a stronger inhibitory effect than those of these extracts on antilipase activity. However, it was not more effective than orlistat (IC_{50} = 0.076 $\mu\text{g/mL}$), a positive control. Orlistat, one of the clinically approved drugs for obesity treatment, has been shown to act by inhibition of pancreatic lipase. Although it is one of the best-selling drugs worldwide, it has serious side effects (Kim *et al.*, 2007). In this study, *Sorbus commixta* and *Viscum album* showed inhibitory effects on PDE activity in dose-dependent manners (Fig. 1). *Sorbus commixta* shows strong inhibitory activity against protein tyrosine phosphatase 1B (PTP1B), an attractive target for the development of new drugs for type 2 diabetes (Ahmed *et al.*, 2008; Na *et al.*, 2009). Furthermore, it has also antioxidative activity (Bae *et al.*, 2007). *Viscum album* (mistletoe), which is distributed in Korea and other East-Asian countries, has long been recognized as a therapeutic herb. It has been traditionally used as a sedative, analgesic, antispasmodic, cardiostonic and anticancer agent (Lyu and Park, 2006). *Acer ginnala* shows the inhibitory effect on rat lens aldose reductase activity *in vitro* (Kim and Oh, 1999). Saponins isolated from *Camellia japonica* leaves have been shown to have antifungal activity and antifeedant activity and tannins from *Camellia japonica* leaves have an inhibitory effect on HIV-1 protease (Onodera *et al.*, 2006). Flower extracts of *Castanea crenata* are reported to have antimicrobial activity (Lee *et al.*, 1999).

To date, *Sorbus commixta* (twig, leaf) extracts have not been reported to have lipase and PDE inhibitory activities. Thus, it is worthwhile to further investigate these extracts for their potential pharmacological effect in obesity *in vivo* and attempts should be made to purify their active components to be used as safer and cheaper therapeutic agents in future.

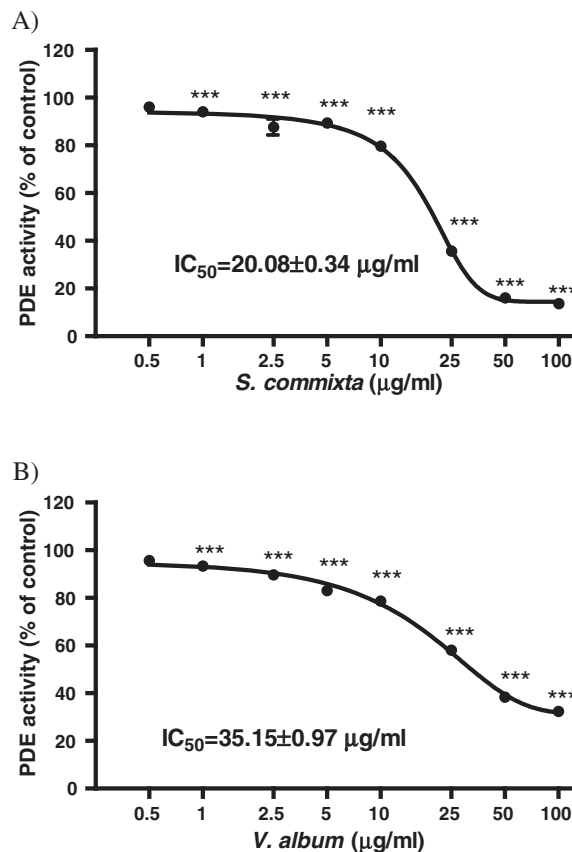


Figure 1. Inhibitory effects of *S. commixta* and *V. album* on PDE activity. (A) *S. commixta*. Data are expressed as the mean \pm SEM ($n = 3$). *** $p < 0.001$ vs. untreated group. (B) *V. album*. Data are expressed as the mean \pm SEM ($n = 3$). *** $p < 0.001$ vs. untreated group.

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Conflict of Interest

The authors report no conflict of interest.

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